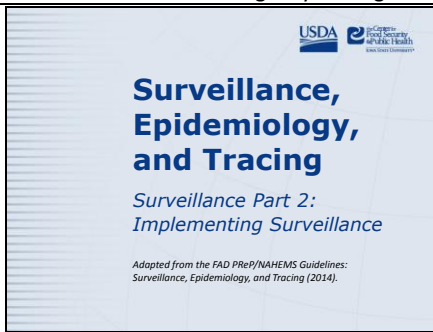
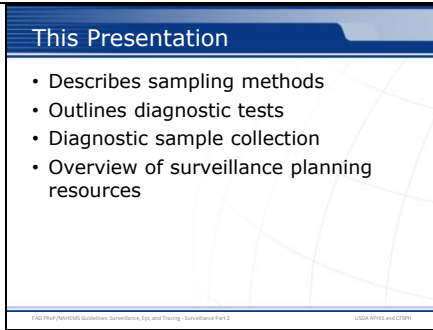


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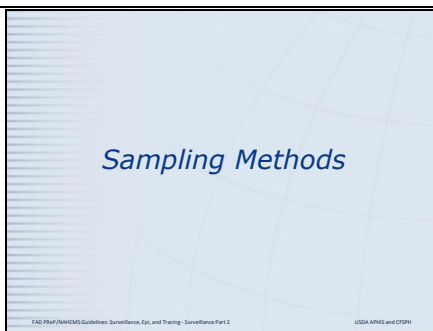
An animal health emergency could have a detrimental effect on the nation's agriculture, food supply, and economy. Veterinary responders, animal health technicians, and other trained personnel may assist with surveillance, epidemiology, and tracing activities. In order to perform these job duties, a broad understanding of surveillance and epidemiological concepts is required. This presentation is Part 2 in describing the process of implementing surveillance. [This information was derived from the *Foreign Animal Disease Preparedness and Response (FAD PReP)/National Animal Health Emergency Management System (NAHEMS) Guidelines: Surveillance, Epidemiology, and Tracing (2014).*]

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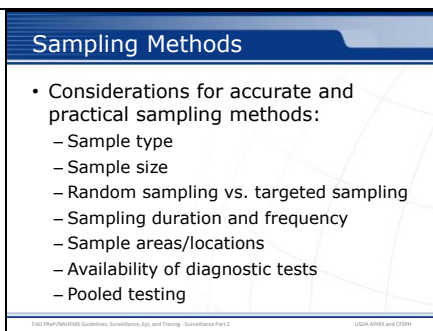
This presentation provides more information about the development of a surveillance plan implemented in a foreign animal disease (FAD) outbreak. It describes the different sampling methods, how those samples are collected, and the different diagnostic tests that are run. It also identifies different resources that may be helpful in surveillance planning.

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This presentation will focus on some considerations for implementing some of the operational aspects of surveillance activities. It will discuss sampling methods, diagnostic tests, and sample collection, as examples that need to be considered. Sampling methods for a surveillance system must be described in detail for a foreign animal disease (FAD) outbreak. Data collection methods must be determined and statistical concerns must also be addressed. This section describes considerations for developing accurate and practical sampling methods.

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Considerations for developing accurate and practical sampling methods include sample type; sample size; random sampling vs. targeted sampling; sampling duration and frequency; sample areas/locations; availability of diagnostic tests; and pooled testing.

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Sample Type and Size

- Sample type
 - Disease agent
 - Available tests
 - Laboratory capabilities
- Sample size
 - Population size
 - Disease prevalence
 - Diagnostic test sensitivity
 - Confidence level



The type of sample collected depends on the FAD agent, available tests, laboratory capabilities/preferences, and resources. Information can be collected via surveys, questionnaires, visual inspections, and collection of diagnostic specimens (ante or post mortem). It is rarely feasible to test all susceptible animals in an outbreak. In most cases, a subset of the herd or group is selected.

The size of the group sampled is affected by:

- Population size;
- Disease prevalence;
- Diagnostic test sensitivity (the likelihood that a test will accurately identify infected animals); and
- Confidence level (the degree of certainty that test results reflect true disease status in the animal population). Generally speaking, the larger the sample selected, the greater the confidence that can be placed in the results.

[The top photo is a photo of properly labeled sample tubes ready for use in collecting samples. The bottom photo illustrates a population of susceptible animals from which a subset will be sampled. Top photo source: Alex Ramirez, Iowa State University. Bottom photo source: Iowa State University, VDPAM]

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Random vs. Targeted

- Random sampling
 - Every animal in the targeted population has an equal chance of getting selected for testing
- Targeted sampling
 - May choose animals for convenience or because a certain group has a specific risk factor or higher prevalence of disease

Random sampling assures every animal in the targeted population has an equal chance of getting selected for testing. Targeted sampling may choose animals for convenience or because a certain group has a specific risk factor or higher prevalence of disease. On the following slides we will further look into these two methods of sampling and examples to better help explain these concepts.

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Comparison

<p>Random sampling</p> <ul style="list-style-type: none"> • Every animal has an equal chance of being selected for testing • Preferred over non-random sampling for determining prevalence or incidence of disease <ul style="list-style-type: none"> – Results are better extrapolated 	<p>Targeted sampling</p> <ul style="list-style-type: none"> • Animals not chosen randomly • Usually preferred during a disease outbreak <ul style="list-style-type: none"> – Primary objective is to identify cases of disease • Cost effective and increases likelihood of finding new cases
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Random Sampling assures that every animal in the target population has an equal chance of being selected for testing. Random sampling is generally preferred over non-random sampling for determining the prevalence or incidence of disease because results can be better extrapolated to the population at risk. Example: Assume that 50 dairy cows per day are examined (sampled) to determine the incidence of lameness in a herd. Examining every *n*th cow in the milk string would provide a sample that represents the whole group.

Targeted (non-random) Sampling may choose animals for convenience or because a certain group has a specific risk factor or higher prevalence of disease. Non-random samples are often preferred during a disease outbreak because the primary objective of surveillance is to identify cases of the disease. This method is cost effective and increases the likelihood of finding new cases. Example: Using the previous example, targeted or non-random sampling of the herd may test the last 50 cows in the milk string because cows with lameness are more likely to be at the end of the string.

Sampling Frequency

- Surveillance
 - Begins as soon as possible in an FAD outbreak
- Susceptible animals
 - Sampled a minimum of three times during the maximum incubation period

Maximum Incubation Period

The longest period that elapses between the introduction of the FAD agent into a susceptible animal and the occurrence of the first clinical signs compatible with the FAD agent.

FAD PReP/NAHEMS Guidelines: Surveillance, Epi, and Tracing – Surveillance Part 2 USDA APHIS and OIE/WHO

Surveillance should begin as soon as possible in an FAD response. In general, it is necessary to inspect/sample susceptible animals a minimum of three times during the maximum incubation period for the disease under investigation. The **maximum incubation period** is defined as the longest period that elapses between the introduction of the FAD agent into a susceptible animal and the occurrence of the first clinical signs compatible with the FAD agent. *[The text in this illustration provides the definition of Maximum Incubation Period. Illustration by: Bridget Wedemeier, Iowa State University]*

Determining Frequency

- Latent period
 - Period of time between host infection and ability to infect others
- Incubation Period
 - Period of time between infection and development of clinical signs
- Infectious period
 - Period of time that an infected animal can transmit the pathogen to another susceptible animal
- Rapidity of disease transmission
- Likelihood of disease spread

FAD PReP/NAHEMS Guidelines: Surveillance, Epi, and Tracing – Surveillance Part 2 USDA APHIS and OIE/WHO

In most cases, susceptible animals will be placed under surveillance for at least two maximum incubation periods. Sampling frequency for animals, herds or premises is based on a number of factors including:

- Latent period: period of time between host infection and the ability to infect others
- Incubation period: period of time between infection and the development of clinical signs
- Infectious period: period of time that an infected animal can transmit the pathogen to another susceptible animal
- Rapidity of disease transmission between animals and premises
- Likelihood of disease spread

NOTE: Repeated tests are often necessary. When repeated tests are conducted, a previous negative test result can strengthen a subsequent negative test result when the interval between tests is short. Two negative test results occurring within days of each other are more reliable than two negative test results occurring within weeks of each other.

Sampling Areas/Locations

- Target population may be selected based on area or location

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A target population may be selected based on area or location. The nature of the FAD agent and its ability to spread long distances through aerosol transmission may place livestock miles away from the infected premises under surveillance. The process of premises/zone classification is very important for determining surveillance needs by location. Specific surveillance activities to locate new cases may be important throughout all zones, but may be focused on the Control Area (Infected Zone and Buffer Zone). Gathering information to demonstrate freedom from infection, using a different set of protocols, may focus on the Free Area. *[These maps visually illustrate disease Zone, Area, and Premises designations. Content provided by: USDA. Illustration by: Dani Ausen, Iowa State University]*

Other Considerations

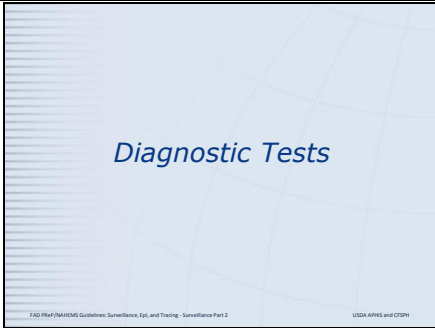
- Diagnostic test availability
 - Validated and approved
 - Affected by manufacturer capacity, reagent availability, etc.
- Pooled testing
 - Samples from multiple animals are combined into a single test
 - Cost effective and useful when resources are limited
 - May not be appropriate for all samples

FAD PReP/NAHEMS Guidelines: Surveillance, Epi, and Tracing – Surveillance Part 2 USDA APHIS and OIE/WHO

Other factors to consider when choosing sampling methods include the following. The chosen sampling methods for an FAD outbreak will consider the laboratory tests that are validated, approved, and available for the disease agent (**diagnostic test availability**). Test availability may be affected by manufacturer capacity, reagent availability, etc. **Pooled testing** is a testing method where samples from multiple animals are combined into a single test. This testing method is cost effective and useful when time and resources are limited. Pooled testing may not be appropriate for all sample types or pathogens and, in some cases, may increase the likelihood of false negative results. The availability of a validated and approved diagnostic test for the FAD agent may also affect sample choice. Additional information on diagnostic tests is found later in this presentation.

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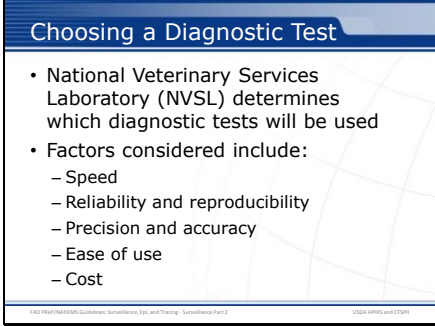
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Surveillance planning includes consideration of appropriate diagnostic tests. The next section discusses choosing a diagnostic test, as well as sensitivity and specificity, and laboratory capacity.

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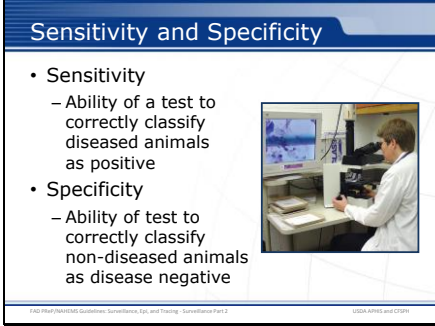
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The National Veterinary Services Laboratories (NVSL) will determine which diagnostic tests will be used in an FAD outbreak. When choosing a diagnostic test, NVSL will consider factors such as: speed, reliability, reproducibility, precision, accuracy, ease of use, and cost for each available diagnostic test.

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Sensitivity and specificity are important criteria to consider when choosing a diagnostic test. A variety of diagnostic tests may be available for a given disease. Each diagnostic test has a different ability to correctly identify diseased and non-diseased animals. Sensitivity is the ability of a test to correctly classify a percentage of diseased animals as positive. For example, if the test sensitivity is 95 percent, 95 out of 100 sick animals will be detected. Specificity is the ability of a test to correctly classify non-diseased animals as disease negative. *[This photo shows a sample being examined microscopically. Photo source: Danelle Bickett-Weddle, Iowa State University]*

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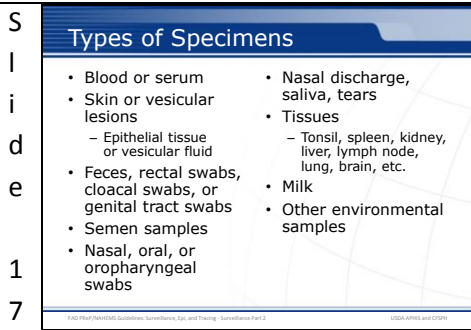
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As previously mentioned, multiple factors will affect diagnostic test availability. Laboratory capacity, or the ability of a laboratory to complete necessary FAD testing, may also be a limiting factor. In an FAD response, samples will be sent to the Foreign Animal Disease Diagnostic Laboratory (FADDL) in Plum Island, NY or the NVSL in Ames, IA depending on the disease and test to be performed. Alternatively, the National Animal Health Laboratory Network (NAHLN), a network of State, university, and other approved laboratories, maintains the capacity and capability to provide laboratory services in support of FAD outbreaks. For FAD investigations, if only one sample is collected, the sample will be sent to NVSL (either FADDL or NVSL Ames). If there are two samples collected, the first sample will be sent to NVSL (either FADDL or NVSL Ames), and the second sample can be sent to a NAHLN laboratory. In all cases, confirmation of FADs is performed by NVSL. For FAD surveillance, NAHLN laboratories will be involved based on their capacity and available assays. For more information on sample collection and testing, please see the *FAD Investigation Manual (FAD PRoP Manual 4-0)*.



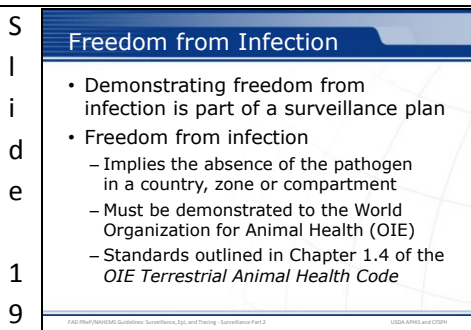
The type of specimen collected is determined by the disease of concern, available diagnostic tests, and the ability to obtain samples from target species. The number and type of specimens to collect will be communicated by Incident Command (IC).



Diagnostic specimens that may be collected include those listed on the slide. Specimens may include animal tissues or fluids, animal products like milk, or environmental samples. For in-depth information on collecting diagnostic specimens, please see the *FAD Investigation Manual (FAD PReP Manual 4-0)*.



Collecting Specimens: Collect specimens according to the protocol and in a manner that prevents cross-contamination and sample degradation. Clearly and legibly label individual specimen tubes/containers with permanent, waterproof ink in a manner that allows identification of the specimen (animal, location, date, type, etc.)
Packaging specimens for shipment: Another factor that can impact sample quality is transportation and shipping of specimens. The diagnostic lab, shipping company and the U.S. Department of Transportation establish packaging and shipping requirements which must be followed. For further information see: (http://www.aphis.usda.gov/wps/portal/aphis/ourfocus/animalhealth?1dmy&urile=wc%3apath%3a%2Faphis_content_library%2Fsa_our_focus%2Fsa_animal_health%2Fsa_lab_information_services%2Fsa_diagnostic_tests%2Fct_packaging_labeling)
Biosecurity protocols: Lastly, adhere to the biosecurity protocols established for sample collection, packaging and shipment in order to prevent cross contamination of samples and the further spread of disease agents. [This photo illustrates packaging and shipping materials. Photo source: Pam Zaabel, Iowa State University]



The surveillance plan must include a plan for demonstrating freedom from infection. Freedom from infection implies the absence of the pathogenic agent in a specified animal population, or population in a country, zone, or compartment. As a member of the World Organization for Animal Health (OIE), the United States makes every effort to demonstrate freedom from infection to a level of confidence acceptable under OIE standards. These standards are outlined in Chapter 1.4 of the *OIE Terrestrial Animal Health Code (2011)*. An example proof of disease freedom surveillance scheme for HPAI is provided in the *HPAI Surveillance SOP*.

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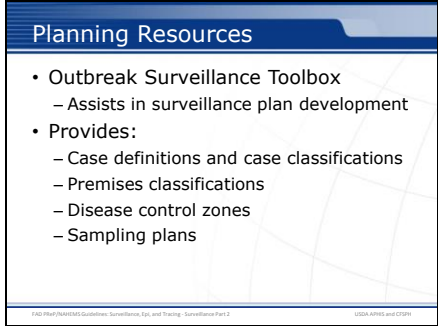
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There are a variety of resources that may be helpful during surveillance planning.

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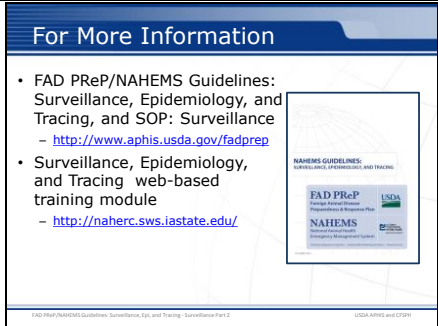


The Outbreak Surveillance Toolbox, available on the APHIS Intranet or on CD, is designed to assist in developing a surveillance plan. The Toolbox provides information and resources to establish:

- Case definitions
- Case classifications
- Premises classifications (Same as found in APHIS Foreign Animal Disease Framework: Response Strategies (FAD PRoP Manual 2-0))
- Disease control zones (Same as found in APHIS Foreign Animal Disease Framework: Response Strategies (FAD PRoP Manual 2-0))
- Sampling plans within each Control Area (or Free Area) to detect disease within individual herds and “prove” disease freedom

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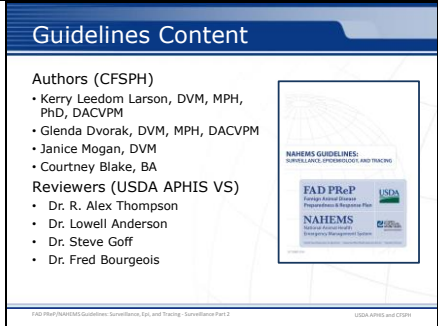
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More details can be obtained from the sources listed on the slide, available on the USDA website (<http://www.aphis.usda.gov/fadprep>) and the NAHERC Training Site (<http://naherc.sws.iastate.edu/>).

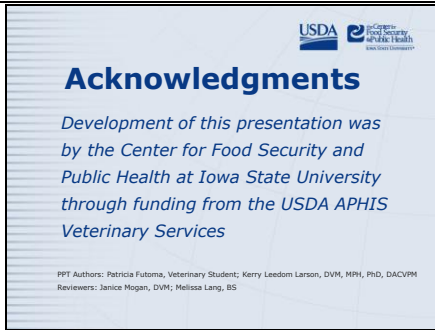
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The print version of the Guidelines document is an excellent source for more detailed information. In particular, the Guidelines document has listings of additional resources. This slide acknowledges the authors and reviewers of the Guidelines document. It can be accessed at <http://www.aphis.usda.gov/fadprep>.

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