S I d e 1	Rift Valley Fever	
S I d e 2	Overview • Organism • History • Epidemiology • Transmission • Disease in Humans • Disease in Animals • Prevention and Control	In today's presentation we will cover information regarding the organism that causes Rift Valley fever and its epidemiology. We will also talk about the history of the disease, how it is transmitted, species that it affects (including humans), and clinical and necropsy signs observed. Finally, we will address prevention and control measures for Rift Valley fever virus.
S I d e 3	THE ORGANISM	
S I d e 4	The Virus• Phlebovirus, Bunyaviridae• Stable at- 60°C to 23°C- 50 to 85% relative humidity• Inactivated by:• Lipid solvents• Detergents• Low pH	 Rift Valley fever (RVF) is caused by a <i>Phlebovirus</i> (Family Bunyaviridae). It is a three stranded RNA virus and requires a mosquito vector. The virus is very stable at temperatures from -60 to 23°C and at 50 to 85% relative humidity. It is inactivated by lipid solvents, detergents and low pH. Photo: Electron micrograph of the Rift Valley fever virus (RVFV), CDC.
S I d 5	 Disease Overview Acute febrile disease Sheep, cattle, goats High abortion rates and death in young Zoonotic Heavy rainfalls Arthropod vector Most commonly mosquito OIE Listed disease 	Rift Valley fever is an acute febrile disease that severely affects sheep, cattle and goats, especially young animals. There is a very high rate of abortion and death in neonates. RVF can also affect humans. Typically cases have mild, flu-like signs that are self-limiting, but severe disease can occur. Rift Valley fever is associated with heavy rainfalls and is transmitted by an arthropod-vector, usually mosquitoes. However, contact with contaminated tissue from infected animals, especially aborted fetuses, can transmit the virus to humans. Currently, RVF is an OIE-listed disease.

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– 1950-51: Kenya

– 1977-78: Egypt

500,000 sheep abortions100,000 sheep deaths

18,000 human cases
598 human deaths

Rift Valley fever was first recognized as a disease in sheep in Rift Valley, Kenya around 1900. The virus was not isolated until 1930. Since then there have been intermittent outbreaks throughout Kenya. A major epizootic occurred in 1950-51 which resulted in 500,000 sheep abortions and 100,000 sheep deaths. In 1977-78, a outbreak of RVF occurred along the Nile in Egypt. This was the first epidemic out of sub-Saharan Africa. This outbreak resulted in an estimated 18,000 human cases. There were 598 human deaths which occurred from encephalitis and hemorrhagic fever. Many abortions and deaths were reported in sheep, goats, cattle, water buffalo and camels.

Image: This map depicts the locations of Kenya and Egypt. Map from www.worldatlas.com.

In 1987, an outbreak of RVF occurred in the Senegal river basin (West Africa). This epizootic differed from prior outbreaks in that it was not associated with the typical heavy rainfall. Mosquito breeding occurred in large rivers and dams. The largest outbreak of RVF reported occurred in Kenya, Africa in 1997-1998. An estimated 89,000 humans were infected with 478 deaths. An RVF outbreak occurred in Saudi Arabia from August 2000 to September 2001; it was the first confirmed outbreak of RVF outside of Africa. This outbreak illustrated the potential for the spread of this disease to other regions of the world. An outbreak of Rift Valley fever occurred in Egypt in August of 2003. Forty-five cases of RFV were diagnosed with 17 deaths. The outbreak occurred in a rural region 150 kilometers north of Cairo and all cases were Egyptian farmers.



In 2006, an outbreak of Rift Valley fever began in Kenya. It soon spread to the surrounding countries of United Republic of Tanzania and Somalia. Over 1000 human case were reported with case-fatality varying between the countries from 23 to 45%. Animal movement and slaughter restrictions were implemented as has vaccination efforts for livestock in affected areas. In 2010, South Africa had over 14,000 cases of RVF; 489 separate outbreaks were observed. Abnormally high rainfall likely contributed to these outbreaks.

Important Outbreaks • 1987: Senegal – Not associated with rainfall • 1997-98: Kenya – Largest outbreak reported – 89,000 humans cases - 478 deaths • 2000-01: Saudi Arabia and Yemen – First outbreak outside of Africa

2003: Egypt

 45 cases; 17 deaths
 All cases were Egyptian farmers



S I d e 1 5	Transmission • Secondary arthropod vectors - Rapidly spread disease - <i>Culex</i> and <i>Anopheles</i> mosquito species - Biting flies • Midges, phlebotomids, stomoxids, simulids	Secondary arthropod vectors can become infected from the ruminant and rapidly spread the disease. <i>Culex</i> and <i>Anopheles</i> mosquitoes can serve as secondary vectors. Several of these species of mosquitoes are found in the United States and could potentially contribute to the spread of RVF in this country. Biting flies such as midges, phlebotomids, stomoxids and simulids may serve as mechanical transmitters of the virus, however this is thought to be minimal route of infection.
	Center far Food Security and Public Health, lows State University, 2011	<u> </u>
S	Additional Modes	Secondary arthropod vectors can become infected from the ruminant and rapidly spread the disease. <i>Cular</i> and <i>Anopheles</i> mosquitoes can serve as
i	• Secondary arthropod vectors	secondary vectors. Biting flies such as midges, phlebotomids, stomoxids
d	 Rapidly spread disease Culex and Anonheles mosquito species 	and simulids may serve as mechanical transmitters of the virus, however
е	 Biting flies (mechanical) Midges, phlebotomids, stomoxids, simulids 	this is thought to be minimal route of infection. The virus can also be transmitted <i>in utero</i> to the fetus. It has also been found in semen and raw
1	Animals In utero	milk.
6	Semen, raw milk	
	Certier for Food Security and Public Health, tons Statis University, 2011	
S	Additional Modes	Humans do not seem to be infected by casual contact with live hosts, but
I i	of Transmission	parturition, necropsy, slaughter, laboratory procedures or meat
d	- Direct contact or aerosol	preparation for cooking. In utero transmission to a human infant was
e	 Tissue or body fluids of infected animals Aborted fetuses, slaughter, necropsy In utero 	first reported in 2006.
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	Carrier for Food Security and Public Health, Iown State University, 2011	
S	TRANSMISSION BOUTSS OF RH Valley Fover	This diagram depicts the various transmission routes of Rift Valley
 ;	And the process of th	Graphic illustration by Clint May, Center for Food Security and Public
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d	DISEASE IN HUMANS	
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S I d e 2 0	Human Disease • Incubation period: 2 to 6 days • Often asymptomatic • Often asymptomatic • Influenza-like illness • Pareu • Recorey in 2 to 7 days • Retinopath • Retinopath • Renorrhagic fever • Bracephalitis	The incubation time in humans has been reported to be anywhere from 2 to 12 days. The average is 2 to 6 days. Typically humans are asymptomatic or have self-limiting flu-like signs. These signs include fever, headache, muscle and joint pain, and possible nausea and vomiting. Recovery is usually in 2 to 7 days. In less than 1% of humans infected, severe disease can occur. This can include retinitis, hemorrhagic fever or encephalitis.
S I d e 2 1	 Human Disease Netinopathy 1 to 3 weeks after onset of symptoms Conjunctivitis Photophobia Can lead to permanent vision loss Death is uncommon 	Of the 1% of cases developing into severe syndromes, 1 to 10% of these cases will develop retinopathy. Clinical signs seen include conjunctivitis and photophobia. Damage from this complication can lead to permanent vision loss. Death from this syndrome is uncommon.
S I d e 2 2	Human Disease • Hemorrhagic fever - 2 to 4 days after fever - Melena, hematemesis, petechia, jaundice, shock, coma - Death - Case-fatality is ~50% • Encephalitis - 1 to 3 weeks after onset of symptoms - Can occur with hemorrhagic fever	Other possible complications from RVF include hemorrhagic fever and encephalitis. People demonstrating hemorrhagic fever syndrome will have fever, melena, hematemesis, petechia and subcutaneous bleeding. Additionally, deep jaundice is present. The syndrome quickly progresses to shock and coma followed by death. The case-fatality rate for persons with this syndrome is approximately 50%. The encephalitic syndrome typically occurs 1 to 3 weeks after the onset of symptoms. It can occur concurrently with the hemorrhagic fever. Both of these syndromes occur in only about 1% of human cases.
S I d e 2 3	<section-header><section-header><section-header><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></section-header></section-header></section-header>	Viral antigens and RNA can be detected in blood and tissue samples by various antigen detection tests and reverse transcription polymerase chain reaction (RT-PCR) assays. Enzyme-linked immunoassay (ELISA) and other serologic tests can detect specific IgM or rising titers. Treatment is symptomatic and supportive therapy. With the hemorrhagic syndrome, blood transfusions may be needed to replace coagulation factors. The anti-viral medication, ribavirin, may prove helpful. Photo: Cataloging samples for testing.

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S I d e 3 0	<section-header><list-item><list-item><list-item><list-item><list-item><list-item><list-item><list-item> <section-header></section-header></list-item></list-item></list-item></list-item></list-item></list-item></list-item></list-item></section-header>	Post mortem lesions found in sheep, cattle and goats indicate hepatic necrosis. This can be quite extensive in younger animals and fetuses. The liver is greatly enlarged, yellow and friable. Petechial hemorrhages may be very prominent and found on cutaneous or serosal surfaces. [Photo shows a sheep liver that is pale, swollen and contains multiple foci of hemorrhage. Photo from Armed Forces Institute of pathology (AFIP)]
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u e	PREVENTION AND	
C	CONTROL	
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S		State and federal veterinarians should be notified immediately of any
I	Recommended Actions	suspicious cases of RVF. While waiting for the authorities or a
i	Notification of Authorities	confirmed diagnosis, all suspect animals should be quarantined.
d	- Federal Area Veterinarian in Charge	
е	http://www.aphis.usda.gov/animal_health/area offices/	Photo from Dr. Katie Steneroden, ISU.
	- State Veterinarians	
3	• Quarantine	
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	Center for Food Security and Public Health, loss State University, 2011	
S		Vaccines are generally used to protect animals from Rift Valley fever in
Ι	Prevention	endemic regions. During epidemics, vaccination of susceptible animals
i	Vaccination Common in endemic areas	can prevent amplification of the virus and protect people as well as
d	- May be used in outbreaks	animals. Attenuated and inactivated Rift Valley fever vaccines are both
е	Attenuated Attenuated Attenuated Attenuated Attenuated Better immunity but may cause abortion and birth	abortions and birth defects can occur in pregnant animals. Subunit
2	efects in pregnant animals • Vector control	vaccines are in development. Additional, less commonly used,
3	Keep livestock at high altitudes Animal housing controls	preventative measures include vector controls, movement of stock to
5		higher altitudes, and the confinement of stock in insect-proof stables.
	Learn for root Security and Public Health, loss State University, 2011	These control methods are often impractical, or are ineffective because they are instituted too late. The measurement of animals from an demis
		areas to RVF-free regions can result in enidemics
S		Mosquito repellents, long shirts and trousers, bednets, and other
Ī	Prevention	arthropod control measures should be used to pre-vent transmission by
i	Vector control	mosquitoes and other potential insect vectors. Outdoor activities should
d	 Mosquito repellents, long shirts and pants, bed nets 	be avoided, if possible, during periods of peak mosquito activity.
е	Barrier precautions	insecticides may be neipiul. During epidemics, vaccination of susceptible animals can prevent amplification of the virus and protect
~	 During exposure to infectious tissues or blood 	people as well as animals. Barrier precautions should be used whenever
3	 Wear personal protective equipment Human vaccine 	contact may occur with infectious tissues or blood from animals:
4	- Limited availability	recommended measures include personal protective equip-ment such as
	Certier for Food Seconty and Public Health, Iona State University, 2011	protective clothing, gloves and goggles. Diagnostic tissue samples

should be processed by trained staff in appropriately equipped laboratories. Universal precautions are recommended for healthcare workers who care for patients with confirmed or suspected Rift Valley fever. Barrier techniques are recommended when nursing hospitalized patients. A human vaccine has been developed, but has limited availability. Additional vaccines are under investigation.

S I d e 3 5	 Cleaning and Disinfection Clean animal housing areas Wear personal protective equipment Remove all organic material from surface (manure, feed, animal tissue) Use soap or detergent with warm water Le dry Disinfect animal housing areas part bleach:10 parts water Virkon-S[®] 	The RVF virus is easily destroyed by disinfectants. However, some disinfectants are easily inactivated by organic material (manure, feed, animal tissues, etc.). Therefore, cleaning is the first important step when treating an area that has become contaminated with RVF virus. When cleaning and disinfecting, it is important to wear personal protective equipment (gloves, coveralls, boots, protective eyewear and a respirator) since the virus may become airborne. Cleaning begins by removing all organic material from the surface. Next, use soap or a detergent with warm water and let the surface dry; then, apply the disinfectant. One part block diluted with 10 meters water on wing the product Virlen S@ by
ļ		DuPont will destroy the RVF virus.
S I d e 3 6	Rift Valley Fever as a Biological Weapon • Aerosol or droplets • 1 km downwind • 35,000 humans incapacitated • 400 deaths (1% mortality) • Human introduction • Animals as sentinels	The use of the RVF virus as a bioterrorism agent would most likely be via virus-laden aerosol or droplets. It has been hypothesized that if RVF virus were disseminated by an airplane, 50 kg of the agent would have a 1 km downwind reach with 35,000 humans incapacitated and 400 deaths (1% mortality). It may be possible for a human to introduce RVF as well. A unsuspecting traveler from Africa may be incubating the disease. Upon arriving in the U.S., if he is bitten by a mosquito, the mosquito then becomes infective. The virus can then be transmitted to an animal which serves as an amplifying host. Currently the U.S. has several
		animals typically occurs before that in humans, animals may serve as sentinel species for this disease.
S	Additional Resources	
i	World Organization for Animal Health	
d	(OIE) – www.oie.int	
e	 U.S. Department of Agriculture (USDA) www.aphis.usda.gov Center for Food Security and Public Health www.cfsph.iastate.edu 	
3	USAHA Foreign Animal Diseases ("The Gray Book")	
7	se/downloads/nahems/fad.pdf	
5 	Acknowledgments	
i	Development of this presentation was funded by grants from	
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e	Management Division, and the Iowa Department of Agriculture and Land Stewardship to the Center for Food Security and Public Health at Iowa State University.	

, DVM, MPH, DACVPM; Ra

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