

Ehrlichiosis and Anaplasmosis in Mammals

Bovine Anaplasmosis,
Ovine Anaplasmosis
Canine Monocytic Ehrlichiosis,
Canine Hemorrhagic Fever,
Tropical Canine Pancytopenia,
Tracker Dog Disease,
Canine Tick Typhus,
Nairobi Bleeding Disorder,
Canine Granulocytic Ehrlichiosis,
Canine Granulocytic Anaplasmosis,
Equine Granulocytic Ehrlichiosis,
Equine Granulocytic Anaplasmosis,
Tick-borne Fever,
Pasture Fever,
Cyclic Canine Thrombocytopenia,
Human Monocytic Ehrlichiosis,
Human Granulocytic Anaplasmosis,
Human Granulocytic Ehrlichiosis,
Human Ewingii Ehrlichiosis

Last Updated: May 2024



IOWA STATE UNIVERSITY
College of Veterinary Medicine



Importance

Ehrlichiosis and anaplasmosis are tick-borne diseases caused by obligate intracellular bacteria in the genera *Ehrlichia* and *Anaplasma*. These organisms are widespread in nature, with reservoir hosts that include numerous wildlife as well as some domestic animals. While many infections in pets, livestock and wildlife are subclinical or mild, some organisms can cause more severe illnesses, and certain diseases, such as tick-borne fever of livestock (*Anaplasma phagocytophilum*) or bovine anaplasmosis caused by *Anaplasma marginale*, can be a significant economic burden. Historically, ehrlichiosis and anaplasmosis were seen as strictly animal diseases, but since the 1980s, some organisms have also been recognized to affect people. While human illnesses initially appeared to be rare, increasing numbers of cases have been attributed to these agents since that time, particularly in the U.S. where mandatory reporting has been in place since 2000, and serological studies have uncovered evidence for many subclinical or mild infections.

Etiology

Ehrlichiosis and anaplasmosis are caused by members of the genera *Ehrlichia* and *Anaplasma*, which are pleomorphic, Gram negative, obligate intracellular bacteria in the family Anaplasmataceae, order Rickettsiales. Some species known to affect mammals include *A. phagocytophilum* (tick-borne fever in ruminants; equine, canine and human granulocytic anaplasmosis), *A. marginale* (bovine anaplasmosis), *A. centrale*, *A. bovis* (bovine ehrlichiosis, bovine anaplasmosis), *A. ovis* (ovine anaplasmosis), *A. capra*, *A. platys* (cyclic canine thrombocytopenia), *A. odocoilei*, *E. canis* (canine monocytic ehrlichiosis), *E. chaffeensis* (human monocytic ehrlichiosis), *E. ewingii* (canine granulocytic ehrlichiosis, human ewingii ehrlichiosis), *E. muris* and its subspecies *E. muris eaucloirensis*, *E. ruminantium* (heartwater), *E. minasensis* (previously *E. mineirensis*) and “the Panola Mountain ehrlichia.” Additional poorly characterized organisms, which often have descriptors such as “organisms related to *E. canis*,” might either be variants of a known species or novel agents. For example, one organism in ruminants originally described as *E. canis*-like became the new species *E. minasensis*, while an *E. muris*-like agent in North America is now *E. muris* subsp. *eaucloirensis*. There are also some *Ehrlichia* and *Anaplasma* that have only been reported in ticks, birds or reptiles but might infect mammals.

Each species of *Anaplasma* or *Ehrlichia* tends to infect certain blood cells or cell fragments (platelets), which can help distinguish these organisms in blood smears and is also reflected in the names of some diseases. *E. canis*, *E. chaffeensis*, *E. muris* and *A. bovis* are mainly found in monocytes, though they sometimes occur in other leukocytes; *E. ewingii* and *A. phagocytophilum* are primarily detected in granulocytes; *A. marginale*, *A. centrale*, *A. ovis* and *A. capra* infect red blood cells; and *A. platys* and *A. odocoilei* circulate in platelets.

This factsheet provides an overview of the illnesses caused by most of the organisms in the genera *Anaplasma* and *Ehrlichia* affecting mammals, except *E. ruminantium* (formerly *Cowdria ruminantium*), which causes heartwater in ruminants and is described in a separate factsheet.

Taxonomy note: Major revisions among the Anaplasmataceae in 2001 resulted in a number of name changes. *Ehrlichia bovis* and *E. platys* became *Anaplasma bovis* and *A. platys*, while the former *Ehrlichia risticii*, which causes Potomac horse fever/ equine monocytic ehrlichiosis, and *Ehrlichia sennetsu*, the agent of sennetsu fever in humans, were transferred to the genus *Neorickettsia*, whose members have life cycles involving trematodes rather than ticks. The most controversial change was consolidating three organisms previously considered separate species - *Ehrlichia equi*, the agent of equine granulocytic ehrlichiosis, *Ehrlichia phagocytophila*, responsible for tick-borne fever in ruminants, and “the agent of human granulocytic ehrlichiosis” - into the single species *A. phagocytophilum*. As a result, *A. phagocytophilum* is a particularly heterogeneous organism, with North American variants, for instance, causing illnesses in humans, dogs and horses but not cattle or small ruminants, while all of these hosts are affected by the variants in Europe.

Ehrlichiosis and Anaplasmosis

Species Affected

Note: Most infections reported in animals are based on PCR, which sometimes detects related ehrlichia or anaplasma; relatively few hosts have been confirmed by isolation of the organism or experimental inoculation.

Ehrlichia canis, *E. minasensis* and related organisms

E. canis affects dogs and other canids, which are also the reservoir hosts. Evidence for this or a closely related organism has been reported in cats and other felids, dromedary camels, a goat, a sika deer (*Cervus nippon*), some South American primates (e.g., *Sapajus apella*, *Callithrix* sp.), coatis (*Nasua* spp.), Eurasian otters (*Lutra lutra*) and raccoons (*Procyon lotor*); however, one attempt to inoculate raccoons with *E. canis* was unsuccessful.

E. minasensis was originally found in cattle but also seems to infect goats, deer and horses.

Ehrlichia chaffeensis

Clinical signs caused by *E. chaffeensis* have been reported in dogs, captive lemurs and experimentally infected calves, though there are currently no reports of cases in naturally infected cattle. This organism also seems to infect some cervids, cats, various wild canids and felids, black-tailed marmosets (*Mico melanurus*), raccoons, badgers (*Meles meles*), American mink (*Neovison vison*) and some rodents. It has been documented in goats, though they do not seem to be readily infected in the laboratory. There are also reports of *E. chaffeensis* DNA in some birds.

Which species maintain *E. chaffeensis* is not entirely clear, though white-tailed deer (*Odocoileus virginianus*) are thought to be a major reservoir host in North America.

Ehrlichia ewingii

E. ewingii is known to affect dogs and experimentally infected goats. Deer and canids have been proposed as reservoir hosts. This organism has also been detected subclinically in other species, such as cats, captive tigers and a goat, and antibodies were found in coyotes.

Ehrlichia muris and the Panola Mountain Ehrlichia

Rodents seem to be the reservoir hosts for *E. muris* subsp. *muris* and *E. muris* subsp. *eaucalarensis*. *E. muris* and the Panola Mountain ehrlichia have been found in deer, and goats are susceptible to experimental inoculation with the latter organism. Two dogs that were PCR positive for either *E. muris* or the Panola Mountain ehrlichia had clinical signs consistent with ehrlichiosis, though a causative role for these organisms as canine pathogens remains to be proven.

Anaplasma phagocytophilum

A. phagocytophilum seems to have an exceptionally wide host range that includes dogs, cats, equids, domestic ruminants, camels, South American camelids, reindeer (*Rangifer tarandus*), and many additional species, such as various cervids and other wild ungulates, bears, wild canids

and felids, raccoon dogs (*Nyctereutes procyonoides*), pine marten (*Martes martes*), wild boar (*Sus scrofa*), raccoons, opossums, striped skunks (*Mephitis mephitis*), hares (*Lepus* spp.), rodents and nonhuman primates. It has been detected in birds and lizards, though the isolates found in lizards seem to differ significantly from those in mammals. The proposed maintenance hosts are diverse and might vary between regions and *A. phagocytophilum* variants. In North America, they are thought to include rodents and cervids.

Clinical cases have been reported in a number of mammals including domestic ruminants, dogs, wolves (*Canis lupus*), cats, equids, llamas, alpacas and non-human primates. The syndrome known as tick-borne fever mainly affects domestic ruminants, particularly sheep and cattle, but it has also been reported in some cervids. North American variants of *A. phagocytophilum* can affect dogs, cats, horses and llamas, but unlike European variants, they do not seem to cause illness in cattle or small ruminants.

Anaplasma bovis

Cattle and water buffalo are the primary hosts for *A. bovis*, but there are also reports of this organism or a closely related species in other mammals including goats, sheep, horses, various cervids, dogs, cats, wild felids and canids, raccoon dogs, coatis, Mongolian gazelles (*Procapra gutturosa*), giraffes (*Giraffa camelopardalis*), raccoons, wild boars, bears, some rabbits and hares, cynomolgus macaques (*Macaca fascicularis*) and rodents/ small mammals. Clinical cases are usually seen in cattle, but an illness in one horse might have been caused by this organism.

Anaplasma marginale and *A. centrale*

A. marginale infects bovids including cattle, water buffalo, yaks, bison (*Bison* spp.) and African buffalo (*Syncerus caffer*), but it or a closely related organism has also been found in sheep, camels, equids, some cervids, sable antelope (*Hippotragus niger*), and tufted capuchins (*Sapajus apella*). Additional species, such as blesbok (*Damaliscus pygargus phillipsi*), common duiker (*Sylvicapra grimmia*), and black wildebeest (*Connochaetes gnou*) can be infected experimentally. Bovids, particularly cattle, appear to be the reservoir hosts, and illnesses are mainly seen in cattle.

A. centrale has been reported in bovids (cattle, water buffalo, African buffalo) and some other ungulates, such as wildebeest (*Connochaetes* spp.), eland (*Taurotragus oryx*), waterbuck (*Kobus ellipsiprymnus*) and sika deer (*Cervus nippon*). Experimental infections have been established in blesbok, common duiker and black wildebeest.

Anaplasma ovis

A. ovis occurs in sheep, goats, some of their wild relatives (e.g., bighorn sheep, *Ovis canadensis*) and various cervids. Small ruminants and some cervids are thought to be reservoir hosts. This organism also seems to infect cattle and camels occasionally, and it has been reported in sable antelope, red foxes (*Vulpes vulpes*) and donkeys.

Ehrlichiosis and Anaplasmosis

Anaplasma capra

A. capra has been detected in sheep, goats, cattle, water buffalo, dogs, wild onagers (*Equus hemionus onager*), Japanese serows (*Capricornis crispus*), takins (*Budorcas taxicolor*) and a number of cervids. At present, it is not known to cause any illness in animals.

Anaplasma platys

A. platys affects dogs, but it or a related organism has also been found in red foxes, cats, cattle, water buffalo, small ruminants, Bactrian and dromedary camels, sable antelope and some cervids. At least some of the organisms identified in camels might belong to a proposed new species, *Candidatus A. camelii*.

Anaplasma odocoilei

A. odocoilei has been identified in various cervids, and it or a related organism was found in goats.

Zoonotic potential

Organisms that have been recognized as human pathogens include *E. chaffeensis* (human monocytic ehrlichiosis), *E. ewingii* (human ewingii ehrlichiosis) and *A. phagocytophilum* (human granulocytic anaplasmosis). However, some *A. phagocytophilum* variants might not affect people, or do so only rarely. *E. canis*, *E. muris*, *E. muris eauclairiensis*, *A. capra*, *A. bovis*, *A. ovis*, *A. platys* and the Panola Mountain ehrlichia have been implicated in a few cases, with varying levels of evidence.

Geographic Distribution

E. canis, *A. phagocytophilum*, *A. marginale*, *A. centrale*, *A. ovis*, *A. bovis* and *A. platys* have been found on most or all continents, though their distribution varies with the occurrence and density of their tick vectors. *A. marginale*, *A. centrale* and *A. platys* are particularly common in tropical and subtropical regions, with a more limited presence in temperate climates. Some *A. phagocytophilum* variants seem to occur in limited locations, which influences the hosts affected in an area. *E. chaffeensis* and *E. minasensis*, which were originally detected in only a few areas, now also appear likely to be cosmopolitan.

Other organisms might either be geographically limited or incompletely surveyed. *A. capra* was originally identified in China, but it has since been found in a number of other countries in Asia, Europe and North Africa. *E. ewingii* has been documented in parts of North and South America and was also reported from Cameroon, Africa. *E. muris* is known to occur in parts of Eurasia, while *E. muris eauclairiensis* was described in North America. *A. odocoilei* has been found in North and South America and the Philippines, and the Panola Mountain ehrlichia has been reported in North America, the Caribbean and Africa.

Transmission

Various hard ticks (family Ixodidae) are the biological vectors for members of the genera *Ehrlichia* and *Anaplasma*. Some organisms seem to be transmitted mainly by a few

specific vectors, while others occur in a wide variety of ticks. How long a tick must remain attached to infect an animal is generally not known, but at least 24-48 hours usually seems to be required for *A. phagocytophilum*, with a few mice seroconverting after 12 hours. Some organisms, including *A. marginale* in cattle and *A. ovis* in sheep, establish chronic, low level infections in their hosts. *E. canis* and *E. ewingii* infections can also persist for years in some dogs, while *E. chaffeensis* seems to be eliminated more rapidly, disappearing in a few weeks or months in one study.

Ehrlichia and *Anaplasma* can be acquired occasionally by other routes, especially in procedures that transfer blood. Mechanical transmission on blood-contaminated fomites (e.g., needles, dehorning equipment) or by biting insects seems to be important for some species that infect RBCs, such as *A. marginale*. However, this is not necessarily the case for all species of *Ehrlichia* and *Anaplasma*, and one study found that insects rarely seemed to transmit *A. phagocytophilum* mechanically. A few cases of ehrlichiosis or anaplasmosis in people were apparently acquired in blood transfusions and bone marrow or solid organ transplants. The risk probably depends on the specific organism and blood components, but transfusions of both leukoreduced and non-leukoreduced RBCs have transmitted ehrlichiosis.

Transplacental transmission is also possible, at least for some organisms. *A. marginale* can cross the placenta in cattle, resulting in healthy but persistently infected calves. There is also evidence for *in utero* infections with *A. phagocytophilum* in cattle, sheep, horses and dogs, and perinatal transmission of this organism, either *in utero* or during delivery, was documented in two women. Another study suggested the possibility of transplacental transmission of *A. platys* in dogs. One report found DNA from *A. ovis*, *A. bovis* and *A. phagocytophilum* in the milk of ruminants, though the significance of this finding remains to be determined.

Other routes of transmission are controversial. Direct contact with deer blood might have caused *A. phagocytophilum* infections in three people, but this is still uncertain as other exposures (e.g., tick bites) could not be ruled out. Possible person-to-person transmission of this organism was suggested in a Chinese hospital, after an outbreak in relatives and healthcare workers who had been in close, direct contact with a severely hemorrhaging, intubated patient. However, it now appears possible that another virus might have been responsible for their symptoms.

Disinfection

Disinfection is unimportant for controlling *Ehrlichia* and *Anaplasma*, which are obligate intracellular organisms. Disinfectants employed against other Gram negative bacteria would probably be effective against these agents.

Ehrlichiosis and Anaplasmosis

Infections in Animals

Incubation Period

Incubation periods ranging from 7 to 100 days have been reported for bovine anaplasmosis caused by *A. marginale*, with most cases becoming apparent in about 2-5 weeks. *A. phagocytophilum* generally affects ruminants and dogs in approximately 1-2 weeks, and horses in 1-3 weeks. Dogs infected with *E. canis* usually develop the acute form of canine monocytic ehrlichiosis within 2-4 weeks of the tick bite, while the chronic form can occur months or years later.

Clinical Signs

Ehrlichiosis and anaplasmosis generally present as a febrile illness with nonspecific clinical signs, often accompanied by elevated liver enzymes and reductions in leukocyte, RBC and/or platelet numbers. Some agents that infect RBCs also tend to cause overt signs of anemia. Concurrent infections may influence the clinical presentation or increase the severity of the illness.

Organisms that affect white blood cells: *E. canis* and other species in dogs and cats

E. canis infections in dogs (canine monocytic ehrlichiosis; CME) range from subclinical to severe. Many dogs have no clinical signs when they become infected, but others develop a nonspecific illness with signs such as fever, lethargy, anorexia, lymphadenopathy and splenomegaly, often accompanied by thrombocytopenia and mildly elevated hepatic enzymes. Gastrointestinal signs (e.g., vomiting, diarrhea), edema in the legs or scrotum, respiratory signs (nasal discharge, coughing, dyspnea), and uveitis or other ocular signs may also be seen in some animals. Bleeding disorders (e.g., petechiae, ecchymoses or mild epistaxis) and neurological signs are possible, though less common than in chronic CME. Polyarthropathy with lameness, stiffness or joint swelling has also been described, but some authors suggest this is caused by coinfections and not *E. canis*.

Dogs with acute CME often recover, though severe cases can be fatal. Recovered dogs, as well as those that never developed clinical signs, sometimes remain subclinically infected for months or years. These animals, which may have mild thrombocytopenia, can eventually clear the organism or develop chronic CME. Chronic CME resembles the acute illness, but is more severe, and often includes additional signs such as chronic weight loss and edema, as well as pancytopenia with various combinations of leukopenia, thrombocytopenia and anemia. Some dogs develop bleeding disorders and/or various neurological signs, or other complications such as renal failure, interstitial pneumonia, liver disease, polymyositis or reproductive disorders with prolonged bleeding during estrus, inability to conceive, abortion and/or neonatal death. In addition to being debilitating, chronic CME can eventually be fatal.

Clinical cases caused by *E. ewingii* (canine granulocytic ehrlichiosis), *A. phagocytophilum* (canine granulocytic anaplasmosis) and *E. chaffeensis* can resemble CME;

however, dogs infected with *A. phagocytophilum* generally seem to have milder illnesses, with fewer animals developing hemorrhagic or neurological signs and only rare fatalities. Healthy dogs experimentally infected with *E. canis*, *E. chaffeensis* or *A. phagocytophilum* usually remained asymptomatic or had only an intermittent fever, with *A. phagocytophilum* causing the mildest signs in one comparative study.

One clinical case attributed to *E. muris* was characterized by fever, thrombocytopenia, joint stiffness and carpal pain, while a dog infected with the Panola Mountain ehrlichia had hepatomegaly, mild thrombocytopenia and lymphocytosis with atypical lymphocytes. Pre-existing hepatitis could have been responsible for some of the clinical signs in the latter case, though the thrombocytopenia and lymphocytosis resolved with doxycycline.

Ehrlichia and *Anaplasma* have been implicated only rarely in clinical cases in cats. Nonspecific signs of fever, lethargy and inappetence are reported most often, but other clinical signs seen in dogs, including joint pain and rare hemorrhagic or neurological signs, have been documented.

Organisms that affect white blood cells: *A. phagocytophilum* in equids

Equine granulocytic anaplasmosis, which is caused by *A. phagocytophilum*, is an acute illness that is generally more severe in horses > 3 years old. While signs in animals under a year of age may be limited to fever, older horses are more likely to appear ill, with a decreased appetite, lethargy and icterus, sometimes followed by petechiae, distal limb edema and myalgia, with stiffness of gait, reluctance to move or lameness. A few reports have described animals with apparent neurological signs (e.g., ataxia, reduced alertness), upper respiratory disease with dysphagia due to laryngeal hemiplegia or pharyngeal edema, bicavitary effusion or transient ventricular arrhythmia. Laboratory abnormalities can include thrombocytopenia, anemia and leukopenia. Deaths seem to be very rare.

Organisms that affect white blood cells: *A. phagocytophilum* in ruminants

Clinical cases caused by *A. phagocytophilum* in sheep (tick-borne fever) mainly occur in young lambs, or in older sheep newly introduced to tick-infested areas. The primary syndrome is a sudden fever that lasts for 4-10 days, which may be accompanied by nonspecific signs of illness, and can be followed by abortions and stillbirths in ewes that become infected during the late stages of gestation. Deaths are uncommon except in some aborting ewes, or in young lambs that develop secondary infections.

In cattle, this disease is most often seen in dairy animals recently turned out to pasture. The clinical signs, which are nonspecific and variable in severity, often include decreased milk production, with some animals also developing respiratory signs and/or edema of the lower limbs. Abortions and stillbirths may be seen in pregnant cows and semen quality is temporarily reduced in bulls.

Ehrlichiosis and Anaplasmosis

Organisms that affect white blood cells: *A. bovis*

A. bovis infections seem to be subclinical in many cattle, but some develop a fever and nonspecific signs of illness, including decreased milk production. Edema, nasal discharge, occasional abortions or rare neurological signs have also been reported. Some infected cattle in Japan had mild to severe anemia, but coinfections with *Anaplasma* and *Theileria* complicate the interpretation of these cases. While deaths are possible, severe cases seem to be uncommon in most regions.

One old horse infected with *A. bovis* had a fever, anorexia, lethargy and severe dehydration.

Organisms that affect white blood cells: *Ehrlichia chaffeensis* and *E. minasensis* in ruminants, cervids and other species

Although clinical cases caused by *E. chaffeensis* have not been reported in naturally infected ruminants, experimentally infected dairy calves sometimes developed mild to severe illnesses that ranged from fever alone, accompanied by decreased leukocyte and platelet numbers, to a febrile illness with progressive weakness in the hindlegs, recumbency and death. Most infections in deer seem to be subclinical, but some experimentally infected fawns had a periodic mild fever on days when the organisms were found in the blood. An outbreak in captive ring-tailed lemurs (*Lemur catta*) and red ruffed lemurs (*Varecia variegata rubra*) appeared as a nonspecific febrile illness accompanied by thrombocytopenia and hyperbilirubinemia.

Clinical cases caused by *E. minasensis* in one naturally infected calf and one experimentally infected calf were characterized by fever, lethargy, thrombocytopenia, leukopenia and anemia. Another attempt to inoculate calves with this organism resulted in no clinical signs. The Panola Mountain ehrlichia caused brief or intermittent fevers in two experimentally infected goats, with serous nasal discharge in one animal.

Organisms that affect red blood cells: *A. marginale*, *A. centrale* and *A. ovis*

Clinical anaplasmosis caused by *A. marginale* in cattle is characterized by nonspecific signs of illness and mild to severe anemia, which may result in pale mucous membranes with elevated heart and respiratory rates, and icterus in the later stages. Neurological signs have been reported (probably from oxygen limitation to the CNS), pregnant cows may abort, and bulls may have a transient decrease in fertility. Severely affected animals may die, especially when stressed. Peracute cases, which are uncommon and occur mainly in high-producing dairy cattle, can be fatal within a few hours of the onset of clinical signs. Cases are typically milder in young animals than those first infected as adults, and infections in cattle less than a year of age are usually subclinical. Chronically infected cattle are asymptomatic but may relapse if immunosuppressed.

A. ovis infections in sheep are similar but often milder or subclinical. Overt signs of illness are most common in animals that are stressed or coinfecting with other organisms. *A. centrale* infections in cattle also tend to be mild, and this organism is used as a vaccine for *A. marginale*.

Organisms that affect platelets: *A. platys* and *A. odocoilei*

Many or most *A. platys* infections in dogs seem to be subclinical, but there are also reports of nonspecific febrile illnesses, accompanied by thrombocytopenia and anemia, with petechiae and other signs of bleeding disorders. Although many cases may be mild, severe hemorrhages and fatalities are possible. Two case reports described sick cats with thrombocytopenia that were infected with *A. platys*, though a causative role in their clinical signs is unclear. One cat had a urinary tract infection that could account for its lethargy and anorexia, while the other had multiple myeloma and several coinfections.

Fawns experimentally infected with *A. odocoilei* sometimes had multiple transient episodes of thrombocytopenia but no overt clinical signs.

Post Mortem Lesions [Click to view images](#)

Gross lesions in ehrlichiosis and anaplasmosis are generally nonspecific, and often include enlargement of the spleen. Dogs with canine monocytic ehrlichiosis may also have edema in the legs, ascites, hydropericardium, lymphadenopathy (particularly of the mesenteric nodes) and/or hemorrhagic lesions in various internal organs, subcutaneous tissues and eyes. Hemorrhages are more common and widespread in chronic than acute CME.

Some illnesses, such as bovine anaplasmosis caused by *A. marginale*, also include signs of anemia (e.g., pallor) and icterus. The liver is often enlarged in bovine anaplasmosis, and may be yellowish-orange and mottled; the hepatic and mediastinal lymph nodes may be brown; and petechiae are often found on the serosa of internal organs, especially the heart and pericardium.

The usual lesions in equine granulocytic anaplasmosis are petechiae, ecchymoses and edema in the subcutaneous tissues and fascia, mainly in the legs, though interstitial pneumonitis has been reported in some animals, and other internal organs may have inflammatory lesions and serosal hemorrhages in severe cases.

Diagnostic Tests

Organisms and/or their nucleic acids can sometimes be found in the blood, bone marrow and/or samples from affected sites (e.g., joint fluid) in live animals or in various tissues, such as the spleen, liver, heart, lung, kidney and blood vessels, at necropsy. PCR tests are available for some common species of *Anaplasma* and *Ehrlichia*, and are increasingly employed in diagnosis. They do not always detect very small numbers of organisms, for instance *E. canis* in the peripheral blood of subclinically or chronically infected dogs, or *A. marginale* in chronically infected cattle.

Ehrlichiosis and Anaplasmosis

Loop-mediated isothermal amplification (LAMP) assays have also been published for some organisms.

Observing intracytoplasmic inclusions in Wright, Giemsa or Romanowsky (e.g., Diff-Quik™) stained blood, bone marrow or fresh tissue impression smears can help support the diagnosis, though it cannot definitively identify these organisms. Inclusion bodies, which are called morulae in WBCs, are generally stippled blue-gray to dark blue or purple, and are visible at 400x or 1000x magnification. *E. canis*, *E. chaffeensis*, *E. muris* and *A. bovis* inclusion bodies are mainly found in monocytes; *A. phagocytophilum* and *E. ewingii* primarily in neutrophils; *A. marginale*, *A. centrale*, *A. ovis* and *A. capra* in RBCs; and *A. platys* and *A. odocoilei* in platelets. Organisms found in WBCs have occasionally been detected in leukocytes other than those they usually infect.

Inclusion bodies are most likely to be observed early in the acute illness, and are rarely found in chronically infected animals. They are also more easily detected in some diseases than others. The morulae of *E. canis* and *E. chaffeensis*, for instance, are uncommon in the blood of dogs (though buffy coat may be more successful). The cyclic nature of *A. platys*, as well as the relatively small percentage of platelets with inclusion bodies, can also make this organism difficult to find.

Clinical cases can also be confirmed serologically by seroconversion or a rising antibody titer. Single titers should be interpreted with caution, as antibodies to these organisms are common in healthy animals. Indirect immunofluorescent antibody (IFA) tests and ELISAs, the most commonly used assays, are available for most clinically important organisms. Cross-reactivity can be an issue, but mainly occurs between members of the same genus. It is occasionally exploited to provide evidence of infection with organisms not readily cultivated *in vitro*, such as *E. ewingii*. Immunoblotting (Western blotting), which can distinguish reactivity to different species, is mainly used in research.

Culture is considered impractical for routine diagnosis, as it can take up to 2-6 weeks and requires specialized techniques that are unavailable at most diagnostic laboratories. Most organisms, including some that were previously considered uncultivable (e.g., *E. ewingii*, *A. ovis*) can be grown *in vitro* for at least a short time.

Treatment

Anaplasmosis and ehrlichiosis are resistant to most antibiotics but can be treated with a few drugs, such as tetracyclines, combined with supportive care as needed. Some organisms may not be cleared completely, though this might depend on the specific antibiotic used and the dose and length of time the animal is treated. Treatment of chronic canine monocytic ehrlichiosis (*E. canis*) may be difficult, though uncomplicated acute cases usually respond promptly.

Control

Disease reporting

Veterinarians who suspect an animal is infected with *Ehrlichia* or *Anaplasma* should follow their national and/or

local guidelines for disease reporting. State authorities should be consulted for regulations in the U.S.

Prevention

Limiting tick bites with acaricides, tick repellents and/or environmental modifications that make habitats less attractive to ticks (e.g., brush removal) is expected to reduce the risk of ehrlichiosis and anaplasmosis; however, the widespread nature of some organisms and their tick vectors may make control difficult. Prompt removal of ticks from the animal is also expected to be helpful.

Animal management can reduce disease impacts in livestock, for instance by first introducing animals to infected pastures at an age when they are less likely to develop severe signs. Tetracyclines have sometimes been employed prophylactically in dogs or domestic ruminants. Although vaccines are not available for most organisms, a live vaccine containing *A. centrale* can be used to induce immunity to *A. marginale*.

PCR testing for certain common organisms may be advisable in animals used as blood donors, but might not detect some chronically infected individuals.

Morbidity and Mortality

Asymptomatic infections with some species of *Ehrlichia* and *Anaplasma* seem to be relatively common in mammals, with serological surveillance suggesting that up to 50-60% or more of the animals in some areas have been exposed to these organisms. In one prospective U.S. study, 70-100% of dogs walked in tick habitats without tick control became subclinically infected with *E. ewingii* or *E. chaffeensis*, though none of the 10 dogs acquired *E. canis*.

Clinical cases are seasonal in temperate regions, usually appearing during or soon after the active tick season. Some diseases, such as tick-borne fever, tend to appear as outbreaks when animals are first exposed to tick habitats (e.g., dairy cattle turned out to pasture after the winter); others can be seen sporadically throughout the season. Clinical cases can also present occasionally outside tick season. Some occur when ticks such as *Rhipicephalus sanguineus*, an important vector for *E. canis*, become established indoors in houses and kennels. Others are caused by illnesses with long or indeterminate incubation periods, such as chronic canine monocytic ehrlichiosis.

Clinical cases vary in severity, depending on the specific organism and host. Bovine anaplasmosis from *A. marginale* can be a serious illness, with reported mortality rates of approximately 30-50% if cattle are first exposed as adults (> 2 years). It is particularly severe in high-producing dairy cows. However, animals raised in some endemic regions are likely to encounter this organism at a younger age, which can reduce its impact. Deaths are uncommon in equine granulocytic anaplasmosis, and tick-borne fever of ruminants is mainly an economic issue, with low mortality but significant decreases in milk production and the potential for reproductive losses. Abortion storms with losses of up to 90% are sometimes seen in previously

Ehrlichiosis and Anaplasmosis

unexposed pregnant ewes. *A. bovis* infections are also said to be economically important in Africa, though they tend to be mild or subclinical in many other regions. Immunity to some organisms does not seem to be absolute, and may fall quickly if animals are removed from tick-infested areas.

Canine monocytic ehrlichiosis, caused by *E. canis*, is usually the most serious illness in dogs. While asymptomatic infections with this organism are fairly common, and most animals that become acutely ill recover, the disease is potentially severe and chronic cases can be difficult to cure. Infections with *E. chaffeensis* or *A. phagocytophilum* are thought to be overall milder in dogs, though severe illnesses and deaths are possible.

Infections in Humans

Incubation Period

Incubation periods from 5 to 21 days have been reported for human ehrlichiosis and anaplasmosis, with most cases appearing in about 1-2 weeks.

Clinical Signs

As in animals, the consequences of infection with *Ehrlichia* or *Anaplasma* range from asymptomatic infections to a severe, potentially fatal illness. Clinical cases caused by *E. chaffeensis* (human monocytic ehrlichiosis; HME) generally appear as an acute febrile illness with nonspecific signs such as headache, myalgia and arthralgia, often accompanied by thrombocytopenia, mild to moderate leukopenia, elevated levels of liver enzymes, and in some cases anemia. Some patients may also have respiratory or gastrointestinal signs (vomiting, diarrhea, abdominal pain) or a nonpruritic rash that usually spares the palms and soles and may be maculopapular, petechial or characterized by diffuse erythroderma. Gastrointestinal signs and rashes are more common in children than adults.

The reported complications of HME are diverse and include opportunistic infections, meningitis or meningoencephalitis, cardiovascular failure, myocarditis, liver dysfunction, acute renal failure, interstitial pneumonia, respiratory distress syndrome and hemorrhages, as well as a multisystemic disease that resembles toxic shock syndrome or septic shock. Complications, severe cases and deaths are more likely in elderly, very young, or immunocompromised patients, or in those with other concurrent illnesses, but fatal cases have been seen even in previously healthy young patients.

Illnesses caused by *A. phagocytophilum* (human granulocytic anaplasmosis; HGA) and *E. ewingii* (human ewingii ehrlichiosis; HEE) are similar, but tend to be less severe, and leukopenia, thrombocytopenia and abnormal liver function tests are sometimes absent in HEE. Rashes are generally said to be infrequent (< 10%) in both diseases; however, some recent reports found them in up to 15-17% of patients with HGA in the U.S. In European HGA patients, a rash was mostly seen in people coinfecting with *Borrelia*. Complications seem to be less frequent in HGA than HME,

and have rarely been reported in HEE. CNS involvement is uncommon.

The few clinical cases attributed to *E. canis*, *E. muris*, *E. muris euclairensis*, *A. ovis*, *A. bovis* or *A. platys* were mostly characterized by similar nonspecific febrile illnesses, though one person PCR positive for *A. platys* had a complicated medical history that included hospitalization for encephalitis of uncertain etiology, with ongoing seizures and migraines after discharge. Because she was not tested for *A. platys* until later, and also had two tickborne coinfections, a role for *A. platys* in her neurological signs is speculative. The Panola Mountain ehrlichia case was also atypical: the main symptom, which resolved promptly with doxycycline, was persistent neck soreness after a tick bite. Some *A. capra* infections and one of the *A. bovis* infections reported from China included one or more eschars, which is not typical of anaplasmosis or ehrlichiosis (but can be caused by some *Rickettsia*), in addition to signs more characteristic of these illnesses. Five of 28 cases attributed to *A. capra* were severe enough to require hospitalization, with one patient developing CNS signs.

Diagnostic Tests

Ehrlichiosis or anaplasmosis in humans is diagnosed similarly to cases in animals. Morulae can be found in the blood or buffy coat of 25-75% of the patients infected with *A. phagocytophilum*, but <10% of those infected with *E. chaffeensis*, and are most likely to be detected during the first week of illness. In rare cases, they may be found in CSF. PCR is more sensitive than microscopic examination for morulae, though it may not detect DNA after the first 2 weeks. Organisms can also be found sometimes in formalin-fixed tissue samples (e.g., bone marrow biopsies; autopsy samples from spleen, lymph node, liver or lung) with immunohistochemistry. Serology, with seroconversion or a fourfold rise in titer, is diagnostic, though single high titers are sometimes employed for a presumptive diagnosis. As in animals, culture is mainly used in research.

Treatment

Ehrlichiosis and anaplasmosis are usually treated with antibiotics, with supportive care as necessary. Uncomplicated early cases usually respond promptly, but prolonged treatment may be necessary for severe illnesses.

Prevention

The risk of ehrlichiosis or anaplasmosis can be reduced by avoidance of tick habitats or by discouraging tick bites with appropriate clothing (e.g., pyrethrin-impregnated garments, long-sleeved shirts and trousers tucked into socks), repellents (e.g., DEET) and insecticides. Any attached ticks should be removed promptly. Treatment of the environment with acaricides and/or modification of tick habitats can also be used to decrease tick populations, but acaricide use may have detrimental effects on non-target species and promotes acaricide resistance.

Ehrlichiosis and Anaplasmosis

Morbidity and Mortality

Significant numbers of healthy people, up to 20% or more, have antibodies to *Ehrlichia* or *Anaplasma* in some regions, suggesting that symptomatic infections or mild illnesses might be relatively common. Some prospective studies have also reported subclinical seroconversion in many or most of those who are naturally exposed. Clinical cases tend to be more severe in immunocompromised or elderly patients, very young children, and individuals with other illnesses, though serious illnesses can be seen occasionally in healthy people of all ages. The estimated case fatality rate in the U.S. is approximately 1% for HME, with higher rates of 3-4% in young children less than 5 years of age and adults over 70, and < 1% for HGA. Because many mild cases could be missed, it is possible that these values overestimate the severity of these illnesses.

In the U.S., where mandatory reporting of human ehrlichiosis and anaplasmosis was implemented in 1999, the recorded incidence of HGA rose from 273 cases (1.4 per million persons) in 2000 to approximately 4000 to 5760 cases per year between 2017 and 2019. The incidence of HME likewise increased from 201 cases (< 1 per million persons) to about 1300 to 2100 cases/ year between 2013 and 2019. It should be noted that more than half of these reports describe probable HGA and HME cases, based on a single antibody titer, rather than confirmed cases. The number of clinical cases documented in countries without mandatory reporting is generally much lower, though seroprevalence in some regions is higher than in North America.

Fewer clinical cases have been reported for other organisms, though this might also be influenced by the availability of diagnostic tests. Human ewingii ehrlichiosis (*E. ewingii*) was initially diagnosed in only a few patients, most of whom were immunosuppressed, but an additional 55 cases affecting both healthy and immunocompromised or elderly individuals were reported to U.S. federal authorities between 2008 and 2012. No deaths and few complications have been associated with this organism, as of 2022. Most other species of *Ehrlichia* and *Anaplasma* have been implicated in very few human cases; however, hospital surveillance attributed 28 clinical cases to *A. capra* in China in 2015, and *E. muris* has been implicated in at least 2 PCR-confirmed and 84 serologically diagnosed cases in Russia. Approximately 1% of the residents in Tokyo, Japan had antibodies to the latter organism, though cross-reactions with other organisms could not be ruled out.

Internet Resources

[Centers for Disease Control and Prevention \(CDC\). Anaplasmosis](#)

[Centers for Disease Control and Prevention \(CDC\). Ehrlichiosis](#)

[Department of Agriculture, Fisheries and Forestry, Australia. Ehrlichiosis in Dogs](#)

[eMedicine.com. Ehrlichiosis](#)

[European Advisory Board on Cat Diseases. Guideline for Anaplasma, Ehrlichia, Rickettsia infections](#)

[International Veterinary Information Service \(IVIS\)](#)

[Medscape. Anaplasma phagocytophilum](#)

Sainz et al. [Guideline for veterinary practitioners on canine ehrlichiosis and anaplasmosis in Europe](#)

[StatPearls. Anaplasma phagocytophilum](#)

[StatPearls. Ehrlichiosis](#)

[The Merck Manual](#)

[The Merck Veterinary Manual](#)

[World Organization for Animal Health \(WOAH\). Bovine anaplasmosis](#)

Acknowledgements

This factsheet was written by Anna Rovid Spickler, DVM, PhD from the Center for Food Security and Public Health. The U.S. Department of Agriculture Animal and Plant Health Inspection Service (USDA APHIS) provided funding for this factsheet through a series of cooperative agreements related to the development of resources for initial accreditation training.

The following format can be used to cite this factsheet. Spickler, Anna Rovid. 2024. Ehrlichiosis and anaplasmosis. Retrieved from <http://www.cfsph.iastate.edu/DiseaseInfo/factsheets.php>.

References

- Abdullah HHAM, Aboelsoued D, Farag TK, Abdel-Shafy S, Abdel Megeed KN, Parola P, Raoult D, Mediannikov O. Molecular characterization of some equine vector-borne diseases and associated arthropods in Egypt. *Acta Trop.* 2022;227:106274.
- Adaszek L, Górna M, Skrzypczak M, Buczek K, Balicki I, Winiarczyk S. Three clinical cases of *Anaplasma phagocytophilum* infection in cats in Poland. *J Feline Med Surg.* 2013;15(4):333-7.
- Adaszek L, Wilczyńska A, Ziętek J, Kalinowski M, Teodorowski O, Winiarczyk D, Skrzypczak M, Winiarczyk S. Granulocytic anaplasmosis in captive ring-tailed lemur (*Lemur catta*) in Poland. *BMC Vet Res.* 2021;17(1):118.
- Addo SO, Baako BOA, Bentil RE, Addae CA, Behene E, Asoala V, Sallam M, Mate S, Dunford JC, Larbi JA, Baidoo PK, Wilson MD, Diclaro JW, Dadzie SK. Molecular survey of *Anaplasma* and *Ehrlichia* species in livestock ticks from Kassena-Nankana, Ghana; with a first report of *Anaplasma capra* and *Ehrlichia minasensis*. *Arch Microbiol.* 2023;205(3):92.
- Aktas M, Ozubek S. Molecular evidence for trans-stadial transmission of *Anaplasma platys* by *Rhipicephalus sanguineus* sensu lato under field conditions. *Med Vet Entomol.* 2018;32(1):78-83.

- Alanazi AD, Nguyen VL, Alyousif MS, Manoj RRS, Alouffi AS, Donato R, Sazmand A, Mendoza-Roldan JA, Dantas-Torres F, Otranto D. Ticks and associated pathogens in camels (*Camelus dromedarius*) from Riyadh Province, Saudi Arabia. *Parasit Vectors*. 2020;13(1):110.
- Alcántara-Rodríguez VE, Sánchez-Montes S, Contreras H, Colunga-Salas P, Fierro-Flores L, Avalos S, Rodríguez-Rangel F, Becker I, Walker DH. Human monocytic ehrlichiosis, Mexico City, Mexico. *Emerg Infect Dis*. 2020;26(12):3016-9.
- Altay K, Erol U, Sahin OF, Aytmirzakizi A. First molecular detection of *Anaplasma* species in cattle from Kyrgyzstan; molecular identification of human pathogenic novel genotype *Anaplasma capra* and *Anaplasma phagocytophilum* related strain. *Ticks Tick Borne Dis*. 2022;13(1):101861.
- Altay K, Erol U, Sahin OF, Aytmirzakizi A, Temizel EM, Aydin MF, Dumanli N, Aktas M. The detection and phylogenetic analysis of *Anaplasma phagocytophilum*-like 1, *A. ovis* and *A. capra* in sheep: *A. capra* divides into two genogroups. *Vet Res Commun*. 2022;46(4):1271-9.
- André MR. Diversity of *Anaplasma* and *Ehrlichia/Neoehrlichia* agents in terrestrial wild carnivores worldwide: implications for human and domestic animal health and wildlife conservation. *Front Vet Sci*. 2018;5:293.
- Angeloni VL. Rickettsial diseases [monograph online]. In: James WD, editor. *Military dermatology, Part III: Disease and the environment*. Available at: http://www.wrampc.amedd.army.mil/fieldmed/dermatology/Derm_Textbook_Ch11.pdf. * Accessed 18 Aug 2004.
- Animal Health Australia. National Animal Health Information System (NAHIS). Tropical canine pancytopenia [monograph online]. Available at: <http://www.aahc.com.au/nahis/disease/dislist.asp>. * Accessed 20 Aug 2004.
- Animalu. Ehrlichiosis [monograph online]. eMedicine.com; 2021. Available at: <http://emedicine.medscape.com/article/235839-overview>. Accessed 20 Dec 2023.
- Annen K, Friedman K, Eshoa C, Horowitz M, Gottschall J, Straus T. Two cases of transfusion-transmitted *Anaplasma phagocytophilum*. *Am J Clin Pathol*. 2012;137(4):562-5.
- Anonymous. Epidemiologic notes and reports. Human ehrlichiosis - United States. *Morb Mortal Wkly Rep*. 2001. Available at: <http://www.cdc.gov/mmwr/preview/mmwrhtml/00000020.htm>. Accessed 17 Aug 2004.
- Arnez M, Luznik-Bufon T, Avsic-Zupanc T, Ruzic-Sabljić E, Petrovec M, Lotric-Furlan S, Strle F. Causes of febrile illnesses after a tick bite in Slovenian children. *Pediatr Infect Dis J*. 2003;22(12):1078-83.
- Arraga-Alvarado CM, Qurollo BA, Parra OC, Berrueta MA, Hegarty BC, Breitschwerdt EB. Case report: Molecular evidence of *Anaplasma platys* infection in two women from Venezuela. *Am J Trop Med Hyg*. 2014;91(6):1161-5.
- Arsić B, Gligić A, Ristanović E, Lako B, Potkonjak A, Peruničić M, Pavlović M. A case of human monocytic ehrlichiosis in Serbia. *Srp Arh Celok Lek*. 2014;142(1-2):79-82.
- Atif FA, Mehnaz S, Qamar MF, Roheen T, Sajid MS, Ehtisham-Ul-Haque S, Kashif M, Ben Said M. Epidemiology, diagnosis, and control of canine infectious cyclic thrombocytopenia and granulocytic anaplasmosis: emerging diseases of veterinary and public health significance. *Vet Sci*. 2021;8(12):312.
- Aziz MU, Hussain S, Song B, Ghauri HN, Zeb J, Sparagano OA. Ehrlichiosis in dogs: a comprehensive review about the pathogen and its vectors with emphasis on South and East Asian countries. *Vet Sci*. 2022;10(1):21.
- Bahrami S, Hamidinejat H, Tafreshi ARG. First molecular detection of *Anaplasma phagocytophilum* in dromedaries (*Camelus dromedarius*). *J Zoo Wildl Med*. 2018;49(4):844-8.
- Bakken JS, Dumler JS. Clinical diagnosis and treatment of human granulocytotropic anaplasmosis. *Ann NY Acad Sci*. 2006;1078:236-47.
- Beall MJ, Alleman AR, Breitschwerdt EB, Cohn LA, Couto CG, et al. Seroprevalence of *Ehrlichia canis*, *Ehrlichia chaffeensis* and *Ehrlichia ewingii* in dogs in North America. *Parasit Vectors*. 2012;5:29.
- Ben Said M, Belkahia H, El Mabrouk N, Saidani M, Alberti A, Zobba R, Cherif A, Mahjoub T, Bouattour A, Messadi L. *Anaplasma platys*-like strains in ruminants from Tunisia. *Infect Genet Evol*. 2017;49:226-33.
- Bjoersdorff A, Svendenius L, Owens JH, Massung RF. Feline granulocytic ehrlichiosis -- a report of a new clinical entity and characterization of the infectious agent. *J Small Anim Pract*. 1999;40:20-4.
- Bouza-Mora L, Dolz G, Solórzano-Morales A, Romero-Zuñiga JJ, Salazar-Sánchez L, Labruna MB, Aguiar DM. Novel genotype of *Ehrlichia canis* detected in samples of human blood bank donors in Costa Rica. *Ticks Tick Borne Dis*. 2017;8(1):36-40.
- Bown KJ, Lambin X, Ogden NH, Begon M, Telford G, Woldehiwet Z, Birtles RJ. Delineating *Anaplasma phagocytophilum* ecotypes in coexisting, discrete enzootic cycles. *Emerg Infect Dis*. 2009;15(12):1948-54.
- Braga M, André MR, Freschi CR, Teixeira MCA, Machado RZ. Molecular and serological detection of *Ehrlichia* spp. in cats on São Luís island, Maranhão, Brazil. *Rev Bras Parasitol Vet*. 2012; 21:37-41.
- Braund KG, editor. *Clinical neurology in small animals - localization, diagnosis and treatment*. Ithaca, NY: International Veterinary Information Service (IVIS); 2003 Feb. *Inflammatory diseases of the central nervous system*. Available at: http://www.ivis.org/special_books/Braund/braund27/ivis.pdf. * Accessed 11 Aug 2004.
- Breitschwerdt E. Ehrlichiosis: New developments [online]. In: *World Small Animal Veterinary Association [WSAVA] World Congress; 2001 Aug 8-11; Vancouver, Canada*. Available at: <http://www.vin.com/VINDBPub/SearchPB/Proceedings/PR05000/PR00143.htm>. Accessed 17 Aug 2004.
- Breitschwerdt EB, Hegarty BC, Hancock SI. Sequential evaluation of dogs naturally infected with *Ehrlichia canis*, *Ehrlichia chaffeensis*, *Ehrlichia equi*, *Ehrlichia ewingii*, or *Bartonella vinsonii*. *J Clin Microbiol*. 1998;36:2645-51.
- Buoro IB, Atwell RB, Kiptoon JC, Ihiga MA. Feline anaemia associated with *Ehrlichia*-like bodies in three domestic short-haired cats. *Vet Rec*. 1989;125:434-6.
- Cabezas-Cruz A, Zweygarth E, Broniszewska M, Passos LM, Ribeiro MF, Manrique M, Tobes R, de la Fuente J. Complete genome sequence of *Ehrlichia mineirensis*, a novel organism closely related to *Ehrlichia canis* with a new host association. *Genome Announc*. 2015;3(1):e01450-14.

Ehrlichiosis and Anaplasmosis

- Cândido SL, de Assis Pereira N, de Oliveira Rosa Fonseca MJ, de Campos Pacheco R, Morgado TO, Colodel EM, Nakazato L, Dutra V, Vieira TSWJ, de Aguiar DM. Molecular detection and genetic characterization of *Ehrlichia canis* and *Ehrlichia* sp. in neotropical primates from Brazil. *Ticks Tick Borne Dis.* 2023;14(4):102179.
- Carrade DD, Foley JE, Borjesson DL, Sykes JE. Canine granulocytic anaplasmosis: a review. *J Vet Intern Med.* 2009;23(6):1129-41.
- Centers for Disease Control and Prevention. *Anaplasma phagocytophilum* transmitted through blood transfusion--Minnesota, 2007. *MMWR Morb Mortal Wkly Rep.* 2008;57(42):1145-8.
- Centers for Disease Control and Prevention [CDC]. Anaplasmosis [online]. CDC; 2012. Available at: <http://www.cdc.gov/anaplasmosis/>. Accessed 20 Jan 2013.
- Centers for Disease Control and Prevention [CDC]. Anaplasmosis. Epidemiology and statistics. CDC; 2024 Jan. Available at: <https://www.cdc.gov/anaplasmosis/stats/index.html#print>. Accessed 1 Feb 2024.
- Centers for Disease Control and Prevention [CDC]. Ehrlichiosis [online]. CDC; 2011. Available at: <http://www.cdc.gov/ehrlichiosis/>. Accessed 20 Jan 2013.
- Centers for Disease Control and Prevention [CDC]. Ehrlichiosis. Epidemiology and statistics. CDC; 2024 Jan. Available at: <https://www.cdc.gov/ehrlichiosis/stats/index.html>. Accessed 1 Feb 2024.
- Chadi H, Moraga-Fernández A, Sánchez-Sánchez M, Chenchouni H, Fernández de Mera IG, Garigliany MM, de la Fuente J, Tennah S, Sedrati T, Ghalmi F. Molecular detection and associated risk factors of *Anaplasma marginale*, *A. ovis* and *A. platys* in sheep from Algeria with evidence of the absence of *A. phagocytophilum*. *Acta Trop.* 2024;249:107040.
- Chien NTH, Nguyen TL, Bui KL, Nguyen TV, Le TH. *Anaplasma marginale* and *A. platys* characterized from dairy and indigenous cattle and dogs in Northern Vietnam. *Korean J Parasitol.* 2019;57(1):43-7.
- Chirek A, Silaghi C, Pfister K, Kohn B. Granulocytic anaplasmosis in 63 dogs: clinical signs, laboratory results, therapy and course of disease. *J Small Anim Pract.* 2018;59(2):112-20.
- Chochlak D, Ioannou I, Tselentis Y, Psaroulaki A. Human anaplasmosis and *Anaplasma ovis* variant. *Emerg Infect Dis.* 2010;16(6):1031-2.
- Christova I, Van De Pol J, Yazar S, Velo E, Schouls L. Identification of *Borrelia burgdorferi* sensu lato, *Anaplasma* and *Ehrlichia* species, and spotted fever group *Rickettsiae* in ticks from Southeastern Europe. *Eur J Clin Microbiol Infect Dis.* 2003;22(9):535-42.
- Cochez C, Ducoffre G, Vandenvelde C, Luyasu V, Heyman P. Human anaplasmosis in Belgium: a 10-year seroepidemiological study. *Ticks Tick Borne Dis.* 2011;2(3):156-9.
- Crosby FL, Wellehan JFX, Pertierra L, Wendland LD, Lundgren AM, Barbet AF, Brown MB. Molecular characterization of "Candidatus *Anaplasma testudinis*": An emerging pathogen in the threatened Florida gopher tortoise (*Gopherus polyphemus*). *Ticks Tick Borne Dis.* 2021;12(3):101672.
- Cruz AC, Zweygarth E, Ribeiro MF, da Silveira JA, de la Fuente J, Grubhoffer L, Valdés JJ, Passos LM. New species of *Ehrlichia* isolated from *Rhipicephalus (Boophilus) microplus* shows an ortholog of the *E. canis* major immunogenic glycoprotein gp36 with a new sequence of tandem repeats. *Parasit Vectors.* 2012;5(1):291.
- Cunha BA. Ehrlichiosis [monograph online]. eMedicine.com; 2011. Available at: <http://emedicine.medscape.com/article/235839-overview>. Accessed 22 Jan 2013.
- Curtis AK, Kleinhenz MD, Anantatat T, Martin MS, Magnin GC, Coetzee JF, Reif KE. Failure to eliminate persistent *Anaplasma marginale* infection from cattle using labeled doses of chlortetracycline and oxytetracycline antimicrobials. *Vet Sci.* 2021;8(11):283.
- Dahlgren FS, Mandel EJ, Krebs JW, Massung RF, McQuiston JH. Increasing incidence of *Ehrlichia chaffeensis* and *Anaplasma phagocytophilum* in the United States, 2000–2007. *Am J Trop Med Hyg.* 2011;85(1):124-31.
- Daniels TJ, Battaly GR, Liveris D, Falco RC, Schwartz I. Avian reservoirs of the agent of human granulocytic ehrlichiosis? *Emerg Infect Dis.* 2002; 8:1524-5.
- Davidson WR, Lockhart JM, Stallknecht DE, Howerth EW. Susceptibility of red and gray foxes to infection by *Ehrlichia chaffeensis*. *J Wildl Dis.* 1999;35:696-702.
- Davidson WR, Lockhart JM, Stallknecht DE, Howerth EW, Dawson JE, Rechav Y. Persistent *Ehrlichia chaffeensis* infection in white-tailed deer. *J Wildl Dis.* 2001;37:538-46.
- Dawson JE, Paddock CD, Warner CK, Greer PW, Bartlett JH, Ewing SA, Munderloh UG, Zaki SR. Tissue diagnosis of *Ehrlichia chaffeensis* in patients with fatal ehrlichiosis by use of immunohistochemistry, *in situ* hybridization, and polymerase chain reaction. *Am J Trop Med Hyg.* 2001;65:603-9.
- Dawson JE, Stallknecht DE, Howerth EW, Warner C, Biggie K, Davidson WR, Lockhart JM, Nettles VF, Olson JG, Childs JE. Susceptibility of white-tailed deer (*Odocoileus virginianus*) to infection with *Ehrlichia chaffeensis*, the etiologic agent of human ehrlichiosis. *J Clin Microbiol.* 1994;32:2725-8.
- Defaye B, Moutailler S, Grech-Angelini S, Galon C, Ferrandi S, Pasqualini V, Quilichini Y. Detecting zoonotic and non-zoonotic pathogens in livestock and their ticks in Corsican wetlands. *Vet Med Sci.* 2022;8(6):2662-77.
- de los Santos JR, Boughan K, Bremer WG, Rizzo B, Schaefer JJ, Rikihisa Y, Needham GR, Capitini LA, Anderson DE, Oglesbee M, Ewing SA, Stich RW. Experimental infection of dairy calves with *Ehrlichia chaffeensis*. *J Med Microbiol.* 2007; 56(Pt 12): 1660-8.
- de los Santos JR, Oglesbee M, Rikihisa Y, Stich RW. Pathological evidence of ehrlichiosis among calves inoculated with *Ehrlichia chaffeensis*. *Ann N Y Acad Sci.* 2008;1149:103-6.
- Dhand A, Nadelman RB, Aguero-Rosenfeld M, Haddad FA, Stokes DP, Horowitz HW. Human granulocytic anaplasmosis during pregnancy: case series and literature review. *Clin Infect Dis.* 2007;45(5):589-93.
- Doudier B, Olano J, Parola P, Brouqui P. Factors contributing to emergence of *Ehrlichia* and *Anaplasma* spp. as human pathogens. *Vet Parasitol.* 2010;167(2-4):149-54.
- Dugan VG, Little SE, Stallknecht DE, Beall AD. Natural infection of domestic goats with *Ehrlichia chaffeensis*. *J Clin Microbiol.* 2000;38:448-9.

Ehrlichiosis and Anaplasmosis

- Dugan VG, Varela AS, Stallknecht DE, Hurd CC, Little SE. Attempted experimental infection of domestic goats with *Ehrlichia chaffeensis*. *Vector Borne Zoonotic Dis*. 2004;4(2):131-6.
- Dugat T, Lagrée AC, Maillard R, Boulouis HJ, Haddad N. Opening the black box of *Anaplasma phagocytophilum* diversity: current situation and future perspectives. *Front Cell Infect Microbiol*. 2015;5:61.
- Dumic I, Jevtic D, Veselinovic M, Nordstrom CW, Jovanovic M, Mogulla V, Veselinovic EM, Hudson A, Simeunovic G, Petcu E, Ramanan P. Human granulocytic anaplasmosis—a systematic review of published cases. *Microorganisms*. 2022;10(7):1433.
- Dumler JS, Madigan JE, Pusterla N, Bakken JS. Ehrlichioses in humans: Epidemiology, clinical presentation, diagnosis, and treatment. *Clin Infect Dis*. 2007;45 Suppl 1:S45-51.
- Eddlestone SM, Neer TM, Gaunt SD, Corstvet R, Gill A, Hosgood G, Hegarty B, Breitschwerdt EB. Failure of imidocarb dipropionate to clear experimentally induced *Ehrlichia canis* infection in dogs. *J Vet Intern Med*. 2006;20(4):840-4.
- Edouard S, Koebel C, Goehringer F, Socolovschi C, Jaulhac B, Raoult D, Brouqui P. Emergence of human granulocytic anaplasmosis in France. *Ticks Tick Borne Dis*. 2012;3(5-6):403-5.
- Eisawi NM, El Hussein ARM, Hassan DA, Musa AB, Hussien MO, Enan KA, Bakheit MA. A molecular prevalence survey on *Anaplasma* infection among domestic ruminants in Khartoum State, Sudan. *Trop Anim Health Prod*. 2020;52(4):1845-52.
- El-Alfy ES, Abbas I, Elseadawy R, Saleh S, Elmishmishy B, El-Sayed SAE, Rizk MA. Global prevalence and species diversity of tick-borne pathogens in buffaloes worldwide: a systematic review and meta-analysis. *Parasit Vectors*. 2023;16(1):115.
- Elhachimi L, Rogiers C, Casaert S, Fellahi S, Van Leeuwen T, Dermauw W, Valcárcel F, Olmeda AS, Daminet S, Khatat SEH, Sahibi H, Duchateau L. Ticks and tick-borne pathogens abundant in the cattle population of the Rabat-Sale Kenitra region, Morocco. *Pathogens*. 2021;10(12):1594.
- El Hamiani Khatat S, Daminet S, Duchateau L, Elhachimi L, Kachani M, Sahibi H. Epidemiological and clinicopathological features of *Anaplasma phagocytophilum* infection in dogs: a systematic review. *Front Vet Sci*. 2021;8:686644.
- Foley JE. Ehrlichiosis, anaplasmosis, and related infections in animals. *The Merck veterinary manual*. Kenilworth, NJ: Merck and Co; 2020. Available at: <https://www.merckvetmanual.com/generalized-conditions/rickettsial-diseases/ehrlichiosis,-anaplasmosis,-and-related-infections-in-animals>. Accessed 10 Jan 2024.
- Foley JE. Equine granulocytic ehrlichiosis. *The Merck veterinary manual*. Kenilworth, NJ: Merck and Co; 2020. Available at: <https://www.merckvetmanual.com/generalized-conditions/equine-granulocytic-anaplasmosis/equine-granulocytic-anaplasmosis>. Accessed 10 Jan 2024.
- Foley J, Nieto NC, Madigan J, Sykes J. Possible differential host tropism in *Anaplasma phagocytophilum* strains in the western United States. *Ann N Y Acad Sci*. 2008;1149:94-7.
- Frankar H, Le Boedec K, Beurlet S, Cauzinille L. Suppurative spinal meningomyelitis in a dog with intra-neutrophilic cerebrospinal fluid cells *Ehrlichia canis morulae*. *Vet Clin Pathol*. 2023;52(1):119-22.
- Franzén P, Berg AL, Aspan A, Gunnarsson A, Pringle J. Death of a horse infected experimentally with *Anaplasma phagocytophilum*. *Vet Rec*. 2007;160(4):122-5.
- Fröhlich J, Fischer S, Bauer B, Hamel D, Kohn B, Ahlers M, Obiegala A, Overzier E, Pfeffer M, Pfister K, Răileanu C, Rehbein S, Skuballa J, Silaghi C. Host-pathogen associations revealed by genotyping of European strains of *Anaplasma phagocytophilum* to describe natural endemic cycles. *Parasit Vectors*. 2023;16(1):289.
- Gajadhar AA, Lobanov V, Scandrett WB, Campbell J, Al-Adhami B. A novel *Ehrlichia* genotype detected in naturally infected cattle in North America. *Vet Parasitol*. 2010;173(3-4):324-9.
- Galay RL, Llaneta CR, Monreal MKFB, Armero AL, Baluyut ABD, et al. Molecular prevalence of *Anaplasma marginale* and *Ehrlichia* in domestic large ruminants and *Rhipicephalus (Boophilus) microplus* ticks from southern Luzon, Philippines. *Front Vet Sci*. 2021;8:746705.
- Galon EM, Macalanda AM, Garcia MM, Ibasco CJ, Garvida A, Ji S, Zafar I, Hasegawa Y, Liu M, Ybañez RH, Umemiya-Shirafuji R, Ybañez A, Claveria F, Xuan X. Molecular identification of selected tick-borne protozoan and bacterial pathogens in Thoroughbred racehorses in Cavite, Philippines. *Pathogens*. 2021;10(10):1318.
- Galon EM, Ybañez RH, Macalanda AM, Estabillo GR, Montano MTR, et al. First molecular identification of Babesia, Theileria, and Anaplasma in goats from the Philippines. *Pathogens*. 2022;11(10):1109.
- Gayle A. Tick-borne diseases. *Am Fam Physician*. 2001;64:461-6.
- Granick JL, Armstrong PJ, Bender JB. *Anaplasma phagocytophilum* infection in dogs: 34 cases (2000-2007). *J Am Vet Med Assoc*. 2009;234(12):1559-65.
- Granquist EG, Bårdsen K, Bergström K, Stuen S. Variant - and individual dependent nature of persistent *Anaplasma phagocytophilum* infection. *Acta Vet Scand*. 2010;52:25.
- Greene CE. Rocky Mountain spotted fever and ehrlichiosis. In: Kirk RW, editor. *Current veterinary therapy IX*. Philadelphia: WB Saunders; 1986. p. 1080-4.
- Gussmann K, Czech C, Hermann M, Schaarschmidt-Kiener D, von Loewenich FD. *Anaplasma phagocytophilum* infection in a horse from Switzerland with severe neurological symptoms. *Schweiz Arch Tierheilkd*. 2014;156(7):345-8.
- Harris RM, Couturier BA, Sample SC, Coulter KS, Casey KK, Schlaberg R. Expanded geographic distribution and clinical characteristics of *Ehrlichia ewingii* infections, United States. *Emerg Infect Dis*. 2016;22:862-5.
- Harrus S, Waner T. Diagnosis of canine monocytotropic ehrlichiosis (*Ehrlichia canis*): an overview. *Vet J*. 2011;187(3):292-6.
- Havens NS, Kinnear BR, Mató S. Fatal ehrlichial myocarditis in a healthy adolescent: a case report and review of the literature. *Clin Infect Dis*. 2012;54(8):e113-4.
- Hegarty BC, Maggi RG, Koskinen P, Beall MJ, Eberts M, Chandrashekar R, Breitschwerdt EB. *Ehrlichia muris* infection in a dog from Minnesota. *J Vet Intern Med* 2012;26:1217-20.
- Heikkilä HM, Bondarenko A, Mihalkov A, Pfister K, Spillmann T. *Anaplasma phagocytophilum* infection in a domestic cat in Finland: Case report. *Acta Vet Scand*. 2010;52:62.

Ehrlichiosis and Anaplasmosis

- Henniger T, Henniger P, Grossmann T, Distl O, Ganter M, von Loewenich FD. Congenital infection with *Anaplasma phagocytophilum* in a calf in northern Germany. *Acta Vet Scand.* 2013;55(1):38.
- Henningsson AJ, Wilhelmsson P, Gyllemark P, Kozak M, Matussek A, Nyman D, Ekerfelt C, Lindgren PE, Forsberg P. Low risk of seroconversion or clinical disease in humans after a bite by an *Anaplasma phagocytophilum*-infected tick. *Ticks Tick Borne Dis.* 2015;6(6):787-92.
- Hornok S, Boldogh SA, Takács N, Juhász A, Kontschán J, Földi D, Koleszár B, Morandini P, Gyuranecz M, Szekeres S. Anaplasmataceae closely related to *Ehrlichia chaffeensis* and *Neorickettsia helminthoeca* from birds in Central Europe, Hungary. *Antonie Van Leeuwenhoek.* 2020;113(7):1067-73.
- Hornok S, Boldogh SA, Takács N, Sándor AD, Tuska-Szalay B. Zoonotic ecotype-I of *Anaplasma phagocytophilum* in sympatric wildcat, pine marten and red squirrel - Short communication. *Acta Vet Hung.* 2022. doi: 10.1556/004.2022.00021 [Online ahead of print].
- Hove P, Khumalo ZTH, Chaisi ME, Oosthuizen MC, Brayton KA, Collins NE. Detection and characterisation of *Anaplasma marginale* and *A. centrale* in South Africa. *Vet Sci.* 2018;5(1):26.
- Hu H, Liu Z, Fu R, Liu Y, Ma H, Zheng W. Detection and phylogenetic analysis of tick-borne bacterial and protozoan pathogens in a forest province of eastern China. *Acta Trop.* 2022;235:106634.
- Ingram D, Joseph B, Hawkins S, Spain J. Anaplasmosis in Pennsylvania: clinical features, diagnosis, and outcomes of patients diagnosed with *Anaplasma phagocytophilum* infection at Hershey Medical Center from 2008 to 2021. *Open Forum Infect Dis.* 2023;10(4):ofad193.
- Ismail N, Bloch KC, McBride JW. Human ehrlichiosis and anaplasmosis. *Clin Lab Med.* 2010;30(1):261-92.
- Ismail N, McBride JW. Tick-borne emerging infections: ehrlichiosis and anaplasmosis. *Clin Lab Med.* 2017;37:317-40.
- Jablinski AC, Reppert EJ, Huser S, Robért BD, Jaeger JR, Kang Q, Liu R, Anantatat T, Armstrong CL, Reif KE. Kansas beef bulls with chronic anaplasmosis demonstrate satisfactory breeding soundness outcomes at breeding soundness examination. *Am Vet Med Assoc.* 2023;262(1):53-60.
- Jenkins S, Ketzis JK, Dundas J, Scorpio D. Efficacy of minocycline in naturally occurring nonacute *Ehrlichia canis* infection in dogs. *J Vet Intern Med.* 2018;32(1):217-21.
- Jereb M, Pecaver B, Tomazic J, Muzlovic I, Avsic-Zupanc T, Premru-Srsen T, Levicnik-Stezinar S, Karner P, Strle F. Severe human granulocytic anaplasmosis transmitted by blood transfusion. *Emerg Infect Dis.* 2012;18(8):1354-7.
- Jongejan F, Crafford D, Erasmus H, Fourie JJ, Schunack B. Comparative efficacy of oral administered afoxolaner (NexGard™) and fluralaner (Bravecto™) with topically applied permethrin/imidacloprid (Advantix®) against transmission of *Ehrlichia canis* by infected *Rhipicephalus sanguineus* ticks to dogs. *Parasit Vectors.* 20;9(1):348.
- Jouglin M, Blanc B, de la Cotte N, Bastian S, Ortiz K, Malandrin L. First detection and molecular identification of the zoonotic *Anaplasma capra* in deer in France. *PLoS One.* 2019;14(7):e0219184.
- Jouglin M, Rispé C, Grech-Angelini S, Gallois M, Malandrin L. *Anaplasma capra* in sheep and goats on Corsica Island, France: A European lineage within *A. capra* clade II? *Ticks Tick Borne Dis.* 2022;13(3):101934.
- Kaewmongkol G, Maneesaay P, Suwanna N, Tiraphut B, Krajarngjang T, Chouybumrung A, Kaewmongkol S, Sirinarumit T, Jittapalpong S, Fenwick SG. First detection of *Ehrlichia canis* in cerebrospinal fluid from a nonthrombocytopenic dog with meningoencephalitis by broad-range PCR. *J Vet Intern Med.* 2016;30(1):255-9.
- Karpathy SE, Kingry L, Pritt BS, Berry JC, Chilton NB, Dergousoff SJ, Cortinas R, Sheldon SW, Oatman S, Anacker M, Petersen J, Paddock CD. *Anaplasma bovis*-like infections in humans, United States, 2015–2017. *Emerg Infect Dis.* 2023; 29:1904-7.
- Keesing F, Hersh MH, Tibbetts M, McHenry DJ, Duerr S, Brunner J, Killilea M, Logiudice K, Schmidt KA, Ostfeld RS. Reservoir competence of vertebrate hosts for *Anaplasma phagocytophilum*. *Emerg Infect Dis.* 2012;18(12):2013-6.
- Killmaster LF, Levin ML. Isolation and short-term persistence of *Ehrlichia ewingii* in cell culture. *Vector Borne Zoonotic Dis.* 2016;16(7):445-8.
- Kingry L, Sheldon S, Oatman S, Pritt B, Anacker M, et al. Targeted metagenomics for clinical detection and discovery of bacterial tick-borne pathogens. *J Clin Microbiol.* 2020;58:e00147-20.
- Kocan KM, de la Fuente J, Blouin EF, Coetzee JF, Ewing SA. The natural history of *Anaplasma marginale*. *Vet Parasitol.* 2010;167(2-4):95-107.
- Kocan KM, de la Fuente J, Cabezas-Cruz A. The genus *Anaplasma*: new challenges after reclassification. *Rev Sci Tech.* 2015;34(2):577-86.
- Kocan AA, Levesque GC, Whitworth LC, Murphy GL, Ewing SA, Barker RW. Naturally occurring *Ehrlichia chaffeensis* infection in coyotes from Oklahoma. *Emerg Infect Dis.* 2000;6:477-80.
- Koh FX, Kho KL, Panchadcharam C, Sitam FT, Tay ST. Molecular detection of *Anaplasma* spp. in pangolins (*Manis javanica*) and wild boars (*Sus scrofa*) in Peninsular Malaysia. *Vet Parasitol.* 2016;227:73-6.
- Koh FX, Panchadcharam C, Sitam FT, Tay ST. Molecular investigation of *Anaplasma* spp. in domestic and wildlife animals in Peninsular Malaysia. *Vet Parasitol Reg Stud Reports.* 2018;13:141-7.
- Kohn B, Galke D, Beelitz P, Pfister K. Clinical features of canine granulocytic anaplasmosis in 18 naturally infected dogs. *J Vet Intern Med.* 2008;22(6):1289-95.
- Koku R, Herndon DR, Avillan J, Morrison J, Futse JE, Palmer GH, Brayton KA, Noh SM. Both coinfection and superinfection drive complex *Anaplasma marginale* strain structure in a natural transmission setting. *Infect Immun.* 2021;89(11):e0016621.
- Kruppenbacher AS, Müller E, Aardema ML, Schäfer I, von Loewenich FD. Granulocytic anaplasmosis in cats from central Europe and molecular characterization of feline *Anaplasma phagocytophilum* strains by ankA gene, groEL gene and multilocus sequence typing. *Parasit Vectors.* 2023;16(1):348.

- Lagré AC, Rouxel C, Kevin M, Dugat T, Girault G, Durand B, Pfeffer M, Silaghi C, Nieder M, Boulouis HJ, Haddad N. Co-circulation of different *A. phagocytophilum* variants within cattle herds and possible reservoir role for cattle. *Parasit Vectors*. 2018;11(1):163.
- Latrofa MS, Dantas-Torres F, de Caprariis D, Cantacessi C, Capelli G, Lia RP, Breitschwerdt EB, Otranto D. Vertical transmission of *Anaplasma platys* and *Leishmania infantum* in dogs during the first half of gestation. *Parasit Vectors*. 2016;9(1):269.
- Lepidi H, Bunnell JE, Martin ME, Madigan JE, Stuen S, Dumler JS. Comparative pathology and immunohistology associated with clinical illness after *Ehrlichia phagocytophila*-group infections. *Am J Trop Med Hyg*. 2000;62:29-37.
- Leschnik M, Kirtz G, Virányi Z, Wille-Piazzai W, Duscher G. Acute granulocytic anaplasmosis in a captive timber wolf (*Canis lupus occidentalis*). *J Zoo Wildl Med*. 2012;43(3):645-8.
- Lesiczka PM, Hrazdilová K, Majerová K, Fonville M, Sprong H, Hönig V, Hofmannová L, Papežik P, Růžek D, Zurek L, Votýpka J, Modrý D. The role of peridomestic animals in the eco-epidemiology of *Anaplasma phagocytophilum*. *Microb Ecol*. 2021;82(3):602-12.
- Levin ML, Troughton DR, Loftis AD. Duration of tick attachment necessary for transmission of *Anaplasma phagocytophilum* by *Ixodes scapularis* (Acari: Ixodidae) nymphs. *Ticks Tick Borne Dis*. 2021;12(6):101819.
- Li H, Zheng YC, Ma L, Jia N, Jiang BG, et al. Human infection with a novel tick-borne *Anaplasma* species in China: a surveillance study. *Lancet Infect Dis*. 2015;15(6):663-70.
- Li Y, Chen Z, Liu ZJ, Yang J, Li Q, Li Y, Luo J, Yi H. Molecular survey of *Anaplasma* and *Ehrlichia* of red deer and sika deer in Gansu, China in 2013. *Trans. Emerg. Dis*. 2016;63(6):e228-36.
- Li Y, Yang J, Chen Z, Qin G, Li Y, Li Q, Liu J, Liu Z, Guan G, Yin H, Luo J, Zhang L. *Anaplasma* infection of Bactrian camels (*Camelus bactrianus*) and ticks in Xinjiang, China. *Parasit Vectors*. 2015;8:313.
- Little SE. Ehrlichiosis and anaplasmosis in dogs and cats. *Vet Clin North Am Small Anim Pract*. 2010;40(6):1121-40.
- Loftis AD, Kelly PJ, Paddock CD, Blount K, Johnson JW, Gleim ER, Yabsley MJ, Levin ML, Beati L. Panola Mountain *Ehrlichia* in *Amblyomma maculatum* from the United States and *Amblyomma variegatum* (Acari: Ixodidae) from the Caribbean and Africa. *J Med Entomol*. 2016;53(3):696-8.
- Loftis AD, Levin ML, Spurlock JP. Two USA *Ehrlichia* spp. cause febrile illness in goats. *Vet Microbiol*. 2008;130:398-402.
- Loftis AD, Reeves WK, Spurlock JP, Mahan SM, Troughton DR, Dasch GA, Levin ML. Infection of a goat with a tick-transmitted *Ehrlichia* from Georgia, U.S.A., that is closely related to *Ehrlichia ruminantium*. *J Vector Ecol*. 2006;31(2):213-23.
- Lorusso V, Wijnveld M, Latrofa MS, Fajinmi A, Majekodunmi AO, Dogo AG, Igweh AC, Otranto D, Jongejan F, Welburn SC, Picozzi K. Canine and ovine tick-borne pathogens in camels, Nigeria. *Vet Parasitol*. 2016;228:90-2.
- Lovett AC, Reppert EJ, Jaeger JR, Kang Q, Flowers MR, Bickmeier NP, Anantatit T, O'Day SC, Armstrong CL, Reif KE. Satisfactory breeding potential is transiently eliminated in beef bulls with clinical anaplasmosis. *BMC Vet Res*. 2022;18(1):381.
- Lu M, Chen Q, Qin X, Lyu Y, Teng Z, et al. *Anaplasma bovis* infection in fever and thrombocytopenia patients—Anhui Province, China 2021. *China CDC Wkly*. 2022;4:249-53.
- Lu M, Li F, Liao Y, Shen JJ, Xu JM, Chen YZ, Li JH, Holmes EC, Zhang JZ. Epidemiology and diversity of rickettsiales bacteria in humans and animals in Jiangsu and Jiangxi provinces, China. *Sci Rep*. 2019;9(1):13176.
- Lukács RM, Peters IR, Eminaga S, Buckeridge DM. *Ehrlichia canis* infection in the cerebrospinal fluid of a dog characterized by morulae within monocytes and neutrophils. *Vet Clin Pathol*. 2020;49(3):470-5.
- Machado RZ, André MR, Werther K, de Sousa E, Gavioli FA, Alves Junior JR. Migratory and carnivorous birds in Brazil: reservoirs for *Anaplasma* and *Ehrlichia* species? *Vector Borne Zoonotic Dis*. 2012;12(8):705-8.
- Machado RZ, Teixeira MM, Rodrigues AC, André MR, Gonçalves LR, Barbosa da Silva J, Pereira CL. Molecular diagnosis and genetic diversity of tick-borne Anaplasmataceae agents infecting the African buffalo *Syncerus caffer* from Marromeu Reserve in Mozambique. *Parasit Vectors*. 2016;9:454.
- MacQueen D, Centellas F. Human granulocytic anaplasmosis. *Infect Dis Clin North Am*. 2022;36(3):639-54.
- Madison-Antenucci S, Kramer LD, Gebhardt LL, Kauffman E. Emerging tick-borne diseases. *Clin Microbiol Rev*. 2020;33(2):e00083-18.
- Mafra C, Barcelos RM, Mantovani C, Carrizo J, Soares AC, Moreira HN, Maia NL, da Silva Fde F, e Silva VH, Boere V, e Silva Ide O. Occurrence of *Ehrlichia canis* in free-living primates of the genus *Callithrix*. *Rev Bras Parasitol Vet*. 2015;24(1):78-81.
- Maggi RG, Mascarelli PE, Havenga LN, Naidoo V, Breitschwerdt EB. Co-infection with *Anaplasma platys*, *Bartonella henselae* and *Candidatus Mycoplasma haematoparvum* in a veterinarian. *Parasit Vectors*. 2013;6:103-13.
- Mah A, Viola GM, Ariza Heredia E, Rezvani K, Kebriaei P, Bhatti MM, Han X, Shpall EJ, Mulanovich VE. Graft loss attributed to possible transfusion-transmitted ehrlichiosis following cord blood stem cell transplant. *Transpl Infect Dis*. 2018;20:e12899.
- Marwaha S, Ranjan R, Nath K, Singh M, Sawal RK, Sahoo A. Molecular epidemiology of anaplasmosis in Indian dromedary camels. *Vet Res Commun*. 2024. doi: 10.1007/s11259-024-10373-5. [Online ahead of print].
- Masika SJ, Muchemi GM, Okumu TA, Mutura S, Zimmerman D, Kamau J. Molecular evidence of *Anaplasma phagocytophilum* in olive baboons and vervet monkeys in Kenya. *BMC Vet Res*. 2021;17(1):385.
- Matei IA, Estrada-Peña A, Cutler SJ, Vayssier-Taussat M, Varela-Castro L, Potkonjak A, Zeller H, Mihalca AD. A review on the eco-epidemiology and clinical management of human granulocytic anaplasmosis and its agent in Europe. *Parasit Vectors*. 2019;12(1):599.
- Matei IA, Stuen S, Modrý D, Degan A, D'Amico G, Mihalca AD. Neonatal *Anaplasma platys* infection in puppies: Further evidence for possible vertical transmission. *Vet J*. 2017;219:40-1.
- Mazzucco Panizza M, Cutullé C, Primo ME, Morel N, Sebastian PS, Nava S. Assays to evaluate the transovarial transmission of *Anaplasma marginale* by *Rhipicephalus microplus*. *Vet Parasitol*. 2022;311:109808.

- Meichner K, Qurollo BA, Anderson KL, Grindem CB, Savage M, Breitschwerdt EB. Naturally occurring *Ehrlichia ewingii* and *Mycoplasma* sp. co-infection in a goat. *J Vet Intern Med.* 2015;29(6):1735-8.
- Modarelli JJ, Borst MM, Piccione J, Esteve-Gasent MD. Molecular identification of *Ehrlichia ewingii* in a polyarthritic Texas dog. *Vet Clin Pathol.* 2019;48(1):96-9.
- Moura de Aguiar D, Pessoa Araújo Junior J, Nakazato L, Bard E, Aguiar-Bultet L, Vorimore F, Leonidovich Popov V, Moleta Colodel E, Cabezas-Cruz A. Isolation and characterization of a novel pathogenic strain of *Ehrlichia minasensis*. *Microorganisms.* 2019;7(11):528.
- Mukhacheva TA, Shaikhova DR, Kovalev SY, von Loewenich FD. Phylogeographical diversity of *Anaplasma phagocytophilum* in the Asian part of Russia based on multilocus sequence typing and analysis of the ankA gene. *Infect Genet Evol.* 2020;80:104234.
- Mukhtar MU, Iqbal N, Yang J, Nawaz Z, Peng TL. The first molecular identification and phylogenetic analysis of tick-borne pathogens in captive wild animals from Lohi Bher zoo, Pakistan. *Parasitol Res.* 2022;121(11):3321-6.
- Muraro LS, Souza AO, Leite TNS, Cândido SL, Melo ALT, Toma HS, Carvalho MB, Dutra V, Nakazato L, Cabezas-Cruz A, Aguiar DM. First evidence of *Ehrlichia minasensis* infection in horses from Brazil. *Pathogens.* 2021;10(3):265.
- Mwale R, Mulavu M, Khumalo CS, Mukubesa A, Nalubamba K, Mubemba B, Changula K, Simulundu E, Chitanga S, Namangala B, Mataa L, Zulu VC, Munyeme M, Muleya W. Molecular detection and characterization of *Anaplasma* spp. in cattle and sable antelope from Lusaka and North-Western provinces of Zambia. *Vet Parasitol Reg Stud Reports.* 2023;39:100847.
- Mwamuye MM, Kariuki E, Omondi D, Kabii J, Odongo D, Masiga D, Villingier J. Novel *Rickettsia* and emergent tick-borne pathogens: A molecular survey of ticks and tick-borne pathogens in Shimba Hills National Reserve, Kenya. *Ticks Tick Borne Dis.* 2017;8(2):208-18.
- Mylonakis ME, Harrus S, Breitschwerdt EB. An update on the treatment of canine monocytic ehrlichiosis (*Ehrlichia canis*). *Vet J.* 2019;246:45-53.
- Mylonakis ME, Kritsepi-Konstantinou M, Dumler JS, Diniz PP, Day MJ, Siarkou VI, Breitschwerdt EB, Psychas V, Petanides T, Koutinas AF. Severe hepatitis associated with acute *Ehrlichia canis* infection in a dog. *J Vet Intern Med.* 2010;24(3):633-8.
- Nair AD, Cheng C, Ganta CK, Sanderson MW, Alleman AR, Munderloh UG, Ganta RR. Comparative experimental infection study in dogs with *Ehrlichia canis*, *E. chaffeensis*, *Anaplasma platys* and *A. phagocytophilum*. *PLoS One.* 2016;11(2):e0148239.
- Nair AD, Cheng C, Jaworski DC, Willard LH, Sanderson MW, Ganta RR. *Ehrlichia chaffeensis* infection in the reservoir host (white-tailed deer) and in an incidental host (dog) is impacted by its prior growth in macrophage and tick cell environments. *PLoS One.* 2014;9(10):e109056.
- Nakayima J, Hayashida K, Nakao R, Ishii A, Ogawa H, Nakamura I, Moonga L, Hang'ombe BM, Mweene AS, Thomas Y, Orba Y, Sawa H, Sugimoto C. Detection and characterization of zoonotic pathogens of free-ranging non-human primates from Zambia. *Parasit Vectors.* 2014;7:490.
- Nawaz M, Ullah R, Rehman ZU, Naeem M, Khan A, Bourhia M, Sohail MM, Ali T, Khan A, Hussain T, Iqbal F. Leading report of molecular prevalence of tick borne *Anaplasma marginale* and *Theileria ovis* in yaks (*Bos grunniens*) from Pakistan. *Arch Microbiol.* 2024;206(4):149.
- Ndip LM, Ndip RN, Esemu SN, Dickmu VL, Fokam EB, Walker DH, McBride JW. Ehrlichial infection in Cameroonian canines by *Ehrlichia canis* and *Ehrlichia ewingii*. *Vet Microbiol.* 2005;111(1-2):59-66.
- Nichols Heitman K, Dahlgren FS, Drexler NA, Massung RF, Behravesh CB. Increasing incidence of ehrlichiosis in the United States: a summary of national surveillance of *Ehrlichia chaffeensis* and *Ehrlichia ewingii* infections in the United States, 2008-2012. *Am J Trop Med Hyg.* 2016;94(1):52-60.
- Noaman V, Sazmand A. *Anaplasma ovis* infection in sheep from Iran: molecular prevalence, associated risk factors, and spatial clustering. *Trop Anim Health Prod.* 2021;54(1):6.
- Oliveira LS, Oliveira KA, Mourão LC, Pescatore AM, Almeida MR, Conceição LG, Galvão MA, Mafra C. First report of *Ehrlichia ewingii* detected by molecular investigation in dogs from Brazil. *Clin Microbiol Infect.* 2009;15 Suppl 2:55-6.
- Oliver A, Conrado FO, Nolen-Walston R. Equine granulocytic anaplasmosis. *Vet Clin North Am Equine Pract.* 2023;39(1):133-45.
- Orozco MM, Argibay HD, Minatel L, Guillemi EC, Berra Y, Schapira A, Di Nucci D, Marcos A, Lois F, Falzone M, Farber MD. A participatory surveillance of marsh deer (*Blastocercus dichotomus*) morbidity and mortality in Argentina: first results. *BMC Vet Res.* 2020;16(1):321.
- Paddock CD, Childs JE. *Ehrlichia chaffeensis*: a prototypical emerging pathogen. *Clin Microbiol Rev.* 2003; 16(1): 37-64.
- Pantchev N, Pluta S, Huisinga E, Nather S, Scheufelen M, Vrhovec MG, Schweinitz A, Hampel H, Straubinger RK. Tick-borne diseases (borreliosis, anaplasmosis, babesiosis) in German and Austrian dogs: status quo and review of distribution, transmission, clinical findings, diagnostics and prophylaxis. *Parasitol Res.* 2015;114 Suppl 1:S19-54.
- Parvizi O, Akinyemi KO, Roesler U, Neubauer H, Mertens-Scholz K. Retrospective study of anaplasmosis in countries of North Africa and the Middle East. *Rev Sci Tech.* 2020;39(3):1053-68.
- Peng SH, Yang SL, Ho YN, Chen HF, Shu PY. Human case of *Ehrlichia chaffeensis* infection, Taiwan. *Emerg Infect Dis.* 2019;25(11):2141-3.
- Peng Y, Lu C, Yan Y, Song J, Pei Z, Gong P, Wang R, Zhang L, Jian F, Ning C. The novel zoonotic pathogen, *Anaplasma capra*, infects human erythrocytes, HL-60, and TF-1 cells *in vitro*. *Pathogens.* 2021;10(5):600.
- Pennisi MG, Hofmann-Lehmann R, Radford AD, Tasker S, Belák S, et al. *Anaplasma*, *Ehrlichia* and *Rickettsia* species infections in cats: European guidelines from the ABCD on prevention and management. *J Feline Med Surg.* 2017;19(5):542-8.
- Perez M, Bodor M, Zhang C, Xiong Q, Rikihisa Y. Human infection with *Ehrlichia canis* accompanied by clinical signs in Venezuela. *Ann N Y Acad Sci.* 2006;1078:110-7.
- Perez M, Rikihisa Y, Wen B. *Ehrlichia canis*-like agent isolated from a man in Venezuela: antigenic and genetic characterization. *J Clin Microbiol.* 1996;34:2133-9.

Ehrlichiosis and Anaplasmosis

- Pinhanelli VC, Costa PN, Silva G, Aguiar DM, Silva CM, Fachin AL, Marins M. Development and evaluation of a loop-mediated isothermal amplification assay for detection of *Ehrlichia canis* DNA in naturally infected dogs using the p30 gene. *Genet Mol Res.* 2015;14(4):17885-92.
- Pritt BS, Sloan LM, Johnson DK, et al. Emergence of a new pathogenic *Ehrlichia* species, Wisconsin and Minnesota, 2009. *N Engl J Med.* 2011;365(5):422-9.
- Pusterla N, Huder J, Wolfensberger C, Braun U, Lutz H. Laboratory findings in cows after experimental infection with *Ehrlichia phagocytophila*. *Clin Diagn Lab Immunol.* 1997;4:643-7.
- Qiu H, Kelly PJ, Zhang J, Luo Q, Yang Y, Mao Y, Yang Z, Li J, Wu H, Wang C. Molecular detection of *Anaplasma* spp. and *Ehrlichia* spp. in ruminants from twelve provinces of China. *Can J Infect Dis Med Microbiol.* 2016;2016:9183861.
- Quorollo BA, Buch J, Chandrashekar R, Beall MJ, Breitschwerdt EB, Yancey CB, Caudill AH, Comyn A. Clinicopathological findings in 41 dogs (2008-2018) naturally infected with *Ehrlichia ewingii*. *J Vet Intern Med.* 2019;33(2):618-29.
- Quorollo BA, Davenport AC, Sherbert BM, Grindem CB, Birkenheuer AJ, Breitschwerdt EB. Infection with Panola Mountain *Ehrlichia* sp. in a dog with atypical lymphocytes and clonal T-cell expansion. *Vet Intern Med.* 2013;27(5):1251-5.
- Rahamim M, Harrus S, Nachum-Biala Y, Baneth G, Aroch I. *Ehrlichia canis* morulae in peripheral blood lymphocytes of two naturally-infected puppies in Israel. *Vet Parasitol Reg Stud Reports.* 2021;24:100554.
- Ramírez-Hernández A, Arroyave E, Faccini-Martínez AA, Martínez-Díaz HC, Betancourt-Ruiz P, Olaya-M LA, Forero-Becerra EG, Hidalgo M, Blanton LS, Walker DH. Emerging tickborne bacteria in cattle from Colombia. *Emerg Infect Dis.* 2022;28(10):2109-21.
- Rar V, Tkachev S, Tikunova N. Genetic diversity of *Anaplasma* bacteria: Twenty years later. *Infect Genet Evol.* 2021;91:104833.
- Rassouli M, Ardekani AO, Mojaver MJ, Roozbeh M, Beikha M, Sani SER. Molecular detection of *Anaplasma platys* among camels (*Camelus dromedarius*) in Yazd, Iran. *Vet Parasitol Reg Stud Reports.* 2020:100462.
- Reeves WK, Loftis AD, Nicholson WL, Czarkowski AG. The first report of human illness associated with the Panola Mountain *Ehrlichia* species: a case report. *J Med Case Rep.* 2008;2:139.
- Regan J, Matthias J, Green-Murphy A, Stanek D, Bertholf M, Pritt BS, Sloan LM, Kelly AJ, Singleton J, McQuiston JH, Hocevar SN, Whittle JP. A confirmed *Ehrlichia ewingii* infection likely acquired through platelet transfusion. *Clin Infect Dis.* 2013;56(12):e105-7.
- Rejmanek D, Bradburd G, Foley J. Molecular characterization reveals distinct genospecies of *Anaplasma phagocytophilum* from diverse North American hosts. *J Med Microbiol.* 2012;61(Pt 2):204-12.
- Remesar S, Castro-Scholten S, Morrondo P, Díaz P, Jiménez-Martín D, Muñoz-Fernández L, Fajardo T, Cano-Terriza D, García-Bocanegra I. Occurrence of spp. in wild lagomorphs from southern Spain: Molecular detection of new *Anaplasma bovis* lineages. *Res Vet Sci.* 2024;166:105093.
- Remesar S, Prieto A, García-Dios D, López-Lorenzo G, Martínez-Calabuig N, Díaz-Cao JM, Panadero R, López CM, Fernández G, Díez-Baños P, Morrondo P, Díaz P. Diversity of *Anaplasma* species and importance of mixed infections in roe deer from Spain. *Transbound Emerg Dis.* 2022;69(4):e374-85.
- Reppert E., Galindo R., Breshears M., Kocan K., Blouin E., de la Fuente J. Demonstration of transplacental transmission of a human isolate of *Anaplasma phagocytophilum* in an experimentally infected sheep. *Trans Emerg Dis.* 2013;60:93-6.
- Richardson SS, Mainville CA, Arguello-Marin A, Whalley D, Burton W, Breitschwerdt EB, Quorollo BA. A second-generation, point-of-care immunoassay provided improved detection of *Anaplasma* and *Ehrlichia* antibodies in PCR-positive dogs naturally infected with *Anaplasma* or *Ehrlichia* species. *J Vet Diagn Invest.* 2023;35(4):366-3.
- Rikihisa Y. The tribe Ehrlichieae and ehrlichial diseases. *Clin Microbiol Rev.* 1991;4:286-308.
- Rodrigues GD, Lucas M, Ortiz HG, Dos Santos Gonçalves L, Blodorn E, Domingues WB, Nunes LS, Saravia A, Parodi P, Riet-Correa F, Menchaca A, Campos VF, Krolow TK, Krüger RF. Molecular of *Anaplasma marginale* Theiler (Rickettsiales: Anaplasmataceae) in horseflies (Diptera: Tabanidae) in Uruguay. *Sci Rep.* 2022;12(1):22460.
- Rojas N, Castillo D, Marin P. Molecular detection of *Ehrlichia chaffeensis* in humans, Costa Rica. *Emerg Infect Dis.* 2015;21(3):532-4.
- Rymaszewska A, Grenda S. Bacteria of the genus *Anaplasma* – characteristics of *Anaplasma* and their vectors: a review. *Vet Med (Praha).* 2008;53(11): 573-84.
- Sachdev SH, Joshi V, Cox ER, Amoroso A, Palekar S. Severe life-threatening *Ehrlichia chaffeensis* infections transmitted through solid organ transplantation. *Transpl Infect Dis.* 2014;16(1):119-24.
- Sahin OF, Erol U, Altay K. Buffaloes as new hosts for *Anaplasma capra*: Molecular prevalence and phylogeny based on gtlA, groEL, and 16S rRNA genes. *Res Vet Sci.* 2022;152:458-64.
- Sahin OF, Erol U, Duzlu O, Altay K. Molecular survey of *Anaplasma phagocytophilum* and related variants in water buffaloes: The first detection of *Anaplasma phagocytophilum*-like 1. *Comp Immunol Microbiol Infect Dis.* 2023;98:102004.
- Samlaska CP. Arthropod infestations and vectors of disease [monograph online]. In: James WD, editor. *Military dermatology, Part III: Disease and the environment.* Available at: http://www.wramc.amedd.army.mil/fieldmed/dermatology/Derm_Textbook_Ch9.pdf. * Accessed 18 Aug 2004.
- Santoro M, Veneziano V, D'Alessio N, Di Prisco F, Lucibelli MG, Borriello G, Cerrone A, Dantas-Torres F, Latrofa MS, Otranto D, Galiero G. Molecular survey of *Ehrlichia canis* and *Coxiella burnetii* infections in wild mammals of southern Italy. *Parasitol Res.* 2016;115(11):4427-31.
- Sato M, Veir JK, Shropshire SB, Lappin MR. *Ehrlichia canis* in dogs experimentally infected, treated, and then immune suppressed during the acute or subclinical phases. *J Vet Intern Med.* 2020;34(3):1214-21.
- Schäfer I, Kohn B. *Anaplasma phagocytophilum* infection in cats: A literature review to raise clinical awareness. *J Feline Med Surg.* 2020;22(5):428-41.

Ehrlichiosis and Anaplasmosis

- Schäfer I, Kohn B, Müller E. *Anaplasma phagocytophilum* in domestic cats from Germany, Austria and Switzerland and clinical/laboratory findings in 18 PCR-positive cats (2008-2020). *J Feline Med Surg*. 2022;24(4):290-7.
- Scharf W, Schauer S, Freyburger F, Petrovec M, Schaarschmidt-Kiener D, et al. Distinct host species correlate with *Anaplasma phagocytophilum* anka gene clusters. *J Clin Microbiol*. 2011;49(3):790-6.
- Schotthoefer AM, Meece JK, Fritsche TR. A clinical, diagnostic, and ecologic perspective on human anaplasmosis in the Upper Midwest. *WMJ*. 2014;113(3):107-14.
- Schvartz G, Epp T, Burgess HJ, Chilton NB, Lohmann KL. Comparison between available serologic tests for detecting antibodies against *Anaplasma phagocytophilum* and *Borrelia burgdorferi* in horses in Canada. *Vet Diagn Invest*. 2015;27(4):540-6.
- Selmi R, Ben Said M, Dhibi M, Ben Yahia H, Messadi L. Improving specific detection and updating phylogenetic data related to *Anaplasma platys*-like strains infecting camels (*Camelus dromedarius*) and their ticks. *Ticks Tick Borne Dis*. 2019;10(6):101260.
- Seo MG, Ouh IO, Kwak D. Detection and genotypic analysis of *Anaplasma bovis* and *A. phagocytophilum* in horse blood and lung tissue. *Int J Mol Sci*. 2023;24(4):3239.
- Seo MG, Ouh IO, Lee H, Geraldino PJJ, Rhee MH, Kwon OD, Kwak D. Differential identification of *Anaplasma* in cattle and potential of cattle to serve as reservoirs of *Anaplasma capra*, an emerging tick-borne zoonotic pathogen. *Vet Microbiol*. 2018;226:15-22.
- Shi K, Li J, Yan Y, Chen Q, Wang K, Zhou Y, Li D, Chen Y, Yu F, Peng Y, Zhang L, Ning C. Dogs as new hosts for the emerging zoonotic pathogen *Anaplasma capra* in China. *Front Cell Infect Microbiol*. 2019;9:394.
- Shin SU, Park YJ, Ryu JH, Jang DH, Hwang S, Cho HC, Park J, Han JI, Choi KS. Identification of zoonotic tick-borne pathogens from Korean water deer (*Hydropotes inermis argyropus*). *Vector Borne Zoonotic Dis*. 2020;20(10):745-54.
- Silaghi C, Liebisch G, Pfister K. Genetic variants of *Anaplasma phagocytophilum* from 14 equine granulocytic anaplasmosis cases. *Parasit Vectors*. 2011;4:161.
- Silaghi C, Santos AS, Gomes J, Christova I, Matei IA, Walder G, Domingos A, Bell-Sakyi L, Sprong H, von Loewenich FD, Oteo JA, de la Fuente J, Dumler JS. Guidelines for the direct detection of *Anaplasma* spp. in diagnosis and epidemiological studies. *Vector Borne Zoonotic Dis*. 2017;17(1):12-22.
- Silaghi C, Scheuerle MC, Friche Passos LM, Thiel C, Pfister K. PCR detection of *Anaplasma phagocytophilum* in goat flocks in an area endemic for tick-borne fever in Switzerland. *Parasite*. 2011;18(1):57-62.
- Sim RR, Joyner PH, Padilla LR, Anikis P, Aitken-Palmer C. Clinical disease associated with *Anaplasma phagocytophilum* infection in captive Przewalski's horses (*Equus ferus przewalskii*). *J Zoo Wildl Med*. 2017;48(2):497-505.
- Skoracki M, Michalik J, Skotarczak B, Rymaszewska A, Sikora B, Hofman T, Wodecka B, Sawczuk M. First detection of *Anaplasma phagocytophilum* in quill mites (Acari: Symbiophilidae) parasitizing passerine birds. *Microbes Infect*. 2006;8:303-7.
- Skotarczak B. Canine ehrlichiosis. *Ann Agric Environ Med*. 2003;10:137-41.
- Staji H, Yousefi M, Hamedani MA, Tamai IA, Khaligh SG. Genetic characterization and phylogenetic of *Anaplasma capra* in Persian onagers (*Equus hemionus onager*). *Vet Microbiol*. 2021;261:109199.
- Starkey LA, Barrett AW, Beall MJ, Chandrashekar R, Thatcher B, Tyrrell P, Little SE. Persistent *Ehrlichia ewingii* infection in dogs after natural tick infestation. *J Vet Intern Med*. 2015;29(2):552-5.
- Starkey LA, Barrett AW, Chandrashekar R, Stillman BA, Tyrrell P, Thatcher B, Beall MJ, Gruntmeir JM, Meinkoth JH, Little SE. Development of antibodies to and PCR detection of *Ehrlichia* spp. in dogs following natural tick exposure. *Vet Microbiol*. 2014;173(3-4):379-84.
- Stuen S. *Anaplasma phagocytophilum* - the most widespread tick-borne infection in animals in Europe. *Vet Res Commun*. 2007Suppl 1:79-84.
- Stuen S, Grøva L, Granquist EG, Sandstedt K, Olesen I, Steinshamn H. A comparative study of clinical manifestations, haematological and serological responses after experimental infection with *Anaplasma phagocytophilum* in two Norwegian sheep breeds. *Acta Vet Scand*. 2011;53:8.
- Stuen S, Okstad W, Sagen AM. Intrauterine transmission of *Anaplasma phagocytophilum* in persistently infected lambs. *Vet Sci*. 2018;5(1):25.
- Stuen S, Scharf W, Schauer S, Freyburger F, Bergström K, von Loewenich FD. Experimental infection in lambs with a red deer (*Cervus elaphus*) isolate of *Anaplasma phagocytophilum*. *J Wildl Dis*. 2010;46(3):803-9.
- Tabor AE. Anaplasmosis in ruminants. In: Line S, Moses MA, editors. *The Merck veterinary manual*. Kenilworth, NJ: Merck and Co; 2022. Available at: <https://www.merckvetmanual.com/circulatory-system/blood-parasites/anaplasmosis-in-ruminants>. Accessed 10 Jan 2024.
- Tate CM, Howerth EW, Mead DG, Dugan VG, Luttrell MP, Sahara AI, Munderloh UG, Davidson WR, Yabsley MJ. *Anaplasma odocoilei* sp. nov. (family Anaplasmataceae) from white-tailed deer (*Odocoileus virginianus*). *Ticks Tick Borne Dis*. 2013;4(1-2):110-9.
- The Ohio State University Health Sciences Center, College of Veterinary Medicine. Ehrlichial Research Laboratory. New taxonomy of the family Anaplasmataceae. Available at: <http://riki-lb1.vet.ohio-state.edu/ehrlichia/background/ehrlichiaspp.php>. Accessed 17 Aug 2004.
- Theodorou K, Leontides L, Siarkou VI, Petanides T, Tsafas K, Harrus S, Mylonakis ME. Synovial fluid cytology in experimental acute canine monocytic ehrlichiosis (*Ehrlichia canis*). *Vet Microbiol*. 2015;177(1-2):224-7.
- Thomas RJ, Birtles RJ, Radford AD, Woldehiwet Z. Recurrent bacteraemia in sheep infected persistently with *Anaplasma phagocytophilum*. *J Comp Pathol*. 2012;147(2-3):360-7.
- Thomas RJ, Dumler JS, Carlyon JA. Current management of human granulocytic anaplasmosis, human monocytic ehrlichiosis and *Ehrlichia ewingii* ehrlichiosis. *Expert Rev Anti Infect Ther*. 2009;7(6):709-22.
- Tinkler SH, Firshman AM, Sharkey LC. Premature parturition, edema, and ascites in an alpaca infected with *Anaplasma phagocytophilum*. *Can Vet J*. 2012;53(11):1199-202.
- Toom ML, Dobak TP, Broens EM, Valtolina C. Interstitial pneumonia and pulmonary hypertension associated with suspected ehrlichiosis in a dog. *Acta Vet Scand*. 2016;58(1):46.

- Topolovec J, Puntarić D, Antolović-Pozgain A, Vuković D, Topolovec Z, Milas J, Drusko-Barisić V, Venus M. Serologically detected "new" tick-borne zoonoses in eastern Croatia. *Croat Med J*. 2003;44(5):626-9.
- Townsend RL, Moritz ED, Fialkow LB, Berardi V, Stramer SL. Probable transfusion-transmission of *Anaplasma phagocytophilum* by leukoreduced platelets. *Transfusion*. 2014;54(11):2828-32.
- Unver A, Perez M, Orellana N, Huang H, Rikihisa Y. Molecular and antigenic comparison of *Ehrlichia canis* isolates from dogs, ticks, and a human in Venezuela. *J Clin Microbiol*. 2001;39:2788-93.
- Vanstreels RET, Yabsley MJ, Parsons NJ, Swanepoel L, Pistorius PA. A novel candidate species of *Anaplasma* that infects avian erythrocytes. *Parasit Vectors*. 2018;11(1):525.
- Varela AS. Tick-borne ehrlichiae and rickettsiae. In: Bowman DD. Companion and exotic animal parasitology. Ithaca, NY: International Veterinary Information Service [IVIS]; 2003. Available at: http://www.ivis.org/advances/parasit_Bowman/varela/chapter_frm.asp?LA=1. * Accessed 11 Aug 2004.
- Varela AS, Stallknecht DE, Yabsley MJ, Moore VA, Howerth EW, Davidson WR, Little SE. Primary and secondary infection with *Ehrlichia chaffeensis* in white-tailed deer (*Odocoileus virginianus*). *Vector Borne Zoonotic Dis*. 2005;5:48-57.
- Veronesi F, Passamonti F, Moretti A, Morganti G, Vardi DM, Laus F, Marenzoni ML, Spaterna A, Coletti M, Fioretti DP. Evaluation of the performance of a rapid enzyme-linked immunosorbent assay in the detection of *Anaplasma phagocytophilum* antibodies in horses. *Vector Borne Zoonotic Dis*. 2014;14(5):317-23.
- Víchová B, Majláthová V, Nováková M, Straka M, Pet'ko B. First molecular detection of *Anaplasma phagocytophilum* in European brown bear (*Ursus arctos*). *Vector Borne Zoonotic Dis*. 2010;10(5):543-5.
- Vieira RF, Biondo AW, Guimarães AM, Dos Santos AP, Dos Santos RP, Dutra LH, Diniz PP, de Moraes HA, Messick JB, Labruna MB, Vidotto O. Ehrlichiosis in Brazil. *Rev Bras Parasitol Vet*. 2011;20(1):1-12.
- Wallace JW, Nicholson WL, Perniciaro JL, Vaughn MF, Funkhouser S, Juliano JJ, Lee S, Kakumanu ML, Ponnusamy L, Apperson CS, Meshnick SR. Incident tick-borne infections in a cohort of North Carolina outdoor workers. *Vector Borne Zoonotic Dis*. 2016;16(5):302-8.
- Wang F, Yan M, Liu A, Chen T, Luo L, et al. The seroprevalence of *Anaplasma phagocytophilum* in global human populations: A systematic review and meta-analysis. *Transbound Emerg Dis*. 2020. doi: 10.1111/tbed.13548 [Online ahead of print].
- Wang J, Zhang Y, Cui Y, Yan Y, Wang X, Wang R, Jian F, Zhang L, Ning C. A rapid, simple and sensitive loop-mediated isothermal amplification method to detect *Anaplasma bovis* in sheep and goats samples. *Parasitol Int*. 2018;67(1):70-3.
- Wang J, Zhang Y, Wang X, Cui Y, Yan Y, Wang R, Jian F, Zhang L, Ning C. A loop-mediated isothermal amplification assay targeting 16S rRNA gene for rapid detection of *Anaplasma phagocytophilum* infection in sheep and goats. *J Parasitol*. 2017;103(2):187-92.
- Warner T, Harrus S. Canine monocytic ehrlichiosis. In: Recent advances in canine infectious diseases [monograph online]. Carmichael L, editor. Ithaca NY: International Veterinary Information Service [IVIS]; 2000. Available at: http://www.ivis.org/advances/Infect_Dis_Carmichael/toc.asp. * Accessed 19 Aug 2004.
- Wei R, Liu HB, Jongejan F, Jiang BG, Chang QC, Fu X, Jiang JF, Jia N, Cao WC. Cultivation of *Anaplasma ovis* in the HL-60 human promyelocytic leukemia cell line. *Emerg Microbes Infect*. 2017;6(9):e83.
- Williams CV, Van Steenhouse JL, Bradley JM, Hancock SI, Hegarty BC, Breitschwerdt EB. Naturally occurring *Ehrlichia chaffeensis* infection in two prosimian primate species: ring-tailed lemurs (*Lemur catta*) and ruffed lemurs (*Varecia variegata*). *Emerg Infect Dis*. 2002;8:1497-500.
- Woldehiwet Z. *Anaplasma phagocytophilum* in ruminants in Europe. *Ann N Y Acad Sci*. 2006;1078:446-60.
- Woldehiwet Z. The natural history of *Anaplasma phagocytophilum*. *Vet Parasitol*. 2010;167(2-4):108-22.
- Woldehiwet Z. Tickborne fever in ruminants. In: Line S, Moses MA, editors. The Merck veterinary manual. Kenilworth, NJ: Merck and Co; 2020. Available at: <https://www.merckvetmanual.com/generalized-conditions/tickborne-fever/tickborne-fever-in-ruminants>. Accessed 10 Jan 2024.
- Xu J, Gu XL, Jiang ZZ, Cao XQ, Wang R, Peng QM, Li ZM, Zhang L, Zhou CM, Qin XR, Yu XJ. Pathogenic *Rickettsia*, *Anaplasma*, and *Ehrlichia* in *Rhipicephalus microplus* ticks collected from cattle and laboratory hatched tick larvae. *PLoS Negl Trop Dis*. 2023;17(8):e0011546.
- Yabsley MJ. Natural history of *Ehrlichia chaffeensis*: Vertebrate hosts and tick vectors from the United States and evidence for endemic transmission in other countries. *Vet Parasit*. 2010. 167:136-48.
- Yabsley MJ, Adams DS, O'Connor TP, Chandrashekar R, Little SE. Experimental primary and secondary infections of domestic dogs with *Ehrlichia ewingii*. *Vet Microbiol*. 2011;150(3-4):315-21.
- Yabsley MJ, Loftis AD, Little SE. Natural and experimental infection of white-tailed deer (*Odocoileus virginianus*) from the United States with an *Ehrlichia* sp. closely related to *Ehrlichia ruminantium*. *J Wildl Dis*. 2008;44(2):381-7.
- Yancey CB, Diniz PPVP, Breitschwerdt EB, Hegarty BC, Wiesen C, Quorllo BA. Doxycycline treatment efficacy in dogs with naturally occurring *Anaplasma phagocytophilum* infection. *J Small Anim Pract*. 2018;59(5):286-93.
- Yang J, Liu Z, Niu Q, Liu J, Han R, Guan G, Hassan MA, Liu G, Luo J, Yin H. A novel zoonotic *Anaplasma* species is prevalent in small ruminants: potential public health implications. *Parasit Vectors*. 2017;10(1):264.
- Yang J, Liu Z, Niu Q, Tian Z, Liu J, Guan G, Liu G, Luo J, Wang X, Yin H. Tick-borne zoonotic pathogens in birds in Guangxi, Southwest China. *Parasit Vectors*. 2015;8:637.
- Ybañez AP, Inokuma H. *Anaplasma* species of veterinary importance in Japan. *Vet World*. 2016;9(11):1190-6.
- Yu DH, Li YH, Yoon JS, Lee JH, Lee MJ, Yu IJ, Chae JS, Park JH. *Ehrlichia chaffeensis* infection in dogs in South Korea. *Vector Borne Zoonotic Dis*. 2008;8(3):355-8.

Ehrlichiosis and Anaplasmosis

- Zhang J, Wang J, Kelly PJ, Zhang Y, Li M, et al. Experimental infection and co-infection with Chinese strains of *Ehrlichia canis* and *Babesia vogeli* in intact and splenectomized dogs: Insights on clinical, hematologic and treatment responses. *Vet Parasitol.* 2023;323:110032.
- Zhang XF, Zhang JZ, Long SW, Ruble RP, Yu XJ. Experimental *Ehrlichia chaffeensis* infection in beagles. *Med Microbiol.* 2003;52(Pt 11):1021-6.
- Zhang XC, Zhang LX, Li WH, Wang SW, Sun YL, Wang YY, Guan ZZ, Liu XJ, Yang YS, Zhang SG, Yu HL, Zhang LJ. Ehrlichiosis and zoonotic anaplasmosis in suburban areas of Beijing, China. *Vector Borne Zoonotic Dis.* 2012;12(11):932-7.
- Zhang Y, Lv Y, Cui Y, Wang J, Cao S, Jian F, Wang R, Zhang L, Ning C. First molecular evidence for the presence of *Anaplasma* DNA in milk from sheep and goats in China. *Parasitol Res.* 2016;115(7):2789-95.

*Link is defunct