Lyme Borreliosis, Lyme Arthritis, Erythema Migrans with Polyarthritis

Last Updated: January 2021



The Center for Food Security & Public Health



INSTITUTE FOR INTERNATIONAL COOPERATION IN ANIMAL BIOLOGICS

IOWA STATE UNIVERSITY College of Veterinary Medicine



World Organisation for Animal Health Founded as OIE



#### Importance

Lyme disease is a tickborne illness caused by members of the *Borrelia burgdorferi* sensu lato complex. These organisms are maintained in wildlife, but most reported illnesses are in humans, with occasional cases in domestic animals, particularly dogs. Lyme disease was first recognized in the 1970s, when a cluster of juvenile arthritis cases was investigated in the U.S.; however, the causative organisms are now known to be relatively widespread and have been found in Europe as well as parts of Asia, Canada and South America. Clinical cases in humans are readily cured with antibiotics during the initial stage of the illness, when an unusual rash often aids disease recognition. However, people whose infections remain untreated during this early stage sometimes develop other syndromes, such as arthritis or neurological signs, which can be more difficult to diagnose. Lyme disease in animals is still incompletely understood.

#### Etiology

Lyme disease is caused by members of the *Borrelia burgdorferi* sensu lato complex, in the family Spirochaetaceae. A very similar disease in Brazil is sometimes called Lyme-like disease or Baggio-Yoshinari syndrome. There are about 20 recognized genospecies (genomic groups) in the *B. burgdorferi* s.l. complex, of which approximately half have been reported in humans. The organisms found most often in clinical cases are *B. burgdorferi* sensu stricto (*B. burgdorferi* s.s.), *B. garinii, B. bavariensis* (formerly *B. garinii* OspA serotype 4), *B. afzelii*, and to a lesser extent, *B. mayonii* and *B. spielmanii*. *B. bissetii, B. valaisiana* and *B. lusitaniae* have been reported in rare cases, while *B. americana* and *B. andersonii* were each found in the blood of a single person in North America. Little is known about the specific organisms in domestic animals, but *B. burgdorferi* s.s. is presumed to be the agent in North America, and *B. afzelii* and *B. garinii* might also play a role in other parts of the world. Other *B. burgdorferi* s.l. complex members (e.g., *B. lusitaniae*) have only been found in healthy animals, to date.

## **Species Affected**

#### **Reservoir hosts**

Mammals, birds and reptiles can serve as reservoir hosts for the members of the *B. burgdorferi* s.l. complex. Most of the clinically important organisms are maintained in rodents, insectivores (e.g., shrews) or other small mammals. The white-footed mouse (*Peromyscus leucopus*) is the primary reservoir host for *B. burgdorferi* s.s. in the eastern U.S., but various chipmunks, squirrels, mice, voles and shrews may be important in some other parts of North America. *P. leucopus* has also been suggested as a maintenance hosts for *B. mayonii*. Yellow-necked mice (*Apodemus flavicollis*), striped field mice (*A. agrarius*), wood mice (*A. sylvaticus*), and the voles *Clethrionomys glareolus* and *Microtus agrestis*, are among the reservoir hosts for *B. burgdorferi* s.l. complex members (e.g., *B. afzelii, B. bavariensis, B. lusitaniae*) in Europe and Asia. Other possible hosts include European hedgehogs (*Erinaceus europaeus*), and lagomorphs, particularly the brown hare (*Lepus europaeus*) and the varying hare (*L. timidus*). Capybaras (*Hydrochoerus hydrochaeris*) are thought to be maintenance hosts in Brazil. Cervids are important in providing blood meals to ticks, but they do not play a role in amplifying or maintaining the members of the *B. burgdorferi* s.l. complex.

Birds are thought to maintain *B. garinii*, which causes some cases of Lyme disease in people, as well as *B. valaisiana* and *B. turdi. B. garinii* occurs in seabirds as well as terrestrial species such as pheasants (*Phasianus colchicus*) and some passerines. Certain lizards appear to be reservoir hosts for *B. lusitaniae*. However, the complement proteins in the blood of many lizards are highly lytic for some members of the *B. burgdorferi* s.l., complex, and may reduce their prevalence in ticks.

#### **Incidental hosts**

Evidence for *B. burgdorferi* sensu lato infections has been found in many domestic animals and some free-living wildlife, including large mammals such as

wild carnivores. Most surveys have examined mammals, but some organisms, including *B. lusitaniae* and *B. burgdorferi* s.s., have been detected occasionally in birds. Clinical cases have mainly been reported in domestic animals, particularly dogs and horses, with a few putative cases in cats, ruminants and a captive chimpanzee (*Pan troglodytes*). Dogs, horses, cats, rabbits, mice, hamsters, gerbils and guinea pigs have been infected experimentally.

Some incidental hosts, such as dogs, may occasionally transmit *B. burgdorferi* s.l. to ticks, but they are not thought to be significant in maintaining these organisms.

#### **Zoonotic potential**

Organisms known to affect people include *B.* burgdorferi s.s., *B. garinii, B. bavariensis, B. afzelii, B.* mayonii, *B. spielmanii, B. bissetii, B. valaisiana* and *B.* lusitaniae. *B. americana* and *B. andersonii* were each detected in the blood of a single person in North America. Other genospecies might also be zoonotic but appear to be rare in humans.

### **Geographic Distribution**

As of 2020, Lyme disease is known to occur in parts of North and South America, Europe and Asia. There are reports of a Lyme-like disease in Australia; however, researchers have been unable to find evidence for any *B. burgdorferi* s.l. members in his location, suggesting that the Australian cases might either be another disease or acquired elsewhere. Most members of the *B. burgdorferi* s.l complex are limited to temperate areas where their vectors, *Ixodes* ticks, can survive. *B. garinii*, which can be spread by seabirds and their cold-resistant ticks, can be found even in polar regions, where it has been detected in seabird colonies. Migratory birds sometimes distribute infected ticks beyond their usual locations, but the organisms do not necessarily persist long-term.

Different genospecies can cause Lyme disease in different locations. In North America, this disease is usually caused by B. burgdorferi s.s., though B. mayonii has been responsible for some cases in the Upper Midwest. The major endemic foci are in the northeastern, mid-Atlantic and northcentral U.S., and along the Pacific coast, though cases can occur elsewhere, including parts of Canada. B. garinii has been found in seabird colonies in a few North American locations including Alaska and Gull Island, Newfoundland, but there have been no human cases attributed to this organism in the Western Hemisphere. In South America, Lyme disease or a similar illness has been documented in Brazil, where it is thought to be caused by *B. burgdorferi* s.s. Members of the B. burgdorferi s.l. complex have also been reported from Uruguay, Argentina and Chile, with the latter two reports describing the species as *B. chilensis*.

Organisms found in Europe include *B. garinii*, *B. bavariensis*, *B. afzelii*, *B. burgdorferi* s.s., *B. spielmanii*, *B. valaisiana* and *B. lusitaniae*. Some of these agents, such as *B. lusitaniae*, seem to occur in limited locations. *B. garinii* and *B. afzelii* are thought to be the most important

genospecies in Asia, but *B. burgdorferi* s.s. has been detected in a few countries (e.g., Taiwan), and recent research suggests that *B. bavariensis* is widespread. *B. lusitaniae* and either *B. valaisiana* or a related species have also been documented in some parts of Asia.

#### Transmission

Members of the *B. burgdorferi* s.l. complex are usually transmitted by ticks in the genus *Ixodes;* however, other tick genera, including *Amblyomma* and *Rhipicephalus*, were proposed as important vectors in Brazil. *Ixodes scapularis* is the major vector for *B. burgdorferi* s.s. in eastern North America and the Midwest, while *I. pacificus* transmits this organism on the Pacific coast. *B. mayonii* is also thought to be transmitted by *I. scapularis*. *I. ricinus* and *I. persulcatus* are the primary vectors for *B. burgdorferi* s.l. complex members in Europe and Asia, though other *Ixodes* ticks may be important locally. The cold-tolerant seabird tick *I. uriae* has been found to carry other organisms such as *B. bavariensis*, *B. lusitaniae* and *B. burgdorferi* s.s.

Successful transmission of *B. burgdorferi* s.l. to a new host requires that an infected tick remain attached for at least a day or two, though certain circumstances (e.g., bites from partially fed ticks or bites from multiple infected ticks) may shorten this period. The minimum attachment time seems to vary with the organism. *B. burgdorferi* s.s. and *B. mayonii* are not usually acquired with attachment times of less than 48 hours, while *B. afzelii* could infect some mice by 48 hours but not before 24 hours. Transstadial transmission has been demonstrated in some tick species, but transovarial transmission seems to be insignificant.

B. burgdorferi s.l. does not spread directly between infected vertebrates to any significant extent, although there are a few research reports of organisms (or their nucleic acids) occasionally being detected in body fluids such as urine, the breast milk of humans with erythema migrans, and colostrum or milk from cattle. One report described transmission from an experimentally infected dog, which excreted spirochetes in its urine, to a control animal. However, another study found that susceptible dogs cohoused with infected dogs for a year did not seroconvert. Possible transplacental and sexual transmission have been proposed in humans, but definitive evidence to support either route is currently lacking. There are no known cases of Lyme disease resulting from a blood transfusion, though transfusion-acquired disease is theoretically possible and it has been demonstrated in experimentally infected mice.

#### **Disinfection**

*Borrelia burgdorferi* sensu lato does not survive well outside the body, but if necessary it can be inactivated by 1% sodium hypochlorite or 70% ethanol. It is also sensitive to heat and ultraviolet light.

## **Infections in Animals**

#### **Incubation Period**

There is limited information about the incubation period in animals, but some experimentally infected dogs developed arthritis in 2-5 months.

## **Clinical Signs**

Most infections in animals seem to be asymptomatic. The range of clinical syndromes is not completely understood, but they are assumed to be similar to those in humans, with most reports describing arthritis, neurological signs, ocular disease and/or cardiac signs. A possible link with nephritis has also been proposed in dogs. Erythema migrans, a common initial sign in humans, has rarely been documented in animals.

#### Dogs

Arthritis is the most commonly described syndrome in dogs and can affect one to a few joints, especially the carpal joints. The joints may or may not be swollen. The resulting lameness may be intermittent or shift from leg to leg. Nonspecific signs including fever, anorexia, lethargy or lymphadenitis, particularly of the prescapular or popliteal nodes, may be seen concurrently in some animals. Arthritis is the only syndrome that has been reproduced in experimentally infected dogs. In one experiment, it was most likely to occur in younger puppies, transient (1-2 days) in older puppies, and absent in adults, which seroconverted without clinical signs. Arthritis was self-limited in this report, though younger animals sometimes had a few recurrent episodes. However, subsequent studies have suggested that severe arthritis is possible in some animals.

Cardiac dysfunction (myocarditis or conduction abnormalities with bradycardia) and neurological signs (e.g., facial paralysis, seizures, aggression) have been attributed to *B. burgdorferi* in a small number of naturally infected dogs. Some of these animals had a history of arthritis. Based on immunostaining for *Borrelia* antigens, Lyme disease was also proposed to cause a syndrome of acute or chronic nephritis with protein-losing nephropathy and immune-mediated glomerulonephritis. Most affected dogs had signs of renal failure, which often progressed rapidly. The association of this condition with Lyme disease is considered unconfirmed, as it has not been reproduced experimentally, and immunostaining could result from cross-reactivity.

Dogs referred to specialists for Lyme disease that is unresponsive to antibiotics often have other illnesses.

#### Cats

Very little is known about the consequences of infection in cats. While significant numbers of cats are seropositive in some surveys, reports of naturally occurring disease are rare, and definitive attribution of the clinical signs to *B. burgdorferi* is difficult. Most suspected cases have involved lameness or intermittent lameness, with or without fever, that improved after treatment with antibiotics effective for *Borrelia*. There have also been cases submitted for Lyme disease serology with neurological signs (e.g., hind leg paresis, ataxia, trembling). Two cases of bradydysrhythmia in asymptomatic cats were recently attributed to this disease. One cat had a structurally normal heart, with a previous history of an erythema migrans-like lesion on its abdomen. The arhythmia resolved after treatment with doxycycline. The other cat had both dilated cardiomyopathy and arhythmia, and eventually developed congestive heart failure.

Conflicting results have been seen in experimentally infected cats. In one study, the animals remained asymptomatic; in another, they developed fever, lethargy and arthritis with intermittent stiffness.

#### **Horses**

The prevalence of Lyme disease in horses remains controversial. Currently, the syndromes with the most evidence for *B. burgdorferi* in a causative role are uveitis, which is often severe and sometimes progresses to blindness; neurological signs (e.g., meningitis, dysphagia, laryngeal dysfunction, facial paresis, neck and back stiffness, progressive ataxia, encephalitis); and cutaneous pseudolymphoma. Cutaneous pseudolymphomas are papular to nodular lesions that occur at the site of a tick bite. They have been documented in experimentally infected as well as naturally infected horses.

Arthritis, cardiac arhythmias and myocarditis have also been attributed to Lyme disease in horses. A few reports of arthritis supported the diagnosis with evidence for *B. burgdorferi* infection, and documented lymphoplasmacytic synovitis with marked fluid distention of the joints or tendon sheaths. However, most horses with presumed but unconfirmed cases have not had joint swelling. There are also diverse unconfirmed and often anecdotal syndromes attributed to Lyme disease, including rare reports of conditions such as laminitis, hepatitis or nephritis, as well as nonspecific signs (e.g., low grade fever) or multisystemic syndromes.

#### **Other species**

Clinical signs that have been attributed to Lyme disease in cattle include fever, lameness/ stiffness with or without joint swelling, uveitis, skin lesions (erythema, warmth, swelling and hypersensitivity of the skin on the ventral udder, developing into dark sloughing scabs) and nonspecific signs such as chronic weight loss and abortions. In one study, calves inoculated with the three European genospecies remained asymptomatic. One presumptive case was reported in a captive chimpanzee that had recurrent episodes of illness with nonspecific signs and shifting leg lameness. This animal had elevated antibody titers to *B. burgdorferi* at the time of diagnosis, and responded to doxycycline, with subsequent decrease in titers. Erythema migrans skin lesions, polyarthritis and carditis have been reported in experimentally infected rabbits.

#### **Post Mortem Lesions**

#### Solution of the second second

The predominant lesions attributed to Lyme disease are nonerosive arthritis, uveitis, myocarditis and lesions in the central nervous system (CNS). Neurological involvement has been described as neutrophilic or lymphoplasmacytic, histiocytic, perivascular to diffuse inflammation that affects the meninges, ganglia, and cranial and spinal nerve roots, with variable necrosis, fibrosis and neuroparenchymal invasion. Some horses with neuroborreliosis may have visible lesions in the meninges, including opacification, yellowish discoloration and hyperemia, with injected vessels and plaques of edema. Nephritis and various lesions associated with uremia were attributed to Lyme disease in some dogs.

## **Diagnostic Tests**

Antemortem diagnosis of Lyme disease is typically based on a combination of the clinical signs, epidemiology (i.e., a history of exposure to ticks in an endemic area), elimination of other diseases, laboratory data and response to antibiotics. It is most often presumptive, as laboratory confirmation is usually limited to serology. *B. burgdorferi* s.l. is rarely found in blood; however, organisms, nucleic acids or antigens may be detected occasionally in affected tissues, particularly the joints (especially synovium), meninges, cerebrospinal fluid (CSF), ocular fluid or cardiac muscle. Histopathology may be helpful in some instances, particularly at necropsy.

Antibody titers can often be detected within 3-8 weeks of exposure in dogs and horses, depending on the assay. Commonly used serological tests include various ELISAs, line immunoassays (LIA), fluorescent bead-based multiplex assays and immunoblotting. Most tests were developed for dogs; however, some are also used in other species. Indirect immunofluorescence assays (IFA) were employed in the past, but some sources no longer recommend them due to their low specificity. Screening tests such as ELISAs can be confirmed by immunoblotting, but a single C6-peptide-based assay is sometimes used in place of this two-tier testing in dogs. Serological diagnosis is complicated by the long incubation period, prevalence of asymptomatic infections in healthy animals, cross-reactions with some other organisms, and persistence of titers for months or years. Some assays (e.g., immunoblotting, the C6-peptide-based ELISA) can distinguish vaccinated dogs from those that are infected.

PCR is used most often to detect the organism directly, but there are occasional research reports of antigen detection by immunohistochemistry, often postmortem. Whenever possible, PCR or antigen testing should be interpreted in conjunction with histopathology, as nucleic acids or antigens may be present without being the cause of the disease, and cross-reactivity may be an issue with antigen tests. Isolation of *B. burgdorferi* s.l. is uncommonly attempted and often unsuccessful. The organisms in this complex are fastidious and microaerophilic, and must be cultured on enriched bacteriologic media such as Barbour-Stoenner-Kelly (BSK) or modified Kelly-Pettenkofer (MKP) media. Recovered organisms (Gram negative, motile spirochetes) can be visualized with dark-field or phase-contrast microscopy, immunofluorescent microscopy, silver stains, Giemsa, or acridine orange stain. They can be confirmed as members of the *B. burgdorferi* s.l. complex with specific monoclonal antibodies or by PCR.

The genospecies is rarely identified in clinical cases. If necessary, this can be done with molecular methods such as large restriction fragment pattern (LRFP) analysis plus plasmid profiling, PCR-based typing methods that target single genes, PCR-based restriction fragment length polymorphism (RFLP) analysis, MLST of house-keeping genes or whole genome sequencing. Distinguishing some closely-related organisms, such as *B. garinii* and *B. bavariensis*, is impractical outside a specialized facility that can do multilocus genotyping.

#### Treatment

Lyme disease is usually treated with antibiotics, often beta-lactam drugs (e.g., amoxicillin) or tetracyclines (e.g., doxycycline). Steroids are employed in immune-mediated polyarthropathy in dogs. Oral antibiotic treatment of horses with neurological signs is often unsuccessful, and some sources suggest that parenteral treatment focusing on adequate drug concentrations in the target organ might be more effective.

#### Control

#### **Disease reporting**

Veterinarians who encounter or suspect an infection with *B. burgdorferi* sensu lato should follow their national and/or local guidelines for disease reporting. Although Lyme disease in humans is nationally reportable in the U.S., this is not the case for animals. State authorities should be consulted for any reporting requirements.

#### Prevention

Acaricides (e.g., pour-on preparations) and tick repellents can help prevent tick bites. Animals should also be checked frequently (at least daily) for ticks, which should be removed as soon as possible. Avoidance of habitats where ticks are more common, such as the woods, reduces exposure. Environmental modifications such as excluding deer (which provide blood meals to sustain ticks) and removing debris or other features that may shelter ticks and rodents, may be helpful, but most of these measures have not been extensively evaluated. The efficacy of some measures, such as attempts to reduce tick populations on wildlife with acaricides, is inconclusive and the cost-benefit unclear.

Several different types of Lyme disease vaccines are currently available for dogs, but their use is controversial, with varying estimates of efficacy. Canine vaccines have sometimes been used off-label in horses, though responses seem to be inconsistent.

### **Morbidity and Mortality**

In endemic areas, antibodies to *B. burgdorferi* sensu lato have been detected in 5-50% or more of healthy dogs, cats, horses and some ruminants, as well as wildlife. Some studies have found that dogs are more likely to be seropositive than cats. Most infections in animals are thought to be subclinical, with epidemiological studies suggesting that approximately 5% or less of all infected dogs in endemic areas develop Lyme disease. Clinical cases seem to be uncommon in horses and have been rarely reported in other species, but lack of awareness and diagnostic uncertainties might play a role. The consequences of infection vary with the syndrome. Neurological cases in horses typically have a poor prognosis, and vision may not improve in horses with uveitis.

#### **Infections in Humans**

#### **Incubation Period**

Erythema migrans rashes can appear from a few days to a month after exposure, with most cases becoming apparent within a week or two. The incubation period for borrelial lymphocytoma ranges from 10 days to several weeks or more. If Lyme disease is not treated in the early stages, additional signs may be observed weeks to years later.

### **Clinical Signs**

Most, though not all, people with early Lyme disease develop a distinctive skin lesion called erythema migrans, which may or may not be accompanied by a flu-like, sometimes febrile, illness. Erythema migrans typically originates as a macule or papule, which widens into a red or bluish-red rash with distinct and often intensely colored (e.g., bright red), but not raised, borders. The rash, which is usually painless, expands over days or weeks. It often develops a "bullseye" appearance with central clearing. This central area may occasionally have vesicular or necrotic lesions. Secondary erythema migrans lesions can occasionally appear on other parts of the body. A small number of people with Lyme disease develop a painless bluish-red or reddish-purple nodule or plaque on the skin, which called a borrelial lymphocytoma. Borrelial lymphocytomas often occur on the ear, nipple or scrotum; are most common in children; can accompany or follow erythema migrans; and tend to be caused by B. afzelii or B. garinii/ B. bavariensis. They are rare in North America, where these genospecies are absent. Atypical rashes have been reported occasionally in Lyme disease patients, including a diffuse macular rash in some people infected with B. mayonii.

Without treatment, the initial symptoms of Lyme disease usually last a few weeks (untreated borrelial lymphocytoma may persist for months) and may recur, but eventually disappear. Some of these patients can later develop arthritis, neurological signs, or other syndromes. Arthritis, which appears to be particularly common in North America, usually appears as intermittent pain affecting one or a few joints, with or without swelling. Large, weightbearing joints such as the knee are affected most often. The arthritis may be recurrent or long lasting, persisting for months or possibly years. It usually responds well to antibiotics, but a small proportion of nonresponsive cases are thought to be caused by an autoimmune reaction.

Neurological syndromes, which seem to be especially common with B. garinii/ B. bavariensis, can include meningitis, facial palsy, radiculitis (pain or discomfort associated with nerve inflammation) or lymphocytic meningoradiculitis with or without paresis, as well as less common or rare conditions such as myelitis, cranial neuritis, chorea or encephalitis. Acute syndromes such as facial nerve palsy are usually self-limiting, but some signs can persist for months, and sequelae are possible. Occasional cardiac complications tend to be transient atrioventricular blocks of varying severity, but other arhythmias, myocarditis, endocarditis or pericarditis are also possible. Rarely, there may be ocular signs including conjunctivitis during the early stages, and uveitis, keratitis, optic nerve involvement (e.g., optic neuritis, papilledema secondary to elevated intracranial pressure, neuroretinitis) and other conditions late.

Late stage complications occur in some untreated patients months or years after the tick bite, and can include acrodermatitis chronica atrophicans, rare incidents of neurological involvement (e.g., chronic progressive meningoencephalitis, multifocal cerebral vasculitis) and chronic arthritis. Acrodermatitis chronica atrophicans is a skin condition often associated with *B. afzelii*, though it can be caused by other genospecies. It usually affects older adults, is uncommon in children, and occurs most often on the limbs. The typical presentation is an area of red or bluish-red cutaneous discoloration, often accompanied by doughy swelling, which is followed by slow, progressive skin atrophy that eventually results in an area of parchmentlike skin with prominent veins. The skin lesions may be accompanied by pain, pruritus, hyperesthesia or paresthesia. Acrodermatitis chronica atrophicans can be treated successfully in the initial stages, but skin atrophy and neuropathy are often irreversible. Likewise, other late stage Lyme disease complications may result in residual neurological deficits or arthritis, especially if treatment is delayed until the signs are advanced.

Co-infection with another tick-borne disease, especially human granulocytic anaplasmosis or babesiosis, can alter the clinical signs and response to treatment.

# Chronic Lyme disease and post-Lyme disease syndrome

"Chronic Lyme disease" has been used to describe several different groups of patients, including untreated or inadequately treated patients with persistent objective signs of Lyme disease (e.g., acrodermatitis chronica atrophicans) and people with post-Lyme disease syndrome. Patients with post-Lyme disease syndrome complain of persistent or relapsing nonspecific symptoms such as fatigue, headache, sleep disorders, myalgia, arthralgia, paresthesias, or difficulty with memory or concentration after they have been treated for Lyme disease. Possible causes include slow resolution after treatment, autoimmune reactions, postinfective fatigue syndrome (which also occurs after a variety of other infections), and concurrent diseases or conditions. There is no evidence that *B. burgdorferi* persists in this syndrome. The number of patients reporting symptoms decreases over time.

Some practitioners have also applied the term "chronic Lyme disease" to a group of people with vague, nonspecific illnesses and positive serology but no response to antibiotics or other evidence that *B. burgdorferi* s.l. is causing their symptoms.

#### **Diagnostic Tests**

As in animals, human Lyme disease is usually diagnosed by the clinical signs, together with a history of exposure to ticks and supportive evidence from laboratory tests. The diagnosis is typically based on symptoms alone in people with erythema migrans, as this rash usually develops before antibodies can be found. While PCR can often detect nucleic acids in a skin biopsy at this stage, testing is normally done only for atypical lesions. It should be noted that, while erythema migrans lesions are usually caused by Lyme disease, it is possible for other diseases to cause a similar rash.

The diagnosis in other syndromes is most often supported by serology. Patients can be screened with various enzyme immunoassays (e.g., ELISAs, enzyme-linked fluorescent assays, chemiluminescence immunoassays), and positive or indeterminate samples are confirmed by immunoblotting. A second ELISA is now considered acceptable for confirmation in some countries. Serology must be interpreted with caution and in conjunction with the clinical signs, as antibodies to *B*. *burgdorferi* are very common in the human population, including healthy people. The CSF/serum antibody index is helpful in the diagnosis of neurological disease, together with signs of inflammation in the CSF.

PCR is sometimes useful for conditions such as acrodermatitis chronica atrophicans, or on synovial tissue or fluid in arthritis. Although PCR may detect nucleic acids in the CSF of patients with neurological conditions, it is not very sensitive in this syndrome. Culture is rarely attempted.

## Treatment

Early treatment with antibiotics reduces the duration of erythema migrans and prevents later complications such as arthritis. Anti-inflammatory drugs are employed in cases of Lyme arthritis thought to be caused by an autoimmune reaction. Treatment failures are rare with the currently recommended regimens; however, patients treated later in the course of the disease may recover more slowly, and acrodermatitis chronica atrophicans can require several weeks of treatment. Studies have found no benefit to prolonged antibiotic treatment in post-Lyme disease syndrome.

### **Prevention**

Lyme disease prevention consists of avoiding tickinfested locations and/or preventing tick bites with protective footwear, clothing (e.g., long pants tucked into the boots, and shirts tucked into the trousers) and tick repellents. Ticks may be more visible on light-colored clothing. People who enter tick habitats should check frequently for ticks and remove them as soon as possible. Gloves are recommended during tick removal. One study found that showering soon after exposure to a tick-infested area was also helpful. Environmental modifications to decrease tick exposure (see Prevention section for animals, above) are more common in North America, where people are often exposed around their homes, than in Europe, where exposure tends to be occupational or recreational.

Postexposure antimicrobial prophylaxis (a single dose of doxycycline within 72 hours) is sometimes given in parts of North America where the risk of infection is high and the tick has been attached for at least 36 hours. Prophylactic treatment is not usually employed in Europe, but some studies suggest there may be a benefit. There is currently no vaccine for humans; a vaccine licensed in the U.S. was withdrawn from the market by the manufacturer in 2002.

### **Morbidity and Mortality**

In the Northern Hemisphere, most cases of Lyme disease are seen from late spring to summer, when ticks are most active and more people are outdoors. People can become reinfected, usually with a different strain. The reported incidence of Lyme disease is approximately 7-10 cases per 100,000 population in the U.S., where exposure around the home is common in some rural and suburban areas. In Europe, infections tend to result from occupational exposure to ticks or recreation in tick habitats, and the estimated incidence in various countries ranges from < 1 to > 100 cases per 100,000 population. The precise number of Lyme disease cases in both North America and Europe is unclear, as some authors feel that significant numbers of cases might be missed, while others suggest that the reliance on serology leads to overdiagnosis and confusion with other conditions. Many patients with chronic conditions seen at Lyme disease referral centers do not have this disease.

How many infected people become ill is uncertain, with estimates ranging from 5% to 50%. In Europe, 3-6% of people have been estimated to seroconvert after a tick bite, with overt clinical signs developing in 0.3% to 1.4%. In one study, 61% of Lyme disease patients with erythema migrans developed neurological, articular or cardiac symptoms. Fatalities appear to be very rare, with only one human death attributed to Lyme disease,

#### Internet Resources

Centers for Disease Control and Prevention (CDC)

European Centre for Disease Prevention and Control. Borreliosis

Public Health Agency of Canada. Pathogen Safety Data Sheets

The Merck Manual

The Merck Veterinary Manual

### **Acknowledgements**

This factsheet was written by Anna Rovid Spickler, DVM, PhD, Veterinary Specialist from the Center for Food Security and Public Health. The U.S. Department of Agriculture Animal and Plant Health Inspection Service (USDA APHIS) provided funding for this factsheet through a series of cooperative agreements related to the development of resources for initial accreditation training.

The following format can be used to cite this factsheet. Spickler, Anna Rovid. 2020. *Lyme Disease*. Retrieved from http://www.cfsph.iastate.edu/DiseaseInfo/factsheets.php.

#### References

Aberer E. Lyme borreliosis--an update. J Dtsch Dermatol Ges. 2007;5(5):406-14.

- Acha PN, Szyfres B (Pan American Health Organization [PAHO]). Zoonoses and communicable diseases common to man and animals. Volume 1. Bacterioses and mycoses. 3rd ed. Washington DC: PAHO; 2003. Scientific and Technical Publication No. 580. Lyme disease; p. 179-84.
- Adaszek Ł, Gatellet M, Mazurek Ł, Dębiak P, Skrzypczak M, Winiarczyk S. Myocarditis secondary to *Borrelia* infection in a dog: a case report. Ann Parasitol. 2020;66(2):255-7.

Aguero-Rosenfeld ME, Wang G, Schwartz I, Wormser GP. Diagnosis of lyme borreliosis. Clin Microbiol Rev. 2005;18(3):484-509.

- Alho AM, Pita J, Amaro A, Amaro F, Schnyder M, Grimm F, Custódio AC, Cardoso L, Deplazes P, de Carvalho LM. Seroprevalence of vector-borne pathogens and molecular detection of *Borrelia afzelii* in military dogs from Portugal. Parasit Vectors. 2016;9(1):225.
- Bartol J. Is Lyme disease overdiagnosed in horses?Equine Vet J. 2013;45(5):529-30.
- Basile RC, Yoshinari NH, Mantovani E, Bonoldi VN, Macoris DD, Queiroz-Neto A. Brazilian borreliosis with special emphasis on humans and horses. Braz J Microbiol. 2017;48(1):167-72.
- Beckmann S, Freund R, Pehl H, Rodgers A, Venegas T. Rodent species as possible reservoirs of *Borrelia burgdorferi* in a prairie ecosystem. Ticks Tick Borne Dis. 2019;10(5):1162-7.

Bratton RL, Whiteside JW, Hovan MJ, Engle RL, Edwards FD. Diagnosis and treatment of Lyme disease. Mayo Clin Proc. 2008;83(5):566-71.

CAB International. *Ixodes ricinus*. Available at: <u>https://www.cabi.org/isc/datasheet/78352</u>. Accessed 22 Dec 2020.

Centers for Disease Control and Prevention [CDC]. Lyme disease [online]. CDC; 2003 Dec. Available at http://www.cdc.gov/ ncidod/dvbid/lyme/index.htm.\*. Accessed 9 July 2004.

- Chalada MJ, Stenos J, Bradbury RS. Is there a Lyme-like disease in Australia? Summary of the findings to date. One Health. 2016;2:42-54.
- Chou J, Wünschmann A, Hodzic E, Borjesson DL. Detection of *Borrelia burgdorferi* DNA in tissues from dogs with presumptive Lyme borreliosis. J Am Vet Med Assoc. 2006;229(8):1260-5.
- Collignon PJ, Lum GD, Robson JM. Does Lyme disease exist in Australia? Med J Aust. 2016 Nov 7;205(9):413-7.

Comstedt P, Jakobsson T, Bergström S. Global ecology and epidemiology of *Borrelia garinii* spirochetes. Infect Ecol Epidemiol 2011;1: doi: 10.3402/iee.v1i0.9545.

Cutler SJ, Ruzic-Sabljic E, Potkonjak A. Emerging borreliae -Expanding beyond Lyme borreliosis. Mol Cell Probes. 2017;31:22-7.

Dambach DM, Smith CA, Lewis RM, Van Winkle TJ. Morphologic, immunohistochemical, and ultrastructural characterization of a distinctive renal lesion in dogs putatively associated with *Borrelia burgdorferi* infection: 49 cases (1987-1992). Vet Pathol. 1997;34:85-96.

Detmer SE, Bouljihad M, Hayden DW, Schefers JM, Armien A, Wünschmann A. Fatal pyogranulomatous myocarditis in 10 Boxer puppies. J Vet Diagn Invest. 2016;28(2):144-9.

Divers JT. Chang YF, McDonough PL (College of Veterinary Medicine, Cornell University). Equine Lyme disease: A review of experimental disease production, treatment efficacy, and vaccine prevention. In: 49<sup>th</sup> Annual Convention of the American Association of Equine Practitioners. 2003. New Orleans, Louisiana. Available at: <u>https://www.ivis.org/library/aaep/aaep-annual-conventionnew-orleans-2003/equine-lyme-disease-a-review-of-</u>

experimental. Accessed 9 July 2004. Divers TJ, Gardner RB, Madigan JE, Witonsky SG, Bertone JJ, Swinebroad EL, Schutzer SE, Johnson AL. *Borrelia* 

- Swinebroad EL, Schutzer SE, Johnson AL. *Borrelia burgdorferi* infection and Lyme disease in North American horses: a consensus statement. J Vet Intern Med. 2018;32(2):617-32.
- Dolan MC, Breuner NE, Hojgaard A, Boegler KA, Hoxmeier JC, Replogle AJ, Eisen L. Transmission of the Lyme disease spirochete *Borrelia mayonii* in relation to duration of attachment by nymphal *Ixodes scapularis* (Acari: Ixodidae). J Med Entomol. 2017;54(5):1360-4.

Eldin C, Raffetin A, Bouiller K, Hansmann Y, Roblot F, Raoult D, Parola P. Review of European and American guidelines for the diagnosis of Lyme borreliosis. Med Mal Infect. 2019;49(2):121-32.

Fischhoff IR, Keesing F, Pendleton J, DePietro D, Teator M, Duerr STK, Mowry S, Pfister A, LaDeau SL, Ostfeld RS. Assessing effectiveness of recommended residential yard management measures against ticks. J Med Entomol. 2019;56(5):1420-27.

Foley DM, Gayek RJ, Skare JT, Wagar EA, Champion CI, Blanco DR, Lovett MA, Miller JN. Rabbit model of Lyme borreliosis: erythema migrans, infection-derived immunity, and identification of *Borrelia burgdorferi* proteins associated with virulence and protective immunity. J Clin Invest. 1995;96(2):965-75.

Fritz CL, Kjemtrup AM. Lyme borreliosis. J Am Vet Med Assoc. 2003;223:1261-70.

Gabitzsch ES, Piesman J, Dolan MC, Sykes CM, Zeidner NS. Transfer of *Borrelia burgdorferi* s.s. infection via blood transfusion in a murine model. J Parasitol. 2006;92(4):869-70.

Gall Y, Pfister K. Survey on the subject of equine Lyme borreliosis. Int J Med Microbiol. 2006;296 Suppl 40:274-9.

Garcia-Monco JC, Benach JL. Lyme Neuroborreliosis: Clinical outcomes, controversy, pathogenesis, and polymicrobial infections. Ann Neurol. 2019;85(1):21-31.

Gerber B, Haug K, Eichenberger S, Reusch CE, Wittenbrink MM. Follow-up of Bernese Mountain dogs and other dogs with serologically diagnosed *Borrelia burgdorferi* infection: what happens to seropositive animals? BMC Vet Res. 2009;5:18.

Gern L. Life cycle of *Borrelia burgdorferi* sensu lato and transmission to humans. Curr Probl Dermatol. 2009;37:18-30.

Gibson MD, Young CR, Omran MT, Edwards J, Palma K, Russell L, Rawlings J. *Borrelia burgdorferi* infection of cats. J Am Vet Med Assoc. 1993;202:1786.

Giery ST, Ostfeld RS. The role of lizards in the ecology of Lyme disease in two endemic zones of the northeastern United States. J Parasitol. 2007;93(3):511-7.

Ginsberg HS, Buckley PA, Balmforth MG, Zhioua E, Mitra S, Buckley FG. Reservoir competence of native North American birds for the lyme disease spirochete, *Borrelia burgdorfieri*. J Med Entomol. 2005;42(3):445-9.

Glatz M, Resinger A, Semmelweis K, Ambros-Rudolph CM, Müllegger RR. Clinical spectrum of skin manifestations of Lyme borreliosis in 204 children in Austria. Acta Derm Venereol. 2015;95(5):565-71.

Gocko X, Lenormand C, Lemogne C, Bouiller K, Gehanno JF, et al. Lyme borreliosis and other tick-borne diseases. Guidelines from the French scientific societies. Med Mal Infect. 2019;49(5):296-317.

Goddard J. Not all erythema migrans lesions are Lyme disease. Am J Med. 2017;130(2):231-3.

Golovchenko M, Vancová M, Clark K, Oliver JH Jr, Grubhoffer L, Rudenko N. A divergent spirochete strain isolated from a resident of the southeastern United States was identified by multilocus sequence typing as *Borrelia bissettii*. Parasit Vectors. 2016;9:68.

Gonçalves DD, Carreira T, Nunes M, Benitez A, Lopes-Mori FM, Vidotto O, de Freitas JC, Vieira ML. First record of *Borrelia burgdorferi* B31 strain in *Dermacentor nitens* ticks in the northern region of Parana (Brazil). Braz J Microbiol. 2014;44(3):883-7.

Green RT. Canine Lyme borreliosis. In: Kirk RW, editor. Current veterinary therapy X. Philadelphia: WB Saunders; 1989. p. 1086-7.

Halperin JJ. Lyme neuroborreliosis. Curr Opin Infect Dis. 2019;32(3):259-64.

Harms MG, Hofhuis A, Sprong H, Bennema SC, Ferreira JA, Fonville M, Docters van Leeuwen A, Assendelft WJJ, Van Weert HCPM, Van Pelt W, Van den Wijngaard CC. A single dose of doxycycline after an *Ixodes ricinus* tick bite to prevent Lyme borreliosis: an open-label randomized controlled trial. J Infect. 2020 Jun 18 [Epub ahead of print].

Hildenbrand P, Craven DE, Jones R, Nemeskal P. Lyme neuroborreliosis: manifestations of a rapidly emerging zoonosis. AJNR Am J Neuroradiol. 2009;30(6):1079-87. Humair PF. Birds and *Borrelia*. Int J Med Microbiol. 2002;291 Suppl 33:70-4.

Inokuma H, Maetani S, Fujitsuka J, Takano A, Sato K, Fukui T, Masuzawa T, Kawabata H. Astasia and pyrexia related to *Borrelia garinii* infection in two dogs in Hokkaido, Japan. J Vet Med Sci. 2013;75:975-8.

Irwin PJ, Robertson ID, Westman ME, Perkins M, Straubinger RK. Searching for Lyme borreliosis in Australia: results of a canine sentinel study. Parasit Vectors. 2017;10(1):114.

Jaulhac B, Saunier A, Caumes E, Bouiller K, Gehanno JF, et al. Lyme borreliosis and other tick-borne diseases. Guidelines from the French scientific societies (II). Biological diagnosis, treatment, persistent symptoms after documented or suspected Lyme borreliosis. Med Mal Infect. 2019;49(5):335-46.

John TM, Taege AJ. Appropriate laboratory testing in Lyme disease. Cleve Clin J Med. 2019;86(11):751-59.

Johnson AL, Divers TJ, Chang YF. Validation of an in-clinic enzyme-linked immunosorbent assay kit for diagnosis of *Borrelia burgdorferi* infection in horses. J Vet Diagn Invest. 2008;20(3):321-4.

Johnson RC. Leptospira, Borrelia (including Lyme disease) and Spirillum [monograph online]. In: Baron S, editor. Medical Microbiology. 4th ed. New York: Churchill Livingstone; 1996. Available at: http://www.gsbs.utmb.edu/microbook/.\* Accessed 9 July 2004.

Johnstone LK, Engiles JB, Aceto H, Buechner-Maxwell V, Divers T, Gardner R, Levine R, Scherrer N, Tewari D, Tomlinson J, Johnson AL. Retrospective evaluation of horses diagnosed with neuroborreliosis on postmortem examination: 16 cases (2004-2015). J Vet Intern Med. 2016;30(4):1305-12.

Jordan BE, Onks KR, Hamilton SW, Hayslette SE, Wright SM. Detection of *Borrelia burgdorferi* and *Borrelia lonestari* in birds in Tennessee. J Med Entomol. 2009;46(1):131-8.

Jungnick S, Margos G, Rieger M, Dzaferovic E, Bent SJ, Overzier E, Silaghi C, Walder G, Wex F, Koloczek J, Sing A, Fingerle V. *Borrelia burgdorferi* sensu stricto and *Borrelia afzelii*: Population structure and differential pathogenicity. Int J Med Microbiol. 2015;305(7):673-81.

Kawabata H, Takano A, Kadosaka T, Fujita H, Nitta Y, et al. Multilocus sequence typing and DNA similarity analysis implicates that a *Borrelia valaisiana*-related sp. isolated in Japan is distinguishable from European *B. valaisiana*. J Vet Med Sci. 2013;75(9):1201-7.

Kobayashi T, Higgins Y, Samuels R, Moaven A, Sanyal A, Yenokyan G, Lantos PM, Melia MT, Auwaerter PG. Misdiagnosis of Lyme disease with unnecessary antimicrobial treatment characterizes patients referred to an academic infectious diseases clinic. Open Forum Infect Dis. 2019;6(7)::ofz299.

Kortela E, Kanerva M, Kurkela S, Oksi J, Järvinen A. Suspicion of Lyme borreliosis in patients referred to an infectious diseases clinic: what did the patients really have? Clin Microbiol Infect. 2020 Sep 23. [Epub ahead of print].

Kullberg BJ, Vrijmoeth HD, van de Schoor F, Hovius JW. Lyme borreliosis: diagnosis and management. BMJ. 2020;369:m1041.

LaFleur RL, Callister SM, Dant JC, Jobe DA, Lovrich SD, Warner TF, Wasmoen TL, Schell RF. One-year duration of immunity induced by vaccination with a canine Lyme disease bacterin. Clin Vaccine Immunol. 2010;17(5):870-4.

Lane RS, Mun J, Eisen RJ, Eisen L. Western gray squirrel (Rodentia: Sciuridae): a primary reservoir host of *Borrelia burgdorferi* in Californian oak woodlands? J Med Entomol. 2005;42(3):388-96.

Lantos PM, Branda JA, Boggan JC, Chudgar SM, Wilson EA, Ruffin F, Fowler V, Auwaerter PG, Nigrovic LE. Poor positive predictive value of Lyme disease serologic testing in an area of low disease incidence. Clin Infect Dis. 2015;61(9):1374-80.

Larsson C, Comstedt P, Olsen B, Bergström S. First record of Lyme disease *Borrelia* in the Arctic.Vector Borne Zoonotic Dis. 2007;7(3):453-6.

Lenormand C, Jaulhac B, Debarbieux S, Dupin N, Granel-Brocard F, Adamski H, Barthel C, Cribier B, Lipsker D. Expanding the clinicopathological spectrum of late cutaneous Lyme borreliosis (acrodermatitis chronica atrophicans [ACA]): A prospective study of 20 culture- and/or polymerase chain reaction (PCR)-documented cases. J Am Acad Dermatol. 2016;74(4):685-92.

Leschnik M. Canine borreliosis: are we facing the facts? Vet J. 2014;199(2):197-8.

Levy S. Developing an integrated approach for diagnosing, managing tick-borne diseases. Thorough testing aids in determining co-infections in dogs. DVM Newsmagazine [serial online]. 2002 June 1: 14-18. Available at: http://www.dvmnewsmagazine.com/dvm/article/articleDetail.js p?id=32680.\* Accessed 11 July 2004.

Lischer CJ, Leutenegger CM, Braun U, Lutz H. Diagnosis of Lyme disease in two cows by the detection of *Borrelia burgdorferi* DNA. Vet Rec. 2000;146(17):497-9.

Little SE, Heise SR, Blagburn BL, Callister SM, Mead PS. Lyme borreliosis in dogs and humans in the USA. Trends Parasitol. 2010;26(4):213-8.

Littman MP. Canine borreliosis. Vet Clin North Am Small Anim Pract. 2003;33(4):827-62.

Littman MP, Goldstein RE, Labato MA, Lappin MR, Moore GE. ACVIM small animal consensus statement on Lyme disease in dogs: diagnosis, treatment, and prevention. J Vet Intern Med. 2006;20(2):422-34.

Littman MP, Gerber B, Goldstein RE, Labato MA, Lappin MR, Moore GE. ACVIM consensus update on Lyme borreliosis in dogs and cats. J Vet Intern Med. 2018;32(3):887-903.

Lohr B, Fingerle V, Norris DE, Hunfeld KP. Laboratory diagnosis of Lyme borreliosis: Current state of the art and future perspectives. Crit Rev Clin Lab Sci. 2018;55(4):219-45.

Magnarelli LA, Bushmich SL, IJdo JW, Fikrig E. Seroprevalence of antibodies against *Borrelia burgdorferi* and *Anaplasma phagocytophilum* in cats. Am J Vet Res. 2005;66(11):1895-9.

Maraspin V, Mrvič T, Ružić-Sabljić E, Jurčić V, Strle F. Acrodermatitis chronica atrophicans in children: Report on two cases and review of the literature. Ticks Tick Borne Dis. 2019;10(1):180-5.

Margos G, Becker NS, Fingerle V, Sing A, Ramos JA, Lopes de Carvalho I, Norte AC. Core genome phylogenetic analysis of the avian associated *Borrelia turdi* indicates a close relationship to *Borrelia garinii*. Mol Phylogenet Evol. 2019;131:93-8.

Margos G, Castillo-Ramirez S, Cutler S, Dessau RB, Eikeland R, et al. Rejection of the name *Borreliella* and all proposed species comb. nov. placed therein. Int J Syst Evol Microbiol. 2020;70(5):3577-81. Margos G, Chu CY, Takano A, Jiang BG, Liu W, Kurtenbach K, Masuzawa T, Fingerle V, Cao WC, Kawabata H. *Borrelia yangtzensis* sp. nov., a rodent-associated species in Asia, is related to *Borrelia valaisiana*. Int J Syst Evol Microbiol. 2015;65(11):3836-40.

Margos G, Wilske B, Sing A, Hizo-Teufel C, Cao WC, Chu C, Scholz H, Straubinger RK, Fingerle V. *Borrelia bavariensis* sp. nov. is widely distributed in Europe and Asia. Int J Syst Evol Microbiol. 2013;63(Pt 11):4284-8.

Marques A. Chronic Lyme disease: a review. Infect Dis Clin North Am. 2008;22(2):341-60.

Marques AR. Lyme disease: a review. Curr Allergy Asthma Rep. 2010;10(1):13-20.

Masuzawa T. Terrestrial distribution of the Lyme borreliosis agent Borrelia burgdorferi sensu lato in East Asia. Jpn J Infect Dis. 2004;57(6):229-35.

Mead P, Petersen J, Hinckley A. Updated CDC Recommendation for serologic diagnosis of Lyme disease. MMWR Morb Mortal Wkly Rep. 2019;68(32):703.

Morgenstern K,Baljer G, Norris DE, Kraiczy P, Hanssen-Hübner C, Hunfeld K-P. *In vitro* susceptibility of *Borrelia spielmanii* to antimicrobial agents commonly used for treatment of Lyme disease. Antimicrob Agents Chemother. 2009;53(3):1281-4.

Morshed MG, Scott JD, Fernando K, Geddes G, McNabb A, Mak S, Durden LA. Distribution and characterization of *Borrelia burgdorferi* isolates from *Ixodes scapularis* and presence in mammalian hosts in Ontario, Canada. J Med Entomol. 2006;43(4):762-73.

Munro HJ, Ogden NH, Lindsay LR, Robertson GJ, Whitney H, Lang AS. Evidence for *Borrelia bavariensis* infections of *Ixodes uriae* within seabird colonies of the North Atlantic Ocean. Appl Environ Microbiol. 2017;83(20). pii: e01087-17.

Nau R, Christen HJ, Eiffert H. Lyme disease--current state of knowledge. Dtsch Arztebl Int. 2009;106(5):72-81.

Norte AC, Lobato DN, Braga EM, Antonini Y, Lacorte G, Gonçalves M, Lopes de Carvalho I, Gern L, Núncio MS, Ramos JA. Do ticks and *Borrelia burgdorferi* s.l. constitute a burden to birds? Parasitol Res. 2013;112(5):1903-12.

Norte AC, Ramos JA, Gern L, Núncio MS, Lopes de Carvalho I. Birds as reservoirs for *Borrelia burgdorferi* s.l. in Western Europe: circulation of *B. turdi* and other genospecies in birdtick cycles in Portugal. Environ Microbiol. 2013;15(2):386-97.

Pantchev N, Vrhovec MG, Pluta S, Straubinger RK. Seropositivity of *Borrelia burgdorferi* in a cohort of symptomatic cats from Europe based on a C6-peptide assay with discussion of implications in disease aetiology. Berl Munch Tierarztl Wochenschr. 2016;129(7-8):333-9.

Parise CM, Breuner NE, Hojgaard A, Osikowicz LM, Replogle AJ, Eisen RJ, Eisen L. Experimental demonstration of reservoir competence of the white-footed mouse, *Peromyscus leucopus* (Rodentia: Cricetidae), for the Lyme disease spirochete, *Borrelia mayonii* (Spirochaetales: Spirochaetaceae). J Med Entomol. 2020;57(3):927-32.

Pecoraro HL, Felippe MJB, Miller AD, Divers TJ, Simpson KW, Guyer KM, Duhamel GE. Neuroborreliosis in a horse with common variable immunodeficiency. J Vet Diagn Invest. 2019;31(2):241-5.

Pichon B, Astrada-Pena A, Kahl O, Mannelli A, Gray JS. Detection of animal reservoirs of tick-borne zoonoses in Europe. Int J Med Microbiol. 2006;296(S1): 129-30.

Piesman J. Strategies for reducing the risk of Lyme borreliosis in North America. Int J Med Microbiol. 2006;296 Suppl 40: 17-22.

Piesman J, Gern L. Lyme borreliosis in Europe and North America. Parasitology. 2004;129 Suppl:S191-220.

Pospisilova T, Urbanova V, Hes O, Kopacek P, Hajdusek O, Sima R. Tracking of *Borrelia afzelii* transmission from infected *Ixodes ricinus* nymphs to mice. Infect Immun. 2019;87. pii: e00896-18.

Priest HL, Irby NL, Schlafer DH, Divers TJ, Wagner B, Glaser AL, Chang YF, Smith MC. Diagnosis of *Borrelia*-associated uveitis in two horses. Vet Ophthalmol. 2012;15(6):398-405.

Pritt BS, Mead PS, Johnson DKH, Neitzel DF, Respicio-Kingry LB, et al. Identification of a novel pathogenic *Borrelia* species causing Lyme borreliosis with unusually high spirochaetaemia: a descriptive study. Lancet Infect Dis. 2016;16(5):556-4.

Pritt BS, Respicio-Kingry LB, Sloan LM, Schriefer ME, Replogle AJ, et al. *Borrelia mayonii* sp. nov., a member of the *Borrelia burgdorferi* sensu lato complex, detected in patients and ticks in the upper midwestern United States. Int J Syst Evol Microbiol. 2016;66(11):4878-80.

Public Health Agency of Canada (PHAC). Pathogen Safety Data Sheets: Infectious Substances – *Borrelia burgdorferi*. Office of Laboratory Security, PHAC; 1999. Available at: <u>https://www.canada.ca/en/public-health/services/laboratorybiosafety-biosecurity/pathogen-safety-data-sheets-riskassessment/borrelia-burgdorferi-material-safety-data-sheets-<u>msds.html</u>. Accessed 9 July 2004.</u>

Richardson M, Khouja C, Sutcliffe K. Interventions to prevent Lyme disease in humans: A systematic review. Prev Med Rep. 2018 Nov 13;13:16-22.

Richter D, Matuschka FR. Perpetuation of the Lyme disease spirochete *Borrelia lusitaniae* by lizards. Appl Environ Microbiol. 2006;72(7):4627-32.

Rudenko N, Golovchenko M, Grubhoffer L, Oliver JH Jr. *Borrelia* carolinensis sp. nov., a new (14th) member of the *Borrelia burgdorferi* sensu lato complex from the southeastern region of the United States. J Clin Microbiol. 2009;47(1):134-41.

Rudenko N, Golovchenko M, Lin T, Gao L, Grubhoffer L, Oliver JH Jr. Delineation of a new species of the *Borrelia burgdorferi* sensu lato complex, *Borrelia american*a sp. nov. J Clin Microbiol. 2009;47(12):3875-80.

Saito K, Ito T, Asashima N, Ohno M, Nagai R, Fujita H, Koizumi N, Takano A, Watanabe H, Kawabata H. *Borrelia valaisiana* infection in a Japanese man associated with traveling to foreign countries. Am J Trop Med Hyg. 2007;77(6):1124-27.

Scheffold N, Herkommer B, Kandolf R, May AE. Lyme carditisdiagnosis, treatment and prognosis. Dtsch Arztebl Int. 2015;112(12):202-8.

Scherrer NM, Knickelbein KE, Engiles JB, Johnstone LK, Tewari D, Johnson AL. Ocular disease in horses with confirmed ocular or central nervous system *Borrelia* infection: Case series and review of literature. Vet Ophthalmol. 2020 Aug 30. [Epub ahead of print]

Schoen RT. Lyme disease: diagnosis and treatment. Curr Opin Rheumatol. 2020;32(3):247-54.

Schuijt TJ, Hovius JW, van der Poll T, van Dam AP, Fikrig E. Lyme borreliosis vaccination: the facts, the challenge and the future. Trends Parasitol. 2011 Jan;27(1):40-7. Shaw SE, Birtles RJ, Day MJ. Arthropod-transmitted infectious diseases of cats. J Feline Med Surg. 2001;3(4):193-209.

Skotarczak B. Canine borreliosis – epidemiology and diagnostics. Ann Agric Environ Med. 2002;9:137-40.

Skotarczak B. Why are there several species of *Borrelia* burgdorferi sensu lato detected in dogs and humans? Infect Genet Evol. 2014;23:182-8.

Smith RP Jr, Muzaffar SB, Lavers J, Lacombe EH, Cahill BK, Lubelczyk CB, Kinsler A, Mathers AJ, Rand PW. *Borrelia* garinii in seabird ticks (*Ixodes uriae*), Atlantic Coast, North America. Emerg Infect Dis. 2006;12(12):1909-12.

Socoloski SNG, de Castro BG, Cordeiro MD, da Fonseca AH, Cepeda MB, Nicolino RR, Lopes LB. Epidemiological investigation of *Borrelia burgdorferi* in horses in the municipality of Sinop-MT, Brazil. Trop Anim Health Prod. 2018;50(4):831-6.

Speck S, Reiner B, Wittenbrink MM. Isolation of *Borrelia afzelii* from a dog. Vet Rec. 2001;149(1):19-20.

Speck S, Reiner B, Streich WJ, Reusch C, Wittenbrink MM. Canine borreliosis: a laboratory diagnostic trial. Vet Microbiol. 2007;120(1-2):132-41.

Stanek G, Fingerle V, Hunfeld KP, Jaulhac B, Kaiser R, Krause A, Kristoferitsch W, O'Connell S, Ornstein K, Strle F, Gray J. Lyme borreliosis: Clinical case definitions for diagnosis and management in Europe.Clin Microbiol Infect. 2011;17(1): 69-79.

Stanek G, Reiter M. The expanding Lyme *Borrelia* complex clinical significance of genomic species? Clin Microbiol Infect 2011;17:487-93.

Stanek G, Strle F. Lyme borreliosis-from tick bite to diagnosis and treatment. FEMS Microbiol Rev. 2018;42(3):233-58.

Straubinger RK. Lyme disease (Lyme borreliosis) in dogs. In: Line S, Moses MA, editors. The Merck veterinary manual. Kenilworth, NJ: Merck and Co; 2018. Available at: <u>https://www.merckvetmanual.com/dog-owners/disorders-affecting-multiple-body-systems-of-dogs/lyme-disease-lyme-borreliosis-in-dogs</u>. Accessed 19 Dec 2020.

Straubinger RK. Overview of Lyme borreliosis (Lyme disease).In: Line S, Moses MA, editors. The Merck veterinary manual. Kenilworth, NJ: Merck and Co; 2015. Available at: <u>https://www.merckvetmanual.com/generalized-</u> <u>conditions/lyme-borreliosis/overview-of-lyme-borreliosis</u>. Accessed 19 Dec 2020.

Stefanciková A, Adaszek Ł, Pet'ko B, Winiarczyk S, Dudinák V. Serological evidence of *Borrelia burgdorferi* sensu lato in horses and cattle from Poland and diagnostic problems of Lyme borreliosis. Ann Agric Environ Med. 2008;15(1):37-43.

Stefancíková A, Stěpánová G, Derdáková M, Pet'ko B, Kysel'ová J, Cigánek J, Strojný L, Cisláková L, Trávnicek M. Serological evidence for *Borrelia burgdorferi* infection associated with clinical signs in dairy cattle in Slovakia. Vet Res Commun. 2002;26(8):601-11.

Straubinger RK. Lyme borreliosis in dogs. In: Carmichael L, editor. Recent advances in canine infectious diseases [monograph online]. Ithaca NY: International Veterinary Information Service [IVIS]; 2001 Available at: http://www.ivis.org/advances/Infect\_Dis\_Carmichael/toc.asp. \* Accessed 9 July 2004.

Straubinger RK, Summers BA, Chang YF, Appel MJ. Persistence of *Borrelia burgdorferi* in experimentally infected dogs after antibiotic treatment. J Clin Microbiol. 1997;35(1):111-6.

Stromdahl EY, Nadolny RM, Gibbons JA, Auckland LD, Vince MA, Elkins CE, Murphy MP, Hickling GJ, Eshoo MW, Carolan HE, Crowder CD, Pilgard MA, Hamer SA. Borrelia burgdorferi not confirmed in human-biting Amblyomma americanum ticks from the southeastern United States. J Clin Microbiol. 2015;53(5):1697-704.

Susta L, Uhl EW, Grosenbaugh DA, Krimer PM. Synovial lesions in experimental canine Lyme borreliosis. Vet Pathol. 2012;49(3):453-61.

Sutton D, Spry C. Ottawa (ON): Canadian Agency for Drugs and Technologies in Health; 2019 May. CADTH Rapid Response Reports. One dose of doxycycline for the prevention of Lyme disease: a review of clinical effectiveness and guidelines. t]. Available at: <u>https://www.ncbi.nlm.nih.gov/ books/n/rc1121/pdf/</u>. Accessed 15 Dec 2020.

Talhari S, de Souza Santos MN, Talhari C, de Lima Ferreira LC, Silva RM Jr, Zelger B, Massone C, Ribeiro-Rodrigues R. Borrelia burgdorferi "sensu lato" in Brazil: Occurrence confirmed by immunohistochemistry and focus floating microscopy.Acta Trop. 2010;115(3):200-4.

Taragel'ová V, Koci J, Hanincová K, Kurtenbach K, Derdáková M, Ogden NH, Literák I, Kocianová E, Labuda M. Blackbirds and song thrushes constitute a key reservoir of *Borrelia* garinii, the causative agent of borreliosis in Central Europe. Appl Environ Microbiol. 2008;74(4):1289-93.

Tarageľová VR, Mahríková L, Selyemová D, Václav R, Derdáková M. Natural foci of *Borrelia lusitaniae* in a mountain region of Central Europe. Ticks Tick Borne Dis. 2016;7(2):350-6.

Tørnqvist-Johnsen C, Dickson SA, Rolph K, Palermo V, Hodgkiss-Geere H, Gilmore P, Gunn-Moore DA. First report of Lyme borreliosis leading to cardiac bradydysrhythmia in two cats. JFMS Open Rep. 2020;6(1):2055116919898292.

Tuomi J, Rantamäki LK, Tanskanen R. Experimental infection of cattle with several *Borrelia burgdorferi* sensu lato strains; immunological heterogeneity of strains as revealed in serological tests.Vet Microbiol. 1998;60(1):27-43.

Uesaka K, Maezawa M, Inokuma H. Serological survey of *Borrelia* infection of dogs in Sapporo, Japan, where *Borrelia* garinii infection was previously detected. J Vet Med Sci. 2016;78(3):463-5.

Veronesi F, Laus F, Passamonti F, Tesei B, Piergili Fioretti D, Genchi C. Occurrence of *Borrelia lusitaniae* infection in horses. Vet Microbiol. 2012;160(3-4):535-8.

Vogt NA, Sargeant JM, MacKinnon MC, Versluis AM. Efficacy of *Borrelia burgdorferi* vaccine in dogs in North America: A systematic review and meta-analysis. J Vet Intern Med. 2019;33(1):23-36.

Wack AN, Holland CJ, Lopez JE, Schwan TG, Bronson E. Suspected Lyme borreliosis in a captive adult chimpanzee (*Pan troglodytes*). J Zoo Wildl Med. 2015;46(2):423-6.

Waddell LA, Greig J, Mascarenhas M, Harding S, Lindsay R, Ogden N. The accuracy of diagnostic tests for Lyme disease in humans, a systematic review and meta-analysis of North American Research. PLoS One. 2016;11(12):e0168613. Wodecka B. Michalik J, Lane RS, Nowak-Chmura M, Wierzbicka A. Differential associations of *Borrelia* species with European badgers (*Meles meles*) and raccoon dogs (*Nyctereutes procyonoides*) in western Poland.Ticks Tick Borne Dis. 2016;7(5):1010-6.

Yeung C, Baranchuk A. Diagnosis and treatment of Lyme carditis: JACC Review Topic of the Week. J Am Coll Cardiol. 2019;73(6):717-26.

Zygner W, Górski P, Wedrychowicz H. Detection of the DNA of Borrelia afzelii, Anaplasma phagocytophilum and Babesia canis in blood samples from dogs in Warsaw. Vet Rec. 2009;164(15):465-7.

\*Link defunct